

Practical Dosimetry of Peptide Receptor Radionuclide Therapy with ^{90}Y -Labeled Somatostatin Analogs

Stanislas Pauwels, MD, PhD¹; Raffaella Barone, MD¹; Stéphan Walrand, PhD¹; Françoise Borson-Chazot, MD, PhD¹; Roelf Valkema, MD, PhD²; Larry K. Kvols, MD³; Eric P. Krenning, MD, PhD²; and François Jamar, MD, PhD¹

¹Centre de Médecine Nucléaire, Université Catholique de Louvain, Brussels, Belgium; ²Department of Nuclear Medicine, Erasmus Medical Center, Rotterdam, The Netherlands; and ³Lee Moffitt Cancer Center, University of South Florida, Tampa, Florida

The challenge for internal therapy is to deliver the highest possible dose to the tumor while sparing normal organs from damage. Currently, the potential risk of kidney and red marrow toxicity limits the amount of radioactivity that may be administered. An accurate dosimetry method that would provide reliable dose estimates to these critical organs and to tumors before therapy would allow the clinician to plan a specific therapeutic regimen and also select those patients who would benefit the most from treatment. The dosimetry for ^{90}Y -1,4,7,10-tetraazacyclododecane-*N,N',N'',N'''*-tetraacetic acid-D-Phe¹-Tyr³-octreotide is usually based on quantitative imaging at different time points that provides information on activity retention in organs over time and on stylized models representing average individuals. Because the therapeutic agent labeled with ^{90}Y is not suitable for quantitative imaging, the peptide surrogate labeled with the positron emitter ^{86}Y can be considered the most appropriate tracer for measuring distribution and retention of the radiopharmaceutical over time. Dose calculations in target organs are generally performed using the MIRDose program, in which S values from source to target are integrated. Significant improvement of dose estimates may be achieved by introducing patient-specific adjustments to the standard models. The use of individual kidney volumes assessed by CT instead of the use of a fixed volume for males and females may significantly improve the determination of kidney radiation doses. The use of actual CT-derived tumor volumes has also shown a dose–efficacy relationship. Additional improvements in this field include the validation and use of an ^{111}In surrogate to avoid the complexity of ^{86}Y use and the consideration of radiobiologic parameters, such as fractionation effects and the specific biologic efficacy of internally deposited radiation, which are probably underestimated using currently available methods.

Key Words: dosimetry; ^{90}Y -DOTA-Tyr³-octreotide; radionuclide therapy; oncology

J Nucl Med 2005; 46:92S–98S

The recent availability of radiolabeled somatostatin analogs opened a new avenue for therapy of several tumor types bearing a high density of somatostatin receptors (SSR). Until now, the attention has been mainly focused on tumors expressing SSR2, such as neuroendocrine tumors. In the near future, other radiolabeled analogs binding to other receptor subtypes or to other hormone receptors will become available (1).

The challenge of this type of therapy is to deliver the highest possible radiation-absorbed dose to the tumor(s) while sparing normal organs from radiation damage. In the case of neuroendocrine tumors, clinical trials have been conducted with several compounds, including ^{90}Y -[DOTA]-D-Phe¹-Tyr³-octreotide (^{90}Y -DOTATOC, OctreoTher) and ^{111}In -diethylenetriaminepentaacetic acid (DTPA)-D-Phe¹-octreotide (^{111}In -DTPA-OC; OctreoScan111, or SomatoTher). Satisfactory therapeutic results were noted with objective tumor responses or stabilization in patients with progressive disease. However, significant toxicity was also noted. Acute toxicity remained limited, but delayed toxicity was observed, leading to substantial morbidity. In the case of administration of very high activities of ^{111}In -DTPA-OC, bone marrow toxicities in the form of myelodysplastic syndromes and leukemia were reported (2). With ^{90}Y -DOTATOC, reported toxicity was mainly renal, with a significant impairment of renal function and some patients evolving to end-stage renal failure and hemodialysis (3–5). This was particularly true in the phase of early human use, when no attempts at dosimetry were made. Infusion of large amounts of amino acids was proposed to reduce the radiation dose to kidneys by a factor of 20%–40% (6). With dose escalation, however, bone marrow irradiation levels may become critical and also may lead to delayed myelotoxicity.

An accurate knowledge of actual radiation doses to tumor tissues as well as to normal organs would be of great help in planning for therapy. Such knowledge might also aid in predicting which patients might better benefit from the treatment and in identifying those with unfavorable dosim-

Received May 19, 2004; revision accepted Sept. 13, 2004.
For correspondence or reprints contact: Stanislas Pauwels, MD, PhD, Centre de Médecine Nucléaire, Université Catholique de Louvain, UCL 54.30, Avenue Hippocrate, 54, B-1200 Brussels, Belgium.
E-mail: stanislas.pauwels@mnuc.ucl.ac.be

etry. To be optimal, such estimations require time-consuming and sophisticated methods, including pharmacokinetic, biodistribution, and washout studies using the pharmaceutical and the radionuclide to be used for therapy. This may be impossible for practical reasons related to the patient's status and for physical reasons. In the case of ^{90}Y -DOTATOC, the β -emission of ^{90}Y does not allow appropriate quantification from the bremsstrahlung emission. Alternative approaches are therefore envisioned, including imaging with similar analogs or using the same analog labeled with a positron emitter used as a surrogate for the therapeutic radionuclide (e.g., ^{86}Y).

This article reviews experience by several investigators in the estimation of radiation doses to tissues and tumors during therapy with ^{90}Y -labeled somatostatin analogs. Methodologic aspects are emphasized, such as the choice of the radiopharmaceutical, acquisition and processing of data, and dosimetry calculations, usually based on the MIRD formula. Also included is an overview of results reported on radiation doses to kidney, bone marrow, and tumors, as well as to other potential target organs.

CHOICE OF RADIOPHARMACEUTICAL

Because ^{90}Y cannot be imaged accurately to ensure correct quantification of organ activity, different approaches are used to mimic the biodistribution of ^{90}Y -DOTATOC and predict its dosimetric parameters.

The chemical structure (hydrophilicity of the molecule and charge) of the radiolabeled analog is important, because it influences tumor uptake, urinary clearance, renal retention, and residence times in tissues (7,8). The ideal method for mimicking ^{90}Y -DOTATOC is to use the same compound labeled with the positron-emitter ^{86}Y . In this case, the chemical structure of the compound is totally preserved. In 24 patients who were candidates for therapy with ^{90}Y -DOTATOC, we studied the pharmacokinetics and biodistribution of ^{86}Y -DOTATOC (9). Using the same methodology, Förster et al. (10) reported data on 3 patients. However, this application is limited, because it requires a PET facility and a high-energy cyclotron. Moreover, the short half-life of ^{86}Y (14.3 h) does not enable delayed acquisitions beyond 48–72 h after tracer administration. To overcome this problem, biodistribution studies were performed using the commercially available ^{111}In -DTPA-OC to predict the behavior of ^{90}Y -DOTATOC. Results were contradictory. Förster et al. (10) showed an overestimation of renal radiation doses using ^{111}In -DTPA-OC when compared with ^{86}Y -DOTATOC, but they observed a good correlation between both radiopharmaceuticals in individual patients. In our experience, ^{111}In -DTPA-OC clearly underestimates ^{90}Y -DOTATOC renal dose compared with ^{86}Y -DOTATOC, and we found a weak correlation between dose estimates for ^{90}Y -DOTATOC with both tracers as a result of inpatient variation (11). The differences between the 2 studies were probably related to

the acquisition method for ^{111}In -DTPA-OC (SPECT vs. planar) and to the use of amino acid infusion.

The last approach used to mimic ^{90}Y -DOTATOC was reported by Cremonesi et al. (12), who used DOTATOC labeled with ^{111}In . This radiopharmaceutical allows γ -camera imaging, which is accessible everywhere, although the chemical behavior of the analog labeled with ^{111}In is not identical to that of the analog labeled with ^{90}Y .

DOSE CALCULATION METHODOLOGY

In internal radiotherapy the dose (D_t) to a target tissue (t) is defined as:

$$D_t = \sum_s S_{t \leftarrow s} N_s$$

where N_s = number of decays occurring in the source tissue s , and S factor $S_{t \leftarrow s}$ = energy transported per decay from the source tissue s to a unity mass of the target tissue t .

Thus, patient-specific dosimetry requires quantitative patient imaging at different time points to evaluate the retention of activity in various organs over time. After integration of the measured activities, these time integrals of activity are multiplied by energy transport factors to obtain radiation dose estimates in target organs.

Because continuous patient imaging is not feasible, the determination of source organ activities over time is usually achieved by interpolation between the data collected at specific time points and further extrapolation to infinity beyond the last measurement. For generation of such time-activity curves, a set of linear and exponential segments or a smooth curve derived from a compartment model can be used. To minimize the risk of an inaccurate extrapolation that could lead to significant under- or overestimation of the dose to target organs, it is recommended that data points be collected to cover 3 effective half-lives of the therapeutic compound (13). Alternatively, a conservative approach may be chosen by assuming that, beyond the last measurable time point, the compound is retained in the organ and that the activity is declining only by radioactive decay.

As previously discussed, the therapeutic compound DOTATOC labeled with ^{90}Y , a pure β -particle emitter, is not suitable for quantitative imaging. The substitution of an isotope of yttrium (^{86}Y , a positron emitter) emitting an in vivo detectable particle provides an appropriate surrogate to predict the in vivo behavior of the therapeutic compound ^{90}Y -DOTATOC. Positron emitters allow the use of PET imaging that provides increased quantitative accuracy and spatial resolution. Unfortunately, the decay characteristics of ^{86}Y also involve 2 drawbacks requiring careful attention. First, ^{86}Y also emits a high number of prompt γ -rays responsible for overestimations of activity that may reach 100% and 80% in the background and kidneys, respectively, when not properly accounted for. This is particularly true in 3-dimensional-mode acquisition. Several correction meth-

ods have been described, including a simple background subtraction or a sinogram tail subtraction (14,15). We proposed a patient-dependent correction method based on sinogram tail fitting using an ^{86}Y point spread function library, which was shown to significantly improve the accuracy of quantification (16). Second, the relatively low branching (33%) and short physical half-life of ^{86}Y (14.6 h) result in a very low counting rate at 48 h after injection, which implies that beyond this time point activities can be estimated only by extrapolation.

Two somatostatin analogs labeled with ^{111}In , a single-photon emitter, were used in dosimetry studies (10,12). Despite the lower sensitivity of SPECT compared with PET, the longer physical half-life (67.4 h) of this radionuclide allows collection of data up to at least 72 h. In addition to the fact that minor structural changes (metal or metal chelator) may lead to significant changes in biodistribution characteristics, it remains an arduous task to obtain accurate quantitative measurements with single-photon imaging (7,17). Planar acquisitions should be avoided, because they do not allow the actual delineation of organs of interest. The major drawback of quantification with SPECT arises from the important contribution (up to 40%) of scattered γ -rays. No accurate correction has been proposed, especially when the 2 energy peaks must be recorded to preserve the counting rate. The use of an effective attenuation coefficient or the subtraction of events acquired in a lower energy window are the most widely used techniques for scatter correction (18). When the attenuation coefficient map is known, an exact attenuation correction theoretically can be performed with SPECT using an iterative algorithm. Despite all these drawbacks, the use of an ^{111}In -labeled surrogate remains attractive, because it is commercially available and does not require PET technology.

Once the time integrals of activities in organs of interest are measured, absorbed dose calculations in target organs are generally performed using dedicated software, such as the MIRDOSE program, in which S values from any source to any target are integrated (13,19). Although age- and sex-specific reference data for human anatomy are included in the program, the estimated doses derived are far from patient-specific or even accurate, because they take into account neither patient-specific differences in organ shape or size nor nonuniform distribution of activity within the source organs. It is also important to remember that when the absorbed dose in a target is mainly related to self-irradiation (source = target), the S factor highly depends on the target volume. Because of the short range of ^{90}Y decay particles, S factors for targets with a mass greater than 1 cm³ are almost inversely proportional to the target volume. A good approximation is to measure the volume of the patient tissue and rescale the dose obtained from standard man or woman S factors (13). Significant improvement in the accuracy of measured doses may thus be achieved by precise

organ volume measurement with use of high-spatial-resolution techniques such as CT or MR imaging.

Finally, it should be remembered that toxicity results from a competition between DNA damage and enzymatic repair mechanisms (20,21). This is related to the ratio between the dose rate and the DNA repair half-time and can be described by the linear-quadratic model (20). Although correlation between biologic effects and radiation doses can be estimated in healthy people after radiation accidents or in patients treated by external beam radiotherapy, the dose rate with ^{90}Y internal radiotherapy (with a half-life of 62 h) may vary significantly and must be considered for estimation of the biologic effects (21–23). Recent data on low-dose hyper-radiosensitivity-inducible radiation repair could also have significant implications for the understanding of the relationship between toxicity and absorbed dose. Studies by Joiner et al. (24) identified a region of high sensitivity in the radiation survival response of mammalian and human cells at doses lower than 0.5 Gy with low dose rates. This hypersensitivity phenomenon is observed with low linear energy transfer (LET) radiation (e.g., x- and γ -rays) but not with high-LET radiation (e.g., neutron beams). Studying the survival of CA20948 rat pancreatic tumor cells incubated with increasing amounts of ^{131}I leading to doses of 1–10 Gy, Verwijnen et al. (25) found no hypersensitivity effects. Additional studies are needed to evaluate whether and to what extent low-dose hypersensitivity occurs in human tissues during internal therapy using peptide analogs labeled with a β -particle emitter such as ^{90}Y .

KIDNEY DOSIMETRY

The kidney is the major critical organ during therapy with ^{90}Y -DOTATOC. Once filtered by glomeruli, radiolabeled peptides are reabsorbed and retained in proximal tubular cells (6,26). Although kidney uptake is limited, renal toxicity can occur because of irradiation of the radiosensitive glomeruli by activity present in the radioresistant tubular cells (3–5). Thrombotic microangiopathy is the main histologic finding related to sublethal endothelial cell irradiation (4). With external beam radiotherapy, it is recommended that a kidney absorbed dose of 23 Gy not be exceeded. This is the absorbed dose that has been shown to induce a 5% risk of nephrotoxicity at 5-y follow-up (23). Whether this holds true for internal irradiation remains unclear.

In vivo studies showed a wide inter-patient variability of ^{86}Y -DOTATOC kidney uptake, supporting the need to perform direct individual measurements. In 4 patients, no uptake could be delineated in the kidney on PET imaging. These patients were assumed to be able to receive high activities without a risk of renal side effects. In others, the renal uptake, mean residence times (MRTs), and, therefore, radiation doses varied by a factor of 4 (range, 1.2–5.1 mGy/MBq) (9). The same variability in renal absorbed doses (range, 2.9–5.7 mGy/MBq) was reported by Bodei et

al. (27) in a series of 6 patients who underwent dosimetry studies with ^{111}In -DOTATOC without amino acid infusion.

We treated a series of 60 patients after individual kidney dose estimation using ^{86}Y -DOTATOC imaging and the MIRDSE3.1 model. Despite the fact that the activities administered to each patient were calculated so that the expected doses delivered to the kidneys would not exceed a fixed limit of 27 Gy, various degrees of nephropathy were experienced by 5 patients, with 1 patient on hemodialysis at 5-y follow-up. We investigated possible factors that could affect the accuracy of pretherapeutic dose estimates. The most important of these seems to be the actual kidney volume. Indeed, the MIRDSE model refers to standard kidney volumes for males and females (288 and 264 mL, respectively). Retrospective determination of individual kidney volumes on pretherapeutic CT images in 25 patients showed high interindividual variability (range, 231–503 mL), indicating that individual kidney volume determination might be crucial to improve the determination of kidney radiation doses. Taking the actual renal volume measured by CT into account resulted in a median undercorrection of the absorbed dose by 11%. However, patients who developed renal toxicity had significantly smaller kidneys as assessed by CT and, in fact, received significantly higher kidney doses compared with patients who experienced no renal toxicity.

Methods usually proposed to estimate organ-absorbed doses require the assumption of a homogeneous activity distribution over the entire organ. In a recent improvement of the MIRD model, Bouchet et al. (28) proposed a multi-region model for the kidney, including 4 main homogeneous regions as source/target: cortex, medulla, pelvis, and papillae. The kidney dose estimates could be made more patient-specific by rescaling the 4 kidney regions on the basis of the actual kidney volume measured by CT or MR imaging. To make use of this model, it is essential to know the activity distribution inside the kidney. Unfortunately, this can not be achieved by quantitative radionuclide imaging because of the lack of spatial resolution of PET and SPECT cameras. However, a more precise pattern of activity distribution inside the kidney has been established by *ex vivo* studies. Autoradiography of a human kidney obtained after *in vivo* injection of ^{111}In -labeled somatostatin analog showed radioactivity to be concentrated predominantly in the juxtamedullary region of the cortex (29,30). Transposing this heterogeneous pattern of activity in a voxel-based model, the authors showed a 2-fold increase in the dose estimates in 29% of the cortex volume compared with the estimates produced by a single-region model assuming a homogeneous organ activity distribution.

Other factors that may influence the radiobiologic effects of internal irradiation should also be taken into consideration. As noted previously, in fractionated external beam therapy, the dose limit causing a 5% probability of radiation nephropathy within 5 y has been found to be 23 Gy. This

dose is typically given at a high dose rate in fractions of 1.5–2 Gy. In contrast, during ^{90}Y -DOTATOC therapy, radiation exposure is continuous and exponentially decaying with an effective half-life of 48 h. It was recently suggested that the linear–quadratic model could be used to transpose the external beam therapy dose limit (23 Gy) to a corresponding limit for radionuclide therapy with ^{90}Y -DOTATOC. According to this model, which takes into account the rate and fractionation of the dose, the 5% level of probability of nephrotoxicity shifts from the 23 Gy threshold for external beam therapy to 35 ± 7 Gy for ^{90}Y -DOTATOC (31).

Thus, the actual dose to the glomeruli may be twice that estimated with the classic MIRD model based on fixed kidney size and homogeneous activity distribution within the whole organ. On the other hand, the maximum tolerable kidney dose for internal radiotherapy with ^{90}Y -DOTATOC therapy is significantly higher than the dose limit for external beam therapy.

RED MARROW

Assessment of red marrow dosimetry in internal radiotherapy is of great importance (32). With red marrow irradiation, we are entering another world in terms of dose assessment. For most organs, significant toxicity may occur for delivered doses of some 10s of Gy, whereas doses as low as 3 Gy to the red marrow induce 1% of leukemia within 10 y after exposure, and the probability of survival decreases rapidly beyond 4 Gy to reach an LD50 (death of 50% of individuals) between 5 and 6 Gy (33). This implies that the red marrow maximal tolerated dose will be reached for a specific uptake much lower than the uptake in other organs. Another hurdle in estimating the red marrow absorbed dose is its nonhomogeneous nature, with a complex mixture of trabecular bone, cortical bone, active red marrow, and inactive marrow. Improvements to the MIRD model were proposed to take this microstructure into account in S factor computation (34).

It was originally reported that no specific uptake in red marrow was noted on ^{111}In -DTPA-OC scan, and red marrow activity was assumed to be a fraction of plasma activity or equal to the remainder of the body (10,12). Recent studies using ^{86}Y -DOTATOC and PET showed that somatostatin analogs are also taken up in the red marrow but not in the trabecular or cortical bone (Fig. 1) (35). The mechanisms of uptake are not entirely elucidated and may include specific binding to SSR, transchelation of the metal label to transferrin, or plasma activity.

To avoid direct superimposition of organ activity or cross-contamination by scatter γ -rays, it is recommended that the red marrow be measured over a segment of the thoracic spine region and rescaled to the whole red marrow mass using the standard fraction of active red marrow present in the thoracic spine (32,35). The delivered dose can

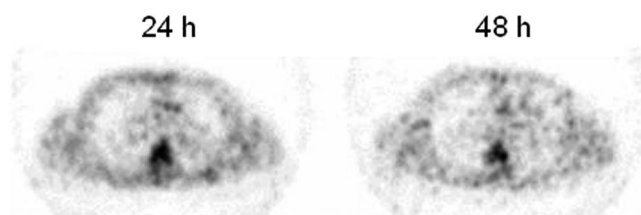


FIGURE 1. Transverse PET slices through the chest obtained 24 and 48 h after injection of 0.4 GBq of ^{86}Y -DOTATOC. Note the clear uptake over the spine area, whereas areas without active red marrow, such as the humerus, were not visualized.

than be computed using the correction of the MIRDose program using MRTs as input (36).

Studies showed that planar imaging techniques for ^{90}Y -labeled antibodies binding to red marrow improved the correlation between myelotoxicity and estimated dose delivered to red marrow (32,37). Some authors suggested that, to predict the myelotoxicity as a function of the delivered dose to the red marrow, patients should first be sorted into populations with different red marrow sensitivities (Fig. 2) (37–40).

Investigations after radiation accidents in healthy people showed that for sublethal doses the platelet count recovery (PCR) 6 wk after irradiation was almost complete (21,22). In ^{90}Y -DOTATOC therapy, the platelet count nadir (PCN) for patients with a normal PCR as a function of the red marrow dose, estimated using ^{86}Y -DOTATOC PET, was in good agreement with observations in radiation accidents (Fig. 2) (35). In contrast, patients with an incomplete PCR already presented an abnormally low PCN. Half of those patients had undergone chemotherapy, known to alter the red marrow reserve. No patients with normal PCRs had undergone chemotherapy.

OTHER ORGANS

For technical reasons, quantification of other organs using the methodology described here is not feasible. Accurate estimation of liver exposure to internal irradiation is hampered by the almost constant and often bulky presence of liver metastases. Absorbed doses to the pituitary and thyroid cannot be precisely measured because of the small size of the organs. However, scintigraphic images show lower uptakes in all these organs than in the kidneys, probably resulting in an absorbed dose far below 20 Gy. Such doses should not be deleterious to these organs. This may not be the case for the testes, which probably receive a substantial radiation dose as illustrated by the drop in inhibin B levels after treatment with ^{90}Y -DOTATOC (41).

TUMOR DOSE-RESPONSE RELATIONSHIPS

Despite differences in the protocols used, tumor size reduction of at least 25% (range, 10%–30%) has been reported in most studies with ^{90}Y -DOTATOC (27,42–44).

This relatively low rate of complete or partial response is a result of the fact that neuroendocrine gastroenteropancreatic tumors are usually radioresistant and also because there is a limit to the therapeutic activities that can be delivered safely. There is a need for predictive parameters that could help the clinician select more appropriate candidates for treatment and, conversely, identify those patients who would not benefit from such treatment.

In a subset of 13 patients treated with ^{90}Y -DOTATOC, we found a significant correlation between dose estimates to the tumors and the tumor size reduction measured at the end of the treatment (Fig. 3) (45). A similar correlation between tumor dose and response was still present 1 y after end of treatment. Dose estimates were derived from quantitative PET with ^{86}Y -DOTATOC, and the MIRDose spheric model was refined by using the actual tumor sizes measured by CT. The median absorbed dose was 6-fold higher in responding tumors (232 Gy) than the median value in nonresponding tumors (37Gy). It is noteworthy that in only 20% of the nonresponding tumors, dose estimates were in the range of those in responding tumors. Although obtained in a limited number of patients, these dose–response relationships suggest the potential usefulness of tumor dosimetry in the management of patients with neuroendocrine tumors.

CONCLUSION

Although it is clear that accurate prediction of absorbed doses is required for internal radiotherapy, methods developed up to now do not enable a simple approach. Even with sophisticated techniques, inaccuracies can occur and result

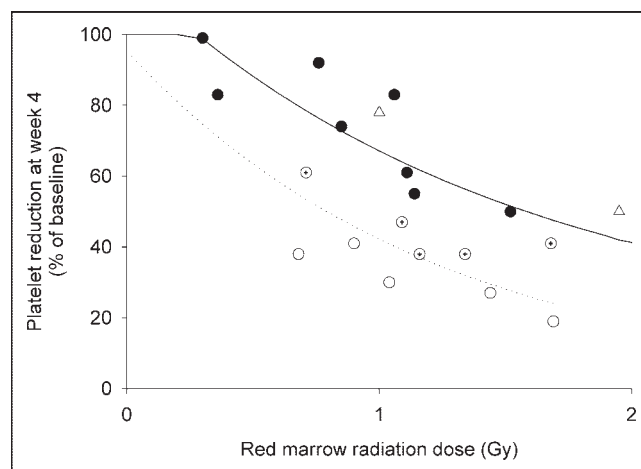


FIGURE 2. Correlation between the platelet count nadir observed after ^{90}Y -DOTATOC therapy and the red marrow radiation doses estimated using ^{86}Y -DOTATOC. ● = patients with subsequent complete platelet count recovery; ○ = patients with a partial platelet count recovery; ⊕ = patients who previously underwent chemotherapy but did not completely recover their platelet counts; △ = platelet count nadir observed in radiation accident in healthy individuals.

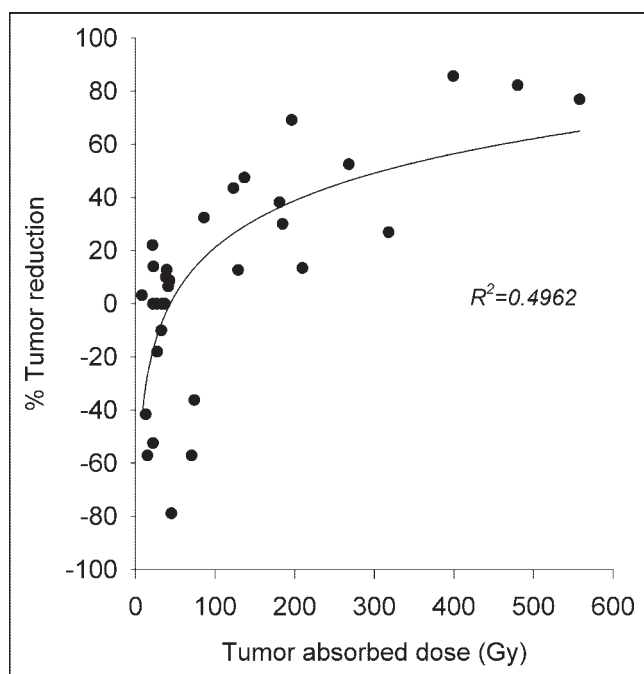


FIGURE 3. Tumor dose-response relationship in 13 patients treated with ^{90}Y -DOTATOC. Tumor volumes were assessed by CT before and after treatment. Tumor dose estimates were derived from CT scan volume measurements and quantitative ^{86}Y -DOTATOC imaging performed before treatment. Data were further computed using the MIRDose spheric model.

in overestimation of the absorbed doses, resulting in undertreatment, or in underestimation, resulting in overtreatment and unpredicted toxicity. One of the major unknowns in internal dosimetry is the effect on cells of very low dose rates and of low-energy γ - and x-ray emissions. Additional research is needed in this field.

REFERENCES

- de Jong M, Kwekkeboom D, Valkema R, Krenning EP. Radiolabelled peptides for tumour therapy: current status and future directions. Plenary lecture at the EANM 2002. *Eur J Nucl Med*. 2003;30:463–469.
- Valkema R, De Jong M, Bakker WH, et al. Phase I study of peptide receptor radionuclide therapy with [^{111}In -DTPA]octreotide: the Rotterdam experience. *Semin Nucl Med*. 2002;32:110–122.
- Otte A, Mueller-Brand J, Dellas S, Nitzsche EU, Herrmann R, Maecke HR. Yttrium-90-labelled somatostatin analogue for cancer treatment. *Lancet*. 1998;351:417–418.
- Moll S, Nickenleit V, Mueller-Brand J, Brunner FP, Maecke HR, Mihatsch MJ. A new cause of renal thrombotic microangiopathy: yttrium 90-DOTATOC internal radiotherapy. *Am J Kidney Dis*. 2001;37:847–851.
- Cybulka M, Weiner SM, Otte A. End-stage renal disease after treatment with ^{90}Y -DOTATOC. *Eur J Nucl Med*. 2001;28:1552–1554.
- Behr TM, Goldenberg DM, Becker W. Reducing the renal uptake of radiolabeled antibody fragments and peptides for diagnosis and therapy: present status, future prospects and limitations. *Eur J Nucl Med*. 1998;25:201–212.
- Reubi JC, Schar JC, Waser B, et al. Affinity profiles for human somatostatin receptor subtypes SST1–SST5 of somatostatin radiotracers selected for scintigraphic and radiotherapeutic use. *Eur J Nucl Med*. 2000;27:273–282.
- Heppeler A, Froidevaux S, Eberle AM, Maecke HR. Receptor targeting for tumor localization and therapy with radiopeptides. *Curr Med Chem*. 2000;7:971–994.
- Jamar F, Barone R, Mathieu I, et al. ^{86}Y -DOTA⁰-D-Phe¹-Tyr³-octreotide (SMT487). A phase I clinical study: pharmacokinetics, biodistribution and renal protective effect of different regimens of amino acid co-infusion. *Eur J Nucl Med*. 2003;30:510–518.
- Förster GJ, Engelbach MJ, Brockmann J, et al. Preliminary data on biodistribution and dosimetry for therapy planning of somatostatin receptor positive tumours: comparison of ^{86}Y -DOTATOC and ^{111}In -DTPA-octreotide. *Eur J Nucl Med*. 2001;28:1743–1750.
- Barone R, Jamar F, Walrand S, et al. Can ^{111}In -DTPA-octreotide (In-OC) predict kidney and tumor exposure during treatment with ^{90}Y -SMT487 (Octreother) [abstract]? *J Nucl Med*. 2000;41(suppl):110P.
- Cremonesi M, Ferrari M, Zoboli S, et al. Biokinetics and dosimetry in patients administered with ^{111}In -DOTA-Tyr³-octreotide: implication for internal radiotherapy with ^{90}Y -DOTATOC. *Eur J Nucl Med*. 1999;26:877–886.
- Stabin MG. Developments in the internal dosimetry of radiopharmaceuticals. *Radiat Prot Dosim*. 2003;105:575–580.
- Buchholz HG, Herzog H, Forster GJ, et al. PET imaging with yttrium-86: comparison of phantom measurements acquired with different PET scanners before and after applying background subtraction. *Eur J Nucl Med Mol Imaging*. 2003;30:716–720.
- Pentlow KS, Finn RD, Larson SM, Erdi YE, Beattie BJ, Humm JL. Quantitative imaging of yttrium-86 with PET: the occurrence and correction of anomalous apparent activity in high density regions. *Clin Positron Imaging*. 2000;3:85–90.
- Walrand SH, Jamar F, Mathieu I, et al. Quantification in PET using isotopes emitting prompt single gammas: application to ^{86}Y . *Eur J Nucl Med*. 2003;30:354–361.
- Pauwels S, Barone R, Carlier P, et al. Biodistribution of yttrium- and lutetium-labeled DOTATOC and DOTATATE in rat tumor model [abstract]. *J Nucl Med*. 2002;43(suppl):123P.
- Buvat I, Benali H, Todd-Pokropek A, Di Paola R. Scatter correction in scintigraphy: the state of the art. *Eur J Nucl Med*. 1994;21:675–694.
- Stabin MG. MIRDose. Personal computer software for internal dose assessment in nuclear medicine. *J Nucl Med*. 1996;37:538–546.
- United Nations Scientific Committee on the Effects of Ionizing Radiation. *Sources and Effects of Ionizing Radiation. Volume 2. Effects. United Nations Scientific Committee on the Effects of Atomic Radiation 2000 Report to the General Assembly with Scientific Annexes. Annex G*. New York, NY: United Nations; 2000:73–175.
- United Nations Scientific Committee on the Effects of Ionizing Radiation. *Sources, Effects and Risks of Ionizing radiation. United Nations Scientific Committee on the Effects of Atomic Radiation 1988 Report to the General Assembly with Scientific Annexes. Annex G*. New York, NY: United Nations; 1988:1–21.
- Space Radiation Study Panel of the Life Sciences Committee. Radiological responses in man. In: Langham WH, ed. *Radiobiological Effects in Manned Space Flight*. Washington, DC: National Academy of Sciences and National Research Council; 1967:57–239.
- Emami B, Lyman J, Brown A, et al. Tolerance of normal tissue to therapeutic irradiation. *Int J Radiat Oncol Biol Phys*. 1991;21:109–122.
- Joiner MC, Marples B, Lambin P, Short SC, Turesson I. Low-dose hypersensitivity: current status and possible mechanisms. *Int J Radiat Oncol Biol Phys*. 2001;49:379–389.
- Verwijnen S, Capello A, Bernard B, et al. Low-dose-rate irradiation by ^{131}I versus high-dose-rate external-beam irradiation in the rat pancreatic tumor cell line CA20948. *Cancer Biother Radiopharm*. 2004;19:285–292.
- Akizawa H, Arano Y, Uezono T, et al. Renal metabolism of ^{111}In -DTPA-D-Phe¹-octreotide in vivo. *Bioconjug Chem*. 1998;9:662–670.
- Bodei L, Cremonesi M, Zoboli S, et al. Receptor-mediated radionuclide therapy with ^{90}Y -DOTATOC in association with amino acid infusion: a phase I study. *Eur J Nucl Med*. 2003;30:207–216.
- Bouchet LG, Bolch WE, Blanco HP, et al. Absorbed fractions and radionuclide S values for six age-dependent multiregion models of the kidney. MIRD pamphlet No. 19. *J Nucl Med*. 2003;44:1113–1147.
- de Jong M, Valkema R, van Gameren A, et al. Inhomogeneous localization of radioactivity in the human kidney after injection of [^{111}In -DTPA]octreotide. *J Nucl Med*. 2004;45:1168–1171.
- Konijnenberg MW, de Jong M, Kooij PP, Krenning EP. Dosimetric consequences for radionuclide therapy because of inhomogeneous radioactivity uptake in human renal cortex by radiolabelled octreotide [abstract]. *J Nucl Med*. 2003;44(suppl):101P.
- Konijnenberg MW. Is the renal dosimetry for [^{90}Y -DOTA⁰,Tyr³]octreotide accurate enough to predict thresholds for individual patients? *Cancer Biother Radiopharm*. 2003;18:619–625.
- Sgouros G, Stabin M, Akabani G, Kwok C, Brill AB, Wessels B. Red marrow dosimetry for radiolabeled antibodies that bind to marrow, bone, or blood components. *Med Phys*. 2000;27:2150–2164.

33. Boice JD, Inskip PD. Radiation-induced leukemia. In: Henderson ES, Lister TA, Greaves, eds. *Leukemia*. Philadelphia, PA: W.B. Saunders; 1996:195–209.
34. Stabin MG, Eckerman KF, Bloch WE, Bouchet LG, Patton PW. Evolution and status of bone and marrow dose models. *Cancer Biother Radiopharm*. 2002;17: 427–433.
35. Walrand S, Barone R, Jamar F, et al. Red marrow ^{90}Y -OctreoTher dosimetry estimated using ^{86}Y -OctreoTher PET and biological correlates [abstract]. *Eur J Nucl Med Mol Imaging*. 2002;29(suppl):S301.
36. Stabin MG, Siegel JA, Sparks RB, Eckerman KF, Breitz HB. Contribution to red marrow absorbed dose from total body activity: a correction to the MIRD method. *J Nucl Med*. 2001;42:492–498.
37. Shen S, Meredith RF, Duan J, et al. Improved prediction of myelotoxicity using a patient specific imaging dose estimate for non-marrow-targeting ^{90}Y -antibody therapy. *J Nucl Med*. 2002;43:1245–1253.
38. Zanzonico P, Sgouros G. Predicting myelotoxicity in radioimmunotherapy: what does dosimetry contribute? *J Nucl Med*. 1997;38:1753–1754.
39. Lim S, DeNardo GL, De Nardo DA, et al. Prediction of myelotoxicity using semiquantitative marrow image scores. *J Nucl Med*. 1997;38:1374–1378.
40. Behr TM, Béhé M, Sgouros G. Correlation of red marrow radiation dosimetry with myelotoxicity: empirical factors influencing the radiation-induced myelo-toxicity of radiolabeled antibodies, fragments and peptides in preclinical and clinical settings. *Cancer Biother Radiopharm*. 2002;17:445–464.
41. Valkema R, Jamar F, Bakker WH, et al. Safety and efficacy of [^{90}Y -DOTA, Tyr(3)]-octreotide (Y-90-SMT487; Octreother) peptide receptor radionuclide therapy (PRRT). Preliminary results of phase-1 study [abstract]. *Eur J Nucl Med*. 2001;28(suppl):1025.
42. Valkema R, Pauwels S, Kvols L, et al. Long-term follow-up of a phase 1 study of peptide receptor radionuclide therapy (PRRT) with [^{90}Y -DOTA⁰,Tyr³]-octreotide in patients with somatostatin receptor positive tumours [abstract]. *Eur J Nucl Med*. 2003;30(suppl):S232.
43. Chinol M, Bodei L, Cremonesi M, Paganelli G. Receptor-mediated radiotherapy with ^{90}Y -DOTA-D-Phe¹-Tyr³-octreotide: the experience of the European Institute of Oncology Group. *Semin Nucl Med*. 2002;32:141–147.
44. Waldherr C, Pless M, Maecke HR, et al. Tumor response and clinical benefit in neuroendocrine tumors after 7.4 GBq ^{90}Y -DOTATOC. *J Nucl Med*. 2002;43: 610–616.
45. Jonard P, Jamar F, Walrand S, et al. Tumor dosimetry based on PET ^{86}Y -DOTA-Tyr³-octreotide (SMT487) and CT-scan predicts tumor response to ^{90}Y -SMT487 (OctreotherTM) therapy [abstract]. *J Nucl Med*. 2000;41(suppl):111P.





The Journal of
NUCLEAR MEDICINE

Practical Dosimetry of Peptide Receptor Radionuclide Therapy with ^{90}Y -Labeled Somatostatin Analogs

Stanislas Pauwels, Raffaella Barone, Stéphan Walrand, Françoise Borson-Chazot, Roelf Valkema, Larry K. Kvols, Eric P. Krenning and François Jamar

J Nucl Med. 2005;46:92S-98S.


This article and updated information are available at:
http://jnm.snmjournals.org/content/46/1_suppl/92S

Information about reproducing figures, tables, or other portions of this article can be found online at:
<http://jnm.snmjournals.org/site/misc/permission.xhtml>

Information about subscriptions to JNM can be found at:
<http://jnm.snmjournals.org/site/subscriptions/online.xhtml>

The Journal of Nuclear Medicine is published monthly.
SNMMI | Society of Nuclear Medicine and Molecular Imaging
1850 Samuel Morse Drive, Reston, VA 20190.
(Print ISSN: 0161-5505, Online ISSN: 2159-662X)

© Copyright 2005 SNMMI; all rights reserved.

 SOCIETY OF
NUCLEAR MEDICINE
AND MOLECULAR IMAGING