

Review

## The Role of Thiamine in Cancer: Possible Genetic and Cellular Signaling Mechanisms

KHANH VINH QUỐC LƯƠNG and LAN THI HOÀNG NGUYỄN

*Vietnamese American Medical Research Foundation, Westminster, CA, U.S.A.*

**Abstract.** *The relationship between supplemental vitamins and various types of cancer has been the focus of recent investigation, and supplemental vitamins have been reported to modulate cancer rates. A significant association has been demonstrated between cancer and low levels of thiamine in the serum. Genetic studies have helped identify a number of factors that link thiamine to cancer, including the solute carrier transporter (SLC19) gene, transketolase, transcription factor p53, poly(ADP-ribose) polymerase-1 gene, and the reduced form of nicotinamide adenine dinucleotide phosphate. Thiamine supplementation may contribute to a high rate of tumor cell survival, proliferation and chemotherapy resistance. Thiamine has also been implicated in cancer through its effects on matrix metalloproteinases, prostaglandins, cyclooxygenase-2, reactive oxygen species, and nitric oxide synthase. However, some studies have suggested that thiamine*

*may exhibit some antitumor effects. The role of thiamine in cancer is controversial. However, thiamine deficiency may occur in patients with cancer and cause serious disorders, including Wernicke's encephalopathy, that require parenteral thiamine supplementation. A very high dose of thiamine produces a growth-inhibitory effect in cancer. Therefore, further investigations of thiamine in cancer are needed to clarify this relationship.*

The relationship of supplemental vitamins and various types of cancer has been the focus of recent investigations, and supplemental vitamins have been reported to modulate cancer rates. In a New Hampshire study, higher total intakes of carotenoids, vitamin D, thiamine, niacin, and vitamin E were inversely related to bladder cancer risk among older individuals (1). Higher intake of vitamin C, beta-carotene, thiamine, and nicotinic acid was also associated with a decrease in gastric cancer risk in a case control study (2). In Hawaii, riboflavin and thiamine levels from food sources exhibited an inverse relationship with incidence of high-grade squamous intraepithelial lesions of the cervix (3). In Poland, higher consumption of alcohol, when combined with a low intake of retinol, thiamine or antioxidant micronutrients, increased the risk of colorectal cancer and established a cancer-preventive role for retinol and thiamine (4). High dietary intake of vitamin C and thiamine has been reported to reduce the risk of prostate cancer in China (5). In another study, Hargreaves *et al.* (6) suggested that low intake of thiamine, riboflavin, vitamins A and C, and iron may contribute to cancer risk in African-Americans. When monitoring micronutrient levels in gastrointestinal cancer, plasma vitamin C and red blood cell thiamine levels were significantly reduced in weight-losing patients with cancer compared with their weight-stable counterparts, and weight-losing patients exhibited lower vitamin C and thiamine intake (7). Furthermore, patients with acute leukemia exhibited low thiamine content in leukocytes and blood plasma (8). The thiamine pyrophosphate (TPP) stimulating effect and urinary excretion of thiamine were high in

**Abbreviations:** 15-PGDH: 15-Hydroxyprostaglandin dehydrogenase; ATP: adenosine thiamine triphosphate; CNS: central nervous system; DKA: diabetic ketoacidosis; ECM: extracellular matrix; COX: cyclooxygenase; DM: diabetes mellitus; HCC: hepatocellular carcinoma; KTS: 3-ethoxy-2-oxobutyraldehyde bis(thiosemicarbazone); iNOS: inducible NOS; MMP: matrix metalloproteinase; NADPH: the reduced form of nicotinamide adenine dinucleotide phosphate; NO: nitric oxide; NOS: nitric oxide synthase; NOX: the NADPH oxidase; LPS: lipopolysaccharide; PARP-1: Poly(ADP-ribose) polymerase-1; PDH: pyruvate dehydrogenase; PG: prostaglandins; RDA: recommended dietary allowance; ROS: reactive oxygen species; SLC: solute carrier transporter; SOD: superoxide dismutase; TD: thiamine-deficient; TDP: thiamine diphosphate; ThTk: thiamine transporter; TKTL-1: transketolase-like 1; TPP: thiamine pyrophosphate; TIMP-1: tissue inhibitor of metalloproteinase-1.

**Correspondence to:** Khanh v.q. Luong, MD, FACP, FACE, FACN, FASN, FCCP, and FACAAI (SC), 14971 Brookhurst St. Westminster, CA 92683, U.S.A. Tel: +1 7148395898, Fax: +1 7148395989, email: lng2687765@aol.com

**Key Words:** Thiamine, cancer, vitamin B1, transketolase, tumor, review.

patients with breast and bronchial carcinomas, suggesting that these patients may be at risk for thiamine deficiency (9). Transketolase reactions play a crucial role in the use of glucose carbons in tumor cell synthesis of nucleic acid ribose. Experimentally, a dramatic decrease in tumor cell proliferation was observed in several *in vitro* and *in vivo* tumor models after administering the transketolase inhibitor oxythiamine (10, 11). Supplementation with excess thiamine and pantothenic acid in tumor-bearing rats reduced growth restriction and enhanced the antitumor activity of 3-ethoxy-2-oxobutyaldehyde bis(thiosemicarbazone) (KTS). Similarly, the addition of thiamine protected the KTS-treated rats from weight loss in a dose-related manner (12). These findings suggest that thiamine may be associated with cancer. Therefore, we reviewed the role of thiamine in cancer.

### The Genomic Factors Associated with Thiamine in Cancer

*The solute carrier (SLC) transport protein.* The *SLC19* gene is a family of three transporter proteins with significant structural similarities, but each protein binds substrates with different structures and ionic charges. The *SLC19* gene family plays an important role in the transport and homeostasis of folate and thiamine in the body (13). *SLC19A2* and *SLC19A3* mediate the transport of thiamine (14, 15). *SLC19A3* RNA levels have been reported to be down-regulated in breast and lung cancer compared with adjacent normal tissues (16, 17). The *SLC19A3* gene is regulated by exogenous thiamine in a manner similar to the effect of *SLC19A3* overexpression (17). *SLC19A3* expression is down-regulated in gastric cancer cell lines (71%) and restored after pharmaceutical demethylation. Exogenous *SLC19A3* expression causes growth inhibition of gastric cancer cells (17). *SLC19A3* gene expression is down-regulated in breast cancer and associated with resistance to apoptosis in these tumors (18). Its mRNA expression is also down-regulated by DNA methylation, and by histone deacetylation in colon cancer cells (19). However, hypoxia induces the up-regulation and function of the *SLC19A3* gene in a breast cancer cell line (20). Aberrant *SLC19A3* has been suggested as a novel biomarker for breast and gastric cancer diagnosis (21). *SLC19A2* has also been shown to transport mono- and pyrophosphate derivatives of thiamine (22, 23). Polymorphisms of *SLC19A1* are reported in many types of cancers, such as osteosarcoma (24), colorectal cancer (25), non-small cell lung cancer (26), bladder cancer (27), prostate cancer (28), diffuse large B-cell lymphoma (29), pediatric acute lymphoblastic leukemia (30), head and neck cancer (31), central nervous system (CNS) tumors in Thai children (32), and human ovarian carcinoma cells (33).

*Transketolase.* Thiamine acts as a co-enzyme for transketolase, pyruvate dehydrogenase (PDH), and  $\alpha$ -ketoglutarate dehydrogenase complexes, which contain enzymes with fundamental roles in intracellular glucose metabolism. Thiamine also regulates the expression of genes that code for enzymes using thiamine as a co-factor. Thiamine deficiency reduces the mRNA levels of transketolase and PDH (34). In the blood of patients with cancer, the concentration of thiamine and thiamine diphosphate (TDP) are lowered by 20%; transketolase activity is decreased by 20%, and the TDP effect by 5-42% (35). Moreover, aberrant levels of transketolase and transketolase-like-1 (TKTL-1) participated in glucose metabolism in malignant cells in pleural effusion (36). TKTL-1 has been reported to play a pivotal role in carcinogenesis and has an important role in controlling the non-oxidative pentose-phosphate pathway, which is significantly up-regulated in cancer. Increased TKTL-1 expression has been shown to be correlated with poor patient outcome and increased tumor progression (37). Additionally, increased TKTL 1 has been observed in many types of cancer, including thyroid cancer (38, 39), nasopharyngeal carcinoma (40, 41), non-small cell lung cancer (42, 43), gastric cancer (44), human hepatoma cell (45), colorectal cancer (46), urothelial cancer (47), anaplastic neuroblastoma (48), breast cancer (49), endometrial cancer (50), ovarian cancer (51, 52), cancer of the uterine cervix (53), head and neck cancer (54), and glioblastoma (55). However, the related proteins TKTL-1 and TKTL-2 lack numerous invariant residues involved in co-factor and substrate binding of TPP and are therefore not expected to possess transketolase activity (56).

*Transcription factor p53.* The p53 gene and protein play a critical role in the regulation of the normal cell cycle, cell cycle arrest, and the apoptotic response. p53 is a transcription factor with a major role in determining cell fate in response to DNA damage. p53 reduces the incidence of cancer by promoting apoptosis in cells that have activated oncogenic pathways. p53 polymorphisms have been demonstrated in breast cancer (57), mesothelioma (58), esophageal squamous cell carcinoma (59), and pancreatic cancer (60). In many meta-analyses, p53 polymorphisms have been reported to be associated with cervical (61), gastric (62), bladder (63), colorectal (64), and endometrial (65) cancer, but not with prostate cancer (66). Increased thiamine transporter levels have been observed in murine cells that overexpress mouse *SLC19A2* or in cells exposed to conditions that induce DNA damage or p53 activation (67). TDP has been shown to inhibit p53 binding and thiamine has been shown to inhibit intracellular p53 activity (68). The expression of p53 decreased significantly in cultured retinal neurons of diabetic rats treated with thiamine (69). These observations suggested that the transcription factor p53 is activated in cancer with

increasing apoptotic response from cellular damage and thiamine ameliorated these effects on cells.

*Poly(ADP-ribose) polymerase-1 (PARP-1)*. PARP 1 is a nuclear protein that contributes to both cell death and survival under stressful conditions. PARP-1 catalytic activity is stimulated by DNA strand breaks. In mice, *Parp-1*-deficient cells exhibited enhanced sensitivity to lethal effects of ionizing radiation and alkylating agents (70), and *Parp-1* knockout mice were found to develop spontaneous mammary and liver tumors (71, 72). Overexpression of PARP-1 has been reported in Ewing's sarcoma (73), malignant lymphoma (74), colorectal cancer (75), hepatocellular carcinoma (HCC) (76), breast cancer (77), pediatric CNS tumors (78), and ovarian cancer (79). In a meta-analysis, *PARP-1* mRNA expression was correlated with high-grade, medullary histological type, tumor size, and worse metastasis-free survival and overall survival in human breast cancer (80). *PARP-1* polymorphisms have been found to be associated with gastric cancer in Han Chinese (81), prostate cancer (82), esophageal squamous cell carcinoma (83), and lung cancer (84), and with a reduced risk of non-Hodgkin lymphoma in Korean males (85). *PARP 1* polymorphisms reduce PARP-1 catalytic activity by 30-40% (86). In a meta-analysis, no significant association was observed between the *PARP-1* V762 polymorphism and cancer risk; however, the variant A allele of the PARP-1 V762 polymorphism was associated with an increased risk of cancer within Asian populations, but with a decreased risk of cancer among Caucasians, particularly for glioma (87). However, thiamine has a cytoprotective effect on cultured neonatal rat cardiomyocytes under hypoxic insult; thiamine also inhibits PARP cleavage and DNA fragmentation (88). Benfotiamine, a fat-soluble thiamine analog, prevents bacterial endotoxin-induced inflammation and PARP cleavage in mouse macrophage cell lines (89). Adenosine thiamine triphosphate (ATTP) has been identified in small amounts in the mouse brain, heart, skeletal muscle, liver and kidneys (90) and has been shown to inhibit PARP-1 activity (91). Taken together, the results indicate that thiamine may play a role in cancer by modulating PARP-1 activity.

*The reduced form of nicotinamide adenine dinucleotide phosphate (NADPH)*. The NADPH oxidase (NOX) enzyme complex mediates critical physiological and pathological processes, including cell signaling, inflammation and mitogenesis, through the generation of reactive oxygen species (ROS) from molecular oxygen. NOX1 is required for RAS oncogene-induced cell transformation (92). The NOX subunit p22<sup>phox</sup> has been reported to inhibit the function of the tumor suppressor protein tuberlin in renal carcinoma cells (93). The activation of NOXs has been demonstrated in the development of cancer, including melanoma (94), leukemia

(95), esophageal adenocarcinoma (96), HCC (97), prostate cancer (98), colon cancer (99), glioblastoma multiforme (100), and multiple myeloma (101). *NADPH* polymorphisms have been reported to be associated with myelodysplastic syndrome and *de novo* acute myeloid leukemia (102, 103), esophagus cancer (104), lung cancer (105, 106), non-Hodgkin's lymphoma (107, 108), childhood acute leukemia (109), postmenopausal breast cancer (110), and gastric cancer (111). The levels of NADPH cytochrome *c* reductase are increased in thiamine-deficient animals (112). The expression of NOX4 is significantly down-regulated by benfotiamine treatment under both normo- and hyperglycemic conditions (113). In addition, animals fed a high thiamine diet exhibited NADPH-cytochrome *c* reductase activity of approximately 57% that of those fed a thiamine-deficient (TD) diet (114). Altogether, these results suggest that thiamine may produce a cancer-protective effect by regulating NADPH-cytochrome *c* activity.

The genetic factors related to thiamine and cancer are summarized in Table I.

### The Role of Thiamine in Cancer

*Diabetes mellitus*. Alterations in glucose metabolism have been linked to a significantly increased risk for many forms of cancer. Gestational diabetes mellitus (DM), a state of glucose intolerance associated with pregnancy, is associated with an increased risk of pancreatic cancer and hematological malignancies (115). DM and impaired glucose metabolism contribute to the risk of postmenopausal breast and endometrial cancer (116). The metabolic syndrome has been reported to be associated with the risk of prostate cancer in Italian populations (117). Type 1 DM is associated with an increase in the risk of cancer, particularly gastric, uterine cervix, endometrial, and ovarian cancer (118, 119). Several meta-analyses have demonstrated an association between DM and increased cancer risk, including esophageal (120), gastric (121), colorectal (122), HCC (123), cholangiocarcinoma (124), and breast cancer (125). The incidence of DM in renal cell carcinoma is higher in females than males and higher in Hispanic populations compared to Caucasian populations and other ethnicities (126). However, a relationship between thiamine and DM has also been reported in the literature. A significant proportion of healthy individuals (36-47%) have been reported to have thiamine deficiency while in a hyperglycemic state (such as on a diet high in carbohydrates or if diabetic or pregnant) (127). Low plasma thiamine levels have been noted in patients with DM type 1 (128). Thiamine reserves are reduced in litters of untreated diabetic rats (129). In children, acute thiamine deficiency can be manifested as diabetic ketoacidosis (DKA), lactic acidosis and hyperglycemia (130, 131). In another study, low blood

Table I. Genetic factors related to thiamine and cancer.

	Cancer	Thiamine
<i>SLC19</i> gene		
A1	<p>*<i>SLC19A1</i> Polymorphisms have been reported in many cancers, including osteosarcoma, colorectal cancer, non-small cell lung cancer, bladder cancer, prostate cancer, diffuse large B-cell lymphoma, pediatric acute lymphoblastic leukemia, head and neck cancer, central nervous system tumors in Thai children, and human ovarian carcinoma cells.</p>	<p>*Plays an important role in the transport and homeostasis of folate and thiamine in the body. *Transports mono- and pyrophosphate derivatives of thiamine.</p>
A2 (ThTr1)	<p>*<i>SLC19A3</i> RNA levels are down-regulated in breast and lung cancers compared with adjacent normal tissues.</p>	<p>*Mediates the transport of thiamine.</p>
A3 (ThTr2)	<p>*<i>SLC19A3</i> expression is down-regulated in gastric cancer cell lines (71%) and restored after pharmaceutical demethylation. Exogenous <i>SLC19A3</i> expression caused growth inhibition of gastric cancer cells. *<i>SLC19A3</i> gene expression is down-regulated in breast cancer and associated with resistance to apoptosis in these tumors. *Aberrant <i>SLC19A3</i> has been suggested as a novel biomarker for breast and gastric cancer diagnosis.</p>	<p>*Mediates the transport of thiamine.  *<i>SLC19A3</i> gene also exhibited evidence of regulation by exogenous thiamine in a manner concordant with the effects of <i>SLC19A3</i> overexpression.</p>
Transketolase (TK)	<p>*Aberrant TK and TK-like 1 (TKTL-1) expression has been reported to participate in glucose metabolism in malignant pleural effusion cells. TKTL-1 has been reported to play a pivotal role in carcinogenesis and has an important role in controlling the non-oxidative pentose-phosphate-pathway, which is significant in cancer. TKTL-1 expression has been correlated with poor patient outcome and tumor progression. *Overexpression of TKTL-1 has been observed in many cancers, including thyroid cancer, nasopharyngeal carcinoma, non-small cell lung cancer, gastric cancer, human hepatoma cell, colorectal cancer, urothelial cancer, anaplastic neuroblastoma, breast cancer, endometrial cancer, ovarian cancer, uterine cervix cancer, head and neck cancer, and glioblastoma.</p>	<p>*Thiamine regulates the expression of genes that code for enzymes that use thiamine as co-factor. Thiamine deficiency diminishes the mRNA levels of TK. In the blood of cancer patients, the concentration of thiamine and thiamine diphosphate (TDP) are decreased by 20%; Tk activity is decreased by 20% and the TDP effect is altered by 5-42%.</p>
<i>p53</i> gene	<p>*Plays a critical role in the regulation of the normal cell cycle, cell cycle arrest, and apoptotic response. *Transcription factor that has a major role in determining cell fates in response to DNA damage. *The function of <i>p53</i> is to mediate apoptosis in cells that have activated oncogenic pathways. *<i>p53</i> polymorphisms have been demonstrated in breast cancer, mesothelioma, esophageal squamous cell carcinoma, and pancreatic cancer. *In many meta-analyses, <i>p53</i> polymorphisms have been associated with cervical, gastric, bladder, colorectal, and endometrial cancer, but not with prostate cancer.</p>	<p>*Increased thiamine transporter activity has been reported in cells that overexpress the genes that code for thiamine transporters and under conditions of DNA damage or <i>p53</i> activation. *TDP has been shown to inhibit <i>p53</i> binding and thiamine has been shown to inhibit intracellular <i>p53</i> activity. *The expression of <i>p53</i> was decreased significantly in cultured retinal neurons of diabetic rats treated with thiamine.</p>
Poly(ADP-ribose) polymerase-1 (PARP 1)	<p>*PARP-1 is a nuclear protein that contributes to both cell death and survival under stressful conditions. The catalytic activity of PARP-1 is stimulated by DNA strand breaks. *Parp-1-deficient cells exhibit enhanced sensitivity to the lethal effects of ionizing radiation and alkylating agents, and <i>Parp-1</i> knockout mice developed spontaneous mammary and liver tumors. *Overexpression of PARP-1 has been reported in Ewing's sarcoma, malignant lymphomas, colorectal cancer, hepatocellular carcinoma, breast cancer, pediatric central nervous system tumors, and ovarian cancer.</p>	<p>*Thiamine has a cytoprotective effect on cultured neonatal rat cardiomyocytes under hypoxic insult. Thiamine also inhibits PARP cleavage and DNA fragmentation. *Benfotiamine, a fat-soluble thiamine analog, prevents bacterial endotoxin-induced inflammation and PARP cleavage in mouse macrophage cell lines. *Adenosine thiamine triphosphate, a thiamine derivative, has been identified in small amounts in the mouse brain, heart, skeletal muscle, liver and kidneys and has been shown to inhibit PARP-1 activity.</p>

Table I. Continued

Table I. *Continued*

	Cancer	Thiamine
	<p>*In a meta-analysis, <i>PARP-1</i> mRNA expression was correlated with high grade, medullary histological type, tumor size, and decreased metastasis-free survival and overall survival in human breast cancer.</p> <p>*<i>PARP-1</i> polymorphism has been associated with gastric cancer in the Han Chinese, prostate cancer, esophageal squamous cell carcinoma, and lung cancer, but it also associated with a reduced risk of non-Hodgkin's lymphoma in Korean males. <i>PARP 1</i> polymorphism reduces PARP-1 catalytic activity by 30-40%.</p> <p>*In a meta-analysis study, no significant association was observed between the <i>PARP-1 V762</i> polymorphism and cancer risk; however, the variant A allele of the <i>PARP-1 V762</i> polymorphism was associated with an increased risk of cancer among Asian populations but a decreased risk of cancer among Caucasians, particular for glioma.</p>	
<p>The reduced form of the nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (NOX) enzyme complex</p>	<p>*Mediates critical physiological and pathological processes including cell signaling, inflammation and mitogenesis through the generation of reactive oxygen species from molecular oxygen.</p> <p>*NOX1 is required for RAS oncogene-induced cell transformation.</p> <p>*The NOX subunit p22<sup>phox</sup> has been reported to inhibit the function of the tumor suppressor protein tuberlin in renal carcinoma cells.</p> <p>*The activation of NOX has been demonstrated in the development of several types of cancer, including melanoma, leukemia, esophageal, hepatocellular carcinoma, prostate, colon, glioblastoma multiforme, and multiple myeloma.</p> <p>*<i>NADPH</i> polymorphisms have been associated with myelodysplastic syndrome and <i>de novo</i> acute myeloid leukemia, esophageal cancer, lung cancer, non-Hodgkin's lymphoma, childhood acute leukemia, postmenopausal breast cancer, and gastric cancer.</p>	<p>*The levels of NADPH cytochrome <i>c</i> reductase are increased in thiamine-deficient (TD) animals.</p> <p>*The expression of NOX4 is significantly down-regulated by benfotiamine treatment under both normo- and hyperglycemic conditions.</p> <p>*Animals fed a high thiamine diet exhibited an NADPH-cytochrome <i>c</i> reductase activity that was approximately 57% of those fed a TD diet.</p>

thiamine levels in the blood and erythrocyte transketolase activity, and high erythrocyte TPP activity were documented in patients with diabetes (132, 133). The activity of transketolase has been used to assess thiamine activity in mammalian tissues. The low thiamine values in patients with diabetes might be a reflection of reduced apo-enzyme levels from the disease itself rather than from a thiamine deficiency (134). In addition, plasma thiamine levels are decreased by 76% in type 1 and 75% in type 2 DM; decreased plasma thiamine levels have also been associated with increased renal clearance and fractional excretion of thiamine (134). Furthermore, thiamine transporter protein concentrations are increased in erythrocyte membranes in type 1 and type 2 DM. Therefore, changes in thiamine levels may be masked by an increase in thiamine transporter expression (134).

*Cancer.* Thiamine levels are decreased in many cancer subjects. Hepatoma induced by butter-yellow exhibited a decrease in thiamine and co-enzyme A concentrations by more than 50% and 92%, respectively, compared to normal livers (135). Thiamine deficiencies have also been noted in patients with uterine cancer (136), B-chronic lymphocytic leukemia (137), and colorectal cancer (138). During the growth of Ehrlich's ascites tumor, the concentration of TDP in the liver of tumor-bearing animals continuously drops for 10 days after inoculation, and the TDP level in the tumor itself decreases more abruptly by the terminal period of tumor growth (10th day) (139). Some authors have suggested that thiamine supplementation may contribute to the high rate of tumor cell survival, proliferation and chemotherapy resistance (140). A significantly higher incidence of urinary bladder carcinomas was induced in rats fed bracken fern,

which contains a carcinogen agent, and additionally supplemented with thiamine than observed in rats fed only bracken fern (141). In melanoma cells, overexpression of the melanoma tumor antigen p97 resulted in decreased thiamine triphosphatase levels that may lead to greater TPP and increased cellular proliferation (142). TPP inhibits cytotoxicity of methotrexate, an antimetabolite used for the treatment of cancer (143). *In vitro*, thiamine blocks the antitumor effect of mechlorethamine in murine leukemia cells (144) and blocks the antitumor effect of cyclophosphan in Ehrlich's ascites tumor (145). The administration of cyclophosphan further increased thiamine deficiency in tumor-bearing mice. Simultaneous administration of thiamine and cyclophosphan abolished the cytostatic toxic effect but did not affect their antitumoral properties (145). Furthermore, low thiamine levels were of benefit against some types of cancers. Thiaminase I digests thiamine and has been reported to have a growth-inhibitory activity against breast cancer cell lines (146) and lymphoid leukemia cell lines (147). In addition, MDA231 breast cancer xenografts exhibited growth delay in mice fed a thiamine-free diet (146). Injections of oxythiamine increased the rate of pathological mitoses in Walker 256 carcinoma cells, reduced the tumor weight by 45%, and inhibited the synthesis of TDP and activity of transketolase in tissues (148). However, marginal dietary thiamine deficiency induces the formation of aberrant colonic crypt foci in rats (149). In Syrian hamsters, which are susceptible to oral tumor induction by chemical agents, a dietary regime low in thiamine resulted in the development of malignant neoplasms arising in the pouch epithelium in a significantly shorter period of time than in hamsters fed a diet containing adequate amounts of this vitamin (150). Moreover, thiamine did not induce death or increase the proliferation rate of Hepa or HepG2 HCC cell lines, either *in vitro* or *in vivo*, and thiamine was reported to attenuate Wilson's disease-induced HCC (151). In another report, thiamine did not stimulate tumor growth in patients with gastric tumors (145). In a patient with osteosarcoma, subcutaneous perfusion of thiamine reduced the tumor circumference from 30 to 20 cm, equivalent to a reduction of 50-75% in volume, within two days (152). A metabolic control analysis demonstrated a high stimulatory effect on tumor growth of 164% compared with controls with a thiamine dose of 25-fold the recommended dietary allowance (RDA); however, at very high doses of thiamine, ~2500-fold the RDA, the opposite effect was observed, producing an inhibitory effect on tumor growth of 36% compared with control animals (153). The Authors suggested that when thiamine supplementation is necessary for patients with cancer, it should be administered at high doses to avoid the tumor-promoting effect of low doses. Furthermore, thiamine analog benfotiamine significantly reduced genomic damage of peripheral lymphocytes in hemodialysis patients, who

exhibited enhanced DNA damage in end-stage renal disease, independent of changes in the plasma levels of advanced glycation end product (154).

*Matrix metalloproteinases (MMPs)*. MMPs are proteolytic enzymes responsible for both extracellular matrix (ECM) remodeling and the regulation of leukocyte migration through the ECM, which is an important step in the inflammatory processes. MMP overexpression has been reported in many types of cancer, including malignant peripheral nerve sheath tumors (155), non-small cell lung cancer (156), glioblastoma multiforme (157), sarcoma (158), colorectal cancer (159), prostate cancer (160), and ovarian carcinoma (161). *MMP* polymorphisms are associated with breast cancer (162), colorectal cancer (163), renal cell carcinoma (164), adult astrocytoma (165), endometrial carcinoma (166), bladder cancer (167), epithelial ovarian cancer (168), oral cancer (169), cervical cancer (170), esophageal squamous cell carcinoma (171), gastric cancer (172), malignant head and neck tumors (173), hypophyseal adenoma (174), prostate cancer (175), glioblastoma multiforme (176), and HCC (177). In meta-analyses, *MMP-2* polymorphisms have been linked to the risk of lung (-735 C/T) (178) and breast cancer (-1306 C/T) (179), but *MMP-2* -1306 C/T allele may be a protective factor for digestive cancer risk (180). In other studies, polymorphisms of the *MMP-1*, -2, 3, and -7 may have played allele-specific roles in cancer development (181, 182). However, *MMP-9* has also been shown to be up-regulated in the TD mouse brain (183, 184). Thiamine reduces the *MMP-2* activity in the heart of diabetic rats and prevents diabetes-induced cardiac fibrosis (185). Moreover, thiamine and benfotiamine correct the increase in *MMP-2* activity that results from high glucose levels in human retinal pericytes, while increasing tissue inhibitor of metalloproteinase-1 (TIMP-1) (186). Altogether, these studies suggest that thiamine may play an important role in the pathological processes of cancer by modulating the levels of MMPs and TIMPs.

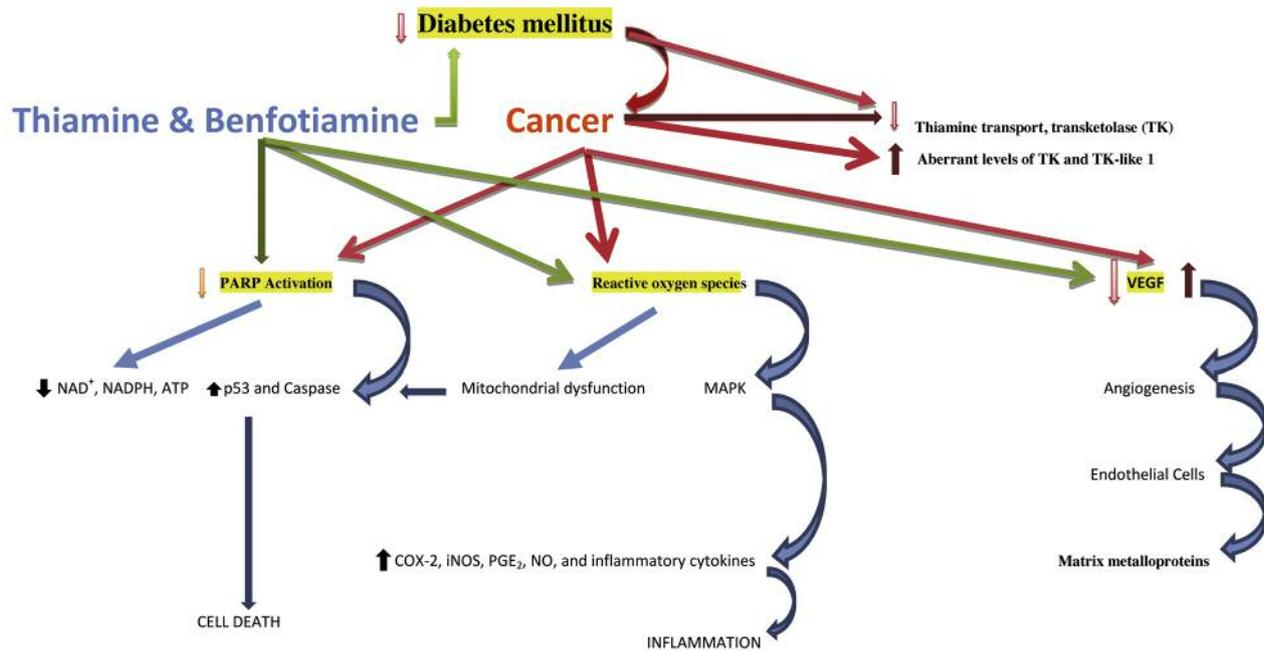
*Prostaglandins (PGs) and Cyclooxygenase-2 (COX2)*. PGS play a role in inflammatory processes. COX participates in the conversion of arachidonic acid into PGs. The expression of COX2 mRNA and PGE<sub>2</sub> were selectively increased in vulnerable regions in the symptomatic stages of TD encephalopathy animal models (187). Up-regulation of 15-hydroxyprostaglandin dehydrogenase (15-PGDH) expression was observed in breast cancer cell lines transfected with *SLC19A3* gene and down-regulation was observed after suppression of *SLC19A3* with siRNA vectors (188). Overexpression of 15-PGDH inhibited interleukin (IL-1 $\beta$ )-induced COX2 expression (189). In murine macrophages, benfotiamine also blocked the expression of COX2 and its product PGE<sub>2</sub> by lipopolysaccharide (LPS)-induced cytotoxicity (190). In addition, benfotiamine significantly

prevented LPS-induced macrophage death and monocyte adhesion to endothelial cells. These anti-inflammatory effects of benfotiamine are mediated through the regulation of the arachidonic acid pathway in macrophages (191). Tumor inflammation is now recognized as one of the hallmarks of cancer. Over-expression of COX2 has been associated with resistance to apoptosis, increased angiogenesis and increased tumor invasiveness in various types of cancer. Increased COX2 expression has been reported in endometrial adenocarcinoma (192), breast cancer (193), reno-medullary interstitial cell tumor (194), colorectal carcinomas (195), gastric cancer (196), carcinoma of the cervix (197), and familial adenomatous polyposis (198). Deletion of *Cox2* in mouse mammary epithelial cells delayed breast cancer onset (199). COX-2 inhibitors also reduced the growth and induced regression of human esophageal adenocarcinoma xenografts in nude mice (200), and retarded murine mammary tumor progression by reducing tumor cell migration, invasiveness, and angiogenesis (201). Genetic variability in enzymes could impact on the risk of the disease. COX2 polymorphisms have been associated with bladder (202), biliary tract (203), lung (204), non-melanoma skin cancer after organ transplantation (205), esophageal squamous cell carcinoma (206), nasopharyngeal carcinoma (207), pancreatic (208), invasive ovarian carcinoma (209), breast (210), gastric carcinoma (211), acute myeloid leukemia (212), prostate (213), head and neck (214), colorectal adenoma (215), and HCC (216). In meta-analyses, the COX2 1195G>A polymorphism was significantly associated with an increased risk of digestive system cancer, especially in Asian populations (217), and the -765G>C variant may confer an increased risk of colorectal carcinoma and esophageal cancer in Asians. The 8473T>C polymorphism may confer a decreased risk of breast and lung cancer (218). In addition, PGE<sub>2</sub> has been reported to be associated with colorectal adenoma (219), pancreatic tumor (220), and childhood neuroblastoma (221). The suppression of PGE<sub>2</sub> receptor inhibited human lung carcinoma cell growth (222). These findings suggest that thiamine may play a role in modulating the inflammatory process of cancer.

**Reactive oxygen species (ROS).** ROS play a major role in various cell-signaling pathways. ROS activates various transcription factors and increases in the expression of proteins that control cellular transformation, tumor cell survival, tumor cell proliferation and invasion, angiogenesis, and metastasis (223). ROS also play an important role in the initiation and progression of many types of cancer (224-229). Single-nucleotide polymorphisms of antioxidant defense genes may significantly modify the functional activity of the encoded proteins. Women with genetic variability in iron-related oxidative stress pathways may be at increased risk for postmenopausal breast cancer (230). The *Ala* variant of *superoxide dismutase* (SOD) has been associated with a moderately increased risk of prostate cancer (231). In meta-

analyses, *manganese* SOD polymorphisms may contribute to cancer development (*Val-9Ala*) (234) and prostate cancer susceptibility (*Val-16Ala*) (232) but not breast cancer susceptibility (*Val-16Ala*) (233). However, oxidative stress has been associated with region-specific neuronal death, and lipid peroxidation products accumulated in the remaining thalamic neurons after 11 days of TD animal models (234). *In vitro*, thiamine inhibits lipid peroxidation in rat liver microsomes and free radical oxidation of oleic acid (235). Benfotiamine promotes a reduction in ROS-induced by advanced glycosylated albumin in macrophages (236). In primary human peritoneal mesothelial cells of a rat model of peritoneal dialysis, the addition of benfotiamine enhanced transketolase activity and reduced expression of advanced glycation end products and their receptor (237). These data suggest that benfotiamine protects the peritoneal membrane and remnant kidney in such a rat model. Thiamin rescues hepatocytes from iron-catalyzed oxidative stress by reducing lipid peroxidation, mitochondrial and protein damage and DNA oxidation (238). These findings suggest that thiamine modulates oxidative stress in cancer.

**Nitric oxide synthase (NOS).** NOS is an enzyme involved in the synthesis of nitric oxide (NO), which regulates a variety of important physiological responses, including cell migration, immune response and apoptosis. NO and calcium have been reported to regulate mitochondrial biogenesis in follicular thyroid carcinoma cells (239). There is a link between NO and the induction of apoptotic cell death in head and neck squamous cell carcinoma development (240). Cytokines especially interferon-gamma was found to induce apoptosis in acute leukemia *via* the NO and caspase-3 pathway (241). The reduction of NO enhanced the radiosensitivity of hypoxic non-small cell lung cancer (242). Increased NO may be a sign of subclinical cardiotoxicity of doxorubicin (243). High NO concentrations at the periphery of a melanoma may contribute to metastasis by stimulating cell proliferation, inhibiting apoptosis, or acting as a lymphangiogenic factor (244). Inducible NOS (*iNOS*) mRNA expression was considerably higher in glioblastoma than meningioma specimens (245). *iNOS* expression has been correlated with angiogenesis, lymphangiogenesis, and poor prognosis in gastric cancer (246) and in estrogen receptor-negative breast cancer (247). NOS inhibition enhanced the antitumor effect of radiation in the treatment of squamous carcinoma xenografts (248). NOS polymorphisms have been associated with bladder (249), urothelial carcinoma (250), gastric (251), colorectal cancer (252), and non-Hodgkin's lymphoma (253). In a meta-analysis study, the endothelial NOS 894G>T polymorphism was associated with breast cancer (254). However, increased brain endothelial NOS expression was demonstrated in TD (255). In murine macrophages, benfotiamine was also found to block the expression of *iNOS* by LPS-induced cytotoxicity (189).



Red Color: Activated, increased.  
 Solid Pink Color: Attenuated, reduced.

Figure 1. Summary the role of thiamine in cancer.

Table II. The role of thiamine in cancer.

	Cancer	Thiamine
Diabetes mellitus	*Increased risk of cancer.	*Low plasma thiamine levels.
Cancer		*Low plasma thiamine levels.
Matrix metalloproteinases (MMP)	*Overexpression of MMPs is observed in cancer *MMP polymorphisms have been associated with cancer.	*Thiamine and benfotiamine reduce MMP levels.
Prostaglandins (PGs) and Cyclooxygenase 2 (COX2)	*Overexpression of COX2 and PGE <sub>2</sub> has been reported in cancer. *COX2 polymorphisms have been associated with cancer.	*Benfotiamine inhibits COX2 and PGE <sub>2</sub> expression.
Reactive oxygen species (ROS)	*ROS have been suggested to play a role in cancer.	*Thiamine reduces ROS.
Nitric oxide synthase (NOS)	*iNOS expression increases in cancer. *NOS polymorphisms have been associated with cancer.	*Benfotiamine inhibits iNOS expression.

Table II illustrates the role of thiamine in cancer and Figure 1 summarizes the role of thiamine in cancer.

### Conclusion

Genetic studies have provided opportunities to determine which proteins may link thiamine to cancer pathology, including *SLC19* gene, transketolase, transcription factor p53, PARP, and NOX. Thiamine can also act through a number of

non-genomic mechanisms, which include protein expression, oxidative stress, inflammation, and cellular metabolism. Thiamine supplementation may stimulate a high rate of tumor cell survival, proliferation and chemotherapy resistance, but other studies have demonstrated the beneficial role of thiamine in cancer. However, thiamine deficiency may occur in patients with cancer and may produce serious disorders, including Wernicke's encephalopathy, that require parenteral thiamine supplementation. In the early stages of TD

encephalopathy (Wernicke's encephalopathy), patients were found to respond rapidly to large doses of parenteral thiamine. The initial dose of thiamine is usually 100 mg two to three times daily for one to two weeks. A very high dose of thiamine has been shown to exhibit a growth inhibitory effect in cancer. Thiamine appears to have both cancer-promoting and anticancer properties, which vary in relation to both genomic and non-genomic factors. However, thiamine absorption decreases with advancing age (256). Therefore, further investigations on parenteral thiamine in cancer are needed. Indeed, a more cautious approach would be advisable before recommending the widespread use of thiamine in patients with cancer.

### Ethical Approval

Not required.

### Funding

The Authors declare that they received no funding.

### Conflicts of Interest

Authors declare that they have no competing interests.

### References

- Brinkman MT, Karagas MR, Zen MS, Schned A, Reulen RC and Zeegers MP: Minerals and vitamins and the risk of bladder cancer: results from the New Hampshire study. *Cancer Causes Control* 21: 609-619, 2010.
- Kaaks R, Tuyns AJ, Haelterman M and Riboli E: Nutrient intake patterns and gastric cancer risk: a case-control study in Belgium. *Int J Cancer* 78: 415-420, 1998.
- Hernandez BY, McDuffie K, Wilkens LR, Kamemoto L and Goodman MT: Diet and premalignant lesions of the cervix: evidence of a protective role for folate, riboflavin, thiamin, and vitamin B12. *Cancer Causes Control* 14: 859-470, 2003.
- Jedrychowski W, Steindorf K, Popiela T, Wahrendorf J, Tobiasz-Adamczyk B, Kulig J and Penar A: Alcohol consumption and the risk of colorectal cancer at low levels of micronutrient intake. *Med Sci Monit* 8: CR357-363, 2002.
- Du S, Shi L, Zhang H and He S: Relationship between dietary nutrients intakes and human prostate cancer. *Wei Sheng Yan Jiu* 26: 122-125, 1997. (Chinese).
- Hargreaves MK, Baquet C and Gamshadzahi A: Diet, nutritional status, and cancer risk in American blacks. *Nutr Cancer* 12: 1-28, 1989.
- Georgiannos SN, Weston PMT and Goode AW: Micronutrients in gastrointestinal cancer. *Br J Cancer* 68: 1195-1198, 1993.
- Nesterova VI and Chebotareva MA: Thiamine content and enzyme activity in blood cells in leukemia]. *Vopr Med Khim* 22: 732-735, 1976. (in Russian).
- Basu TK and Dickerson JW: The thiamin status of early cancer patients with particular reference to those with breast and bronchial carcinomas. *Oncology* 33: 250-252, 1976.
- Boros LG, Puigjaner J, Cascante M, Lee WN, Brandes JL, Bassilian S, Yusuf FI, Williams RD, Muscarella P, Melvin WS and Schirmer WJ: Oxythiamine and dehydroepiandrosterone inhibit the nonoxidative synthesis of ribose and tumor cell proliferation. *Cancer Res* 57: 4242-4248, 1997.
- Raïs B, Comin B, Puigjaner J, Brandes JL, Creppy E, Saboureau D, Ennamany R, Lee WN, Boros LG and Cascante M: Oxythiamine and dehydroepiandrosterone induce a G1 phase cycle arrest in Ehrlich's tumor cells through inhibition of the pentose cycle. *FEBS Lett* 456: 113-118, 1999.
- Crim JA, Buskirk HH and Petering HG: The effect of B vitamin and protein intake of the rat on the antitumor activity of 3-ethoxy-2-oxobutyraldehyde bis(thiosemicarbazone). *Cancer Res* 27: 1109-1114, 1967.
- Ganapathy V, Smith SB and Prasad PD: SLC19: the folate/thiamine transporter family. *Pflugers Arch* 447: 641-646, 2004.
- Dutta B, Huang W, Molero M, Kekuda R, Leibach FH, Devoe LD, Ganapathy V and Prasad PD: Cloning of the human thiamine transporter, a member of the folate transporter family. *J Biol Chem* 274: 31925-31929, 1999.
- Rajgopal A, Edmondson A, Goldman ID and Zhao R: SLA19A3 encodes a second thiamine transporter ThTr2. *Biochim Biophys Acta* 1537: 175-178, 2001.
- Liu S, Huang H, Lu X, Golinski M, Comesse S, Watt D, Grossman RB and Moscow JA: Down-regulation of thiamine transporter THTR2 gene expression in breast cancer and its association with resistance to apoptosis. *Mol Cancer Res* 1: 665-1673, 2003.
- Liu S, Stromberg A, Tai H-H and Moscow JA: Thiamine transporter gene expression and exogenous thiamine modulate the expression of genes involved in drug and prostaglandin metabolism in breast cancer cells. *Mol Cancer Res* 2: 477-487, 2004.
- Liu X, Lam EK, Wang X, Zhang J, Cheng YY, Lam YW, Ng EK, Yu J, Chan FK, Jin H and Sung JJ: Promoter hypermethylation mediates downregulation of thiamine receptor SLC19A3 in gastric cancer. *Tumour Biol* 30: 242-248, 2009.
- Ikehata M, Ueda K and Iwakawa S: Different involvement of DNA methylation and histone deacetylation in the expression of solute-carrier transporters in 4 colon cancer cell lines. *Biol Pharm Bull* 35: 301-307, 2012.
- Sweet R, Paul A and Zastre J: Hypoxia induced upregulation and function of the thiamine transporter, SLC19A3 in a breast cancer cell line. *Cancer Biol Ther* 10: 1101-1111, 2010.
- Ng EK, Leung CP, Shin VY, Wong CL, Ma ES, Jin HC, Chu KM and Kwong A: Quantitative analysis and diagnostic significance of methylated SLC19A3 DNA in the plasma of breast and gastric cancer patients. *PLoS ONE* 6: e22233, 2011.
- Zhao R, Gao F, Wang Y, Diaz GA, Gelb BD and Goldman ID: Impact of the reduced folate carrier on the accumulation of active thiamin metabolites in murine leukemia cells. *J Biol Chem* 276: 1114-1118, 2001.
- Zhao R, Gao F and Goldman ID: Reduced folate carrier transport thiamine monophosphate: an alternative route for thiamine delivery into mammalian cells. *Am J Physiol* 282: C1512-1517, 2002.
- Yang R, Qin J, Hoang BH, Healy JH and Gorlik R: Polymorphisms and methylation of the reduced folate carrier in osteosarcoma. *Clin Orthop Relat Res* 466: 2046-2051, 2008.

- 25 Figueiredo JC, Levine AJ, Lee WH, Conti DV, Poynter JN, Campbell PT, Duggan D, Lewinger JP, Martinez ME, Ulrich CM, Newcomb P, Potter J, Limburg PJ, Hopper J, Jenkins MA, Le Marchand L, Baron JA and Haile RW: Genes involved with folate uptake and distribution and their association with colorectal cancer risk. *Cancer Causes Control* 21: 597-608, 2010.
- 26 Adjei AA, Salavaggione OE, Mandrekar SJ, Dy GK, Ziegler KL, Endo C, Molina JR, Schild SE and Adjei AA: Correlation between polymorphisms of the reduced folate carrier gene (SLC19A1) and survival after premetrexed-based therapy in non-small cell lung cancer: a North central cancer treatment group-based exploratory study. *J Thorac Oncol* 5: 1346-1353, 2010.
- 27 Abdel-Haleem AM, El-Zeiry MI, Mahran LG, Abou-Aisha K, Rady MH, Rohde J, Mostageer M and Spahn-Langguth H: Expression of RFC/SLC19A1 is associated with tumor type in bladder cancer patients. *PLoS ONE* 6: e21820, 2011.
- 28 Collin SM, Metcalfe C, Zuccolo L, Lewis SJ, Chen L, Cox A, Davis M, Lane JA, Donovan J, Smith GD, Neal DE, Hamdy FC, Gudmundsson J, Sulem P, Rafnar T, Benediksdottir KR, Eeles RA, Guy M, Kote-Jarai Z: UK Genetic Prostate Cancer Study Group, Morrison J, Al Olama AA, Stefansson K, Easton DF and Martin RM: Association of folate-pathway gene polymorphisms with the risk of prostate cancer: a population-based nested case-control study, systemic review, and meta-analysis. *Cancer Epidemiol Biomarkers Prev* 18: 2528-2539, 2009.
- 29 Kastrup IB, Worm J, Ralfkiaer E, Guldborg P and Grønbaek K: Genetic and epigenetic alterations of the reduced folate carrier in untreated diffuse large B-cell lymphoma. *Eur J Haematol* 80: 61-66, 2008.
- 30 de Jonge R, Tissing WJ, Hooijberg JH, Jansen G, Kaspers GJ, Lindemans J, Peters GJ and Pieters R: Polymorphisms in folate-related genes and risk of pediatric acute lymphoblastic leukemia. *Blood* 113: 2284-2289, 2009.
- 31 Galbiatti AL, Ruiz MT, Rezende Pinto D, Raposo LS, Maniglia JV, Pavarino-Bertelli EC and Goloni-Bertollo EM: A80G polymorphism of reduced folate carrier 1 (RFC1) gene and head and neck squamous cell carcinoma etiology in Brazilian population. *Mol Biol Rep* 38: 1071-1078, 2011.
- 32 Sirachainan N, Wongruangrsi S, Kajanachumpol S, Pakakasama S, Visudtibhan A, Nuchprayoon I, Lusawat A, Phudhichareerat S, Shuangshoti S and Hongeng S: Folate pathway genetic polymorphisms and susceptibility of central nervous system tumors in Thai children. *Cancer Detect Prev* 32: 72-78, 2008.
- 33 Corona G, Giannini F, Fabris M, Toffoli G and Boiocchi M: Role of folate receptor and reduced folate carrier in the transport of 5-methyltetrahydrofolic acid in human ovarian carcinoma cells. *Int J Cancer* 75: 125-133, 1998.
- 34 Meléndez RR: Importance of water-soluble vitamins as regulatory factors of genetic expression. *Rev Invest Clin* 54: 77-83, 2002. (in Spanish).
- 35 Trebukhina RV, Tumanov VN, Avdeichik LA, Bobko IN and Tsabenko II: Transketolase activity and thiamine diphosphate content in oncological patients. *Vopr Med Khim* 29: 85-90, 1983. (in Russian).
- 36 Lin CC, Chen LC, Tseng VS, Yan JJ, Lai WW, Su WP, Lin CH, Huang CY and Su WC: Malignant pleural effusion cells aberrant glucose metabolism gene expression. *Eur Respir J* 37: 1453-1465, 2011.
- 37 Xu X, Zur Hausen A, Coy JF and Löchelt M: Transketolase-like protein 1 (TKTL1) is required for rapid cell growth and full viability of human tumor cells. *Int J Cancer* 124: 1330-1337, 2009.
- 38 Zerilli M, Amato MC, Martorana A, Cabibi D, Coy JF, Cappello F, Pompei G, Russo A, Giordano C and Rodolico V: Increased expression of transketolase-like-1 in papillary thyroid carcinomas smaller than 1.5 cm in diameter is associated with lymph-node metastases. *Cancer* 113: 936-944, 2008.
- 39 Fröhlich E, Fink I and Wahl R: Is transketolase like 1 a target for the treatment of differentiated thyroid carcinoma? A study on thyroid cancer cell lines. *Invest New Drugs* 27: 297-303, 2009.
- 40 Völker HU, Scheich M, Schmausser B, Kämmerer U and Eck M: Overexpression of transketolase TKTL1 is associated with shorter survival in laryngeal squamous cell carcinomas. *Eur Arch Otorhinolaryngol* 264: 1431-1436, 2007.
- 41 Zhang S, Yue JX, Yang JH, Cai PC and Kong WJ: Overexpression of transketolase protein TKTL1 is associated with occurrence and progression in nasopharyngeal carcinoma: a potential therapeutic target in nasopharyngeal carcinoma. *Cancer Biol Ther* 7: 517-522, 2008.
- 42 Kayser G, Sielen W, Kubitz B, Mattern D, Stickeler E, Passlick B, Werner M and Zur Hausen A: Poor outcome in primary non-small cell lung cancers is predicted by transketolase TKTL1 expression. *Pathology* 43: 719-724, 2011.
- 43 Schultz H, Kähler D, Branscheid D, Vollmer E, Zabel P and Goldmann T: TKTL1 is overexpressed in a large portion of non-small cell lung cancer specimens. *Diagn Pathol* 3: 35, 2008.
- 44 Staiger WI, Coy JF, Grobholz R, Hofheinz RD, Lukan N, Post S, Schwarzbach MH and Willeke F: Expression of the mutated transketolase TKTL1, a molecular marker in gastric cancer. *Oncol Rep* 16: 657-661, 2006.
- 45 Zhang S, Yang JH, Guo CK and Cai PC: Gene silencing of TKTL1 by RNAi inhibits cell proliferation in human hepatoma cells. *Cancer Lett* 253: 108-114, 2007.
- 46 Diaz-Moralli S, Tarrado-Castellarnau M, Alenda C, Castells A and Cascante M: Transketolase-like 1 expression is modulated during colorectal cancer progression and metastasis formation. *PLoS One* 6: e25323, 2011.
- 47 Langbein S, Zerilli M, Zur Hausen A, Staiger W, Rensch-Boschert K, Lukan N, Popa J, Ternullo MP, Steidler A, Weiss C, Grobholz R, Willeke F, Alken P, Stassi G, Schubert P and Coy JF: Expression of transketolase TKTL1 predicts colon and urothelial cancer patient survival: Warburg effect reinterpreted. *Br J Cancer* 94: 578-585, 2006.
- 48 Wu HT, Allie N, Myer L and Govender D: Anaplastic nephroblastomas express transketolase-like enzyme 1. *J Clin Pathol* 62: 460-463, 2009.
- 49 Földi M, Stickeler E, Bau L, Kretz O, Watermann D, Gitsch G, Kayser G, Zur Hausen A and Coy JF: Transketolase protein TKTL1 overexpression: A potential biomarker and therapeutic target in breast cancer. *Oncol Rep* 17: 841-845, 2007.
- 50 Krockenberger M, Engel JB, Schmidt M, Kohrenhagen N, Häusler SF, Dombrowski Y and Kapp M, Diel J, Honig A: Expression of transketolase-like 1 protein (TKTL1) in human endometrial cancer. *Anticancer Res* 30: 1653-1659, 2010.
- 51 Krockenberger M, Honig A, Rieger L, Coy JF, Sutterlin M, Kapp M, Horn E, Diel J and Kammerer U: Transketolase-like 1 expression correlates with subtypes of ovarian cancer and the presence of distant metastases. *Int J Gynecol Cancer* 17: 101-106, 2007.

- 52 Schmidt M, Kammerer U, Segerer S, Cramer A, Kohrenhagen N, Dietsl J and Voelker HU: Glucose metabolism and angiogenesis in granulosa cell tumors of the ovary: activation of Akt, expression of M2PK, TKTL1 and VEGF. *Eur J Obstet Gynecol Reprod Biol* 139: 72-78, 2008.
- 53 Chen H, Yue JX, Yang SH, Ding H, Zhao RW and Zhang S: Overexpression of transketolase-like gene 1 is associated with cell proliferation in uterine cervix cancer. *J Exp Clin Cancer Res* 28: 43, 2009.
- 54 Sun W, Liu Y, Glazer CA, Shao C, Bhan S, Demokan S, Zhao M, Rudek MA, Ha PK and Califano JA: TKTL1 is activated by promoter hypomethylation and contributes to head and neck squamous cell carcinoma carcinogenesis through increased aerobic glycolysis and HIF1 $\alpha$  stabilization. *Clin Cancer Res* 16: 857-866, 2010.
- 55 Völker HU, Hagemann C, Coy J, Wittig R, Sommer S, Stojic J, Haubitz I, Vince GH and Kämmerer U, Monoranu CM: Expression of transketolase-like 1 and activation of Akt in grade IV glioblastomas compared with grades II and III astrocytic gliomas. *Am J Clin Pathol* 130: 50-57, 2008.
- 56 Mitschke L, Parthier C, Schröder-Tittmann K, Coy J, Lüdtko S and Tittmann K: The crystal structure of human transketolase and new insights into its mode of action. *J Biol Chem* 285: 31559-31570, 2010.
- 57 Lo Nigro C, Vivenza D, Monteverde M, Lattanzio L, Gojjs O, Garrone O, Comino A, Merlano M, Quinlan PR, Syed N, Purdie CA, Thompson A, Palmieri C and Crook T: High frequency of complex TP53 mutations in CNS metastases from breast cancer. *Br J Cancer* 106: 397-404, 2012.
- 58 Li Q, Kawamura K, Yamanaka M, Okamoto S, Yang S, Yamauchi S, Fukamachi T, Kobayashi H, Tada Y, Takiguchi Y, Tatsumi K, Shimada H, Hiroshima K and Tagawa M: Upregulated p53 expression activates apoptotic pathways in wild-type p53-bearing mesothelioma and enhances cytotoxicity of cisplatin and pemetrexed. *Cancer Gene Ther* 19: 218-228, 2012.
- 59 Abedi-Ardekani B, Kamangar F, Sotoudeh M, Villar S, Islami F, Aghcheli K, Nasrollahzadeh D, Taghavi N, Dawsey SM, Abnet CC, Hewitt SM, Fahimi S, Saïdi F, Brennan P, Boffetta P, Malekzadeh R and Hainaut P: Extremely high tp53 mutation load in esophageal squamous cell carcinoma in goleslan province, Iran. *PLoS One* 6: e29488, 2011.
- 60 King JC, Lu QY, Li G, Moro A, Takahashi H, Chen M, Go VL, Reber HA, Eibl G and Hines OJ: Evidence for activation of mutated p53 by apigenin in human pancreatic cancer. *Biochim Biophys Acta* 1823: 593-604, 2012.
- 61 Koushik A, Platt RW and Franco EL: p53 codon 72 polymorphism and cervical neoplasia: a meta-analysis review. *Cancer Epidemiol Biomarkers Prev* 13: 11-22, 2004.
- 62 Zhou Y, Li N, Zhuang W, Liu GJ, Wu TX, Yao X, Du L, Wei ML and Wu XT: P53 codon 72 polymorphism and gastric cancer: a meta-analysis of the literature. *Int J Cancer* 121: 1481-1486, 2007.
- 63 Li DB, Wei X, Jiang LH, Wang Yand Xu F: Meta-analysis of epidemiological studies of association of P53 codon 72 polymorphism with bladder cancer. *Genet Mol Res* 9: 1599-1605, 2010.
- 64 Dahabreh IJ, Linardou H, Bouzika P, Varvarigou V and Murray S: TP53 Arg72Pro polymorphism and colorectal cancer risk: a systematic review and meta-analysis. *Cancer Epidemiol Biomarkers Prev* 19: 1840-1847, 2010.
- 65 Gu Y, Zhou X and Zhang SL: Meta-analysis of an association of codon 72 polymorphisms of the p53 gene with increased endometrial cancer risk. *Genet Mol Res* 10: 3609-3619, 2011.
- 66 Li MS, Liu JL, Wu Y, Wang P and Teng H: Meta-analysis demonstrates no association between p53 codon 72 polymorphism and prostate cancer risk. *Genet Mol Res* 10: 2924-2933, 2011.
- 67 Lo PK, Chen JY, Tang PP, Lin J, Lin CH, Su LT, Wu CH, Chen TL, Yang Y and Wang FF: Identification of a mouse thiamine transporter gene as a direct transcriptional target for p53. *J Biol Chem* 276: 37186-37193, 2001.
- 68 McLure KG, Takagi M and Kastan MB: NAD<sup>+</sup> modulates p53 DNA binding specificity and function. *Mol Cellular Biol* 24: 9958-9967, 2004.
- 69 Yang Z, Ge J, Yin W, Shen H, Liu H and Guo Y: The expression of p53, MDM2 and Ref1 gene in cultured retina neurons of SD rats treated with vitamin B1 and/or elevated pressure. *Yan Ke Xue Bao* 20: 259-263, 2004. (in Chinese).
- 70 Masutani M, Nakagama H and Sugimura T: Poly(ADP-ribose) and carcinogenesis. *Genes Chromosomes Cancer* 38: 339-348, 2003.
- 71 Tong WM, Yang YG, Cao WH, Galendo D, Frappart L, Shen Y and Wang ZQ: Poly(ADP-ribose) polymerase-1 plays a role in suppressing mammary tumorigenesis in mice. *Oncogene* 26: 3857-3867, 2007.
- 72 Tong WM, Cortes U, Hande MP, Ohgaki H, Cavalli LR, Lansdorp PM, Haddad BR and Wang ZQ: Synergistic role of Ku80 and poly(ADP-ribose) polymerase in suppressing chromosomal aberrations and liver cancer formation. *Cancer Res* 62: 6990-6996, 2002.
- 73 Prasad SC, Thraves PJ, Bhatia KG, Smulson ME and Dritschilo A: Enhanced poly(adenosine diphosphate ribose) polymerase activity and gene expression in Ewing's sarcoma cells. *Cancer Res* 50: 38-43, 1990.
- 74 Tomoda T, Kurashige T, Moriki T, Yamamoto H, Fujimoto S and Taniguchi T: Enhanced expression of poly(ADP-ribose) synthetase gene in malignant lymphoma. *Am J Hematol* 37: 223-227, 1991.
- 75 Noshio K, Yamamoto H, Mikami M, Taniguchi H, Takahashi T, Adachi Y, Imamura A, Imai K and Shinomura Y: Overexpression of poly(ADP-ribose) polymerase-1 (PARP-1) in the early stage of colorectal carcinogenesis. *Eur J Cancer* 42: 2374-2381, 2006.
- 76 Quiles-Perez R, Muñoz-Gómez JA, Ruiz-Extremera A, O'Valle F, Sanjuán-Nuñez L, Martín-Alvarez AB, Martín-Oliva D, Caballero T, Muñoz de Rueda P, León J, Gonzalez R, Muntané J, Oliver FJ and Salmerón J: Inhibition of poly adenosine diphosphate-ribose polymerase decreases hepatocellular carcinoma growth by modulation of tumor-related gene expression. *Hepatology* 51: 255-266, 2010.
- 77 Rojo F, García-Parra J, Zazo S, Tusquets I, Ferrer-Lozano J, Menendez S, Eroles P, Chamizo C, Servitja S, Ramírez-Merino N, Lobo F, Bellosillo B, Corominas JM, Yelamos J, Serrano S, Lluch A, Rovira A and Albanell J: Nuclear PARP-1 protein overexpression is associated with poor overall survival in early breast cancer. *Ann Oncol* 23: 1156-1164, 2012.
- 78 Barton VN, Donson AM, Kleinschmidt-DeMasters BK, Gore L, Liu AK and Foreman NK: PARP1 expression in pediatric central nervous system tumors. *Pediatr Blood Cancer* 53: 1227-1230, 2009.
- 79 Barnett JC, Bean SM, Nakayama JM, Kondoh E, Murphy SK and Berchuck A: High poly(adenosine diphosphate-ribose) polymerase expression and poor survival in advanced-stage serous ovarian cancer. *Obstet Gynecol* 115: 49-54, 2010.

- 80 Gonçalves A, Finetti P, Sabatier R, Gilabert M, Adelaide J, Borg JP, Chaffanet M, Viens P, Birnbaum D and Bertucci F: Poly(ADP-ribose) polymerase-1 mRNA expression in human breast cancer: a meta-analysis. *Breast Cancer Res Treat* 127: 273-281, 2011
- 81 Zhang Q, Li Y, Li X, Zhou W, Shi B, Chen H and Yuan W: PARP-1 Val762Ala polymorphism, CagA+ H. pylori infection and risk for gastric cancer in Han Chinese population. *Mol Biol Rep* 36: 1461-1467, 2009.
- 82 Lockett KL, Hall MC, Xu J, Zheng SL, Berwick M, Chuang SC, Clark PE, Cramer SD, Lohman K and Hu JJ: The ADPRT V762A genetic variant contributes to prostate cancer susceptibility and deficient enzyme function. *Cancer Res* 64: 6344-6348, 2004.
- 83 Hao B, Wang H, Zhou K, Li Y, Chen X, Zhou G, Zhu Y, Miao X, Tan W, Wei Q, Lin D and He F: Identification of genetic variants in base excision repair pathway and their associations with risk of esophageal squamous cell carcinoma. *Cancer Res* 64: 4378-4384, 2004.
- 84 Zhang X, Miao X, Liang G, Hao B, Wang Y, Tan W, Li Y, Guo Y, He F, Wei Q and Lin D: Polymorphisms in DNA base excision repair genes ADPRT and XRCC1 and risk of lung cancer. *Cancer Res* 65: 722-726, 2005.
- 85 Jin XM, Kim HN, Lee IK, Park KS, Kim HJ, Choi JS, Juhng SW and Choi C: PARP-1 Val762Ala polymorphism is associated with reduced risk of non-Hodgkin lymphoma in Korean males. *BMC Med Genet* 11: 38, 2010.
- 86 Wang XG, Wang ZQ, Tong WM and Shen Y: PARP1 Val762Ala polymorphism reduces enzymatic activity. *Biochem Biophys Res Commun* 354: 122-126, 2007.
- 87 Yu H, Ma H, Yin M and Wei Q: Association between PARP-1 V762A polymorphism and cancer susceptibility: a meta-analysis. *Genet Epidemiol* 36: 56-65, 2011.
- 88 Shin BH, Choi SH, Cho EY, Shin MJ, Hwang KC, Cho HK, Chung JH and Jang Y: Thiamines attenuates hypoxia-induced cell death in cultured neonatal rat cardiomyocytes. *Mol Cells* 18: 133-140, 2004.
- 89 Yadav UCS, Kalariya NM, Srivastava SK and Ramana KV: Protective role of benfotiamine, a fat soluble vitamin B1 analogue, in the lipopolysaccharide-induced cytotoxic signals in murine macrophages. *Free Radic Biol Med* 48: 1423-1434, 2010.
- 90 Frédérich M, Delvaux D, Gigliobianco T, Gangolf M, Dive G, Mazzucchelli G, Elias B, De Pauw E, Angenot L, Wins P and Bettendorff L: Thiaminylated adenine nucleotides. *FEBS J* 276: 3256-3268, 2009.
- 91 Tanaka T, Yamamoto D, Sato T, Tanaka S, Usui K, Manabe M, Aoki Y, Iwashima Y, Saito Y, Mino Y and Deguchi H: Adenosine thiamine triphosphate (ATHTP) inhibits Poly(ADP-Ribose) polymerase-1 (PARP-1) activity. *J Nutr Sci Vitaminol* 57: 192-196, 2011.
- 92 Kamata T: Roles of Nox1 and other Nox isoforms in cancer development. *Cancer Sci* 100: 1382-1388, 2009.
- 93 Block K, Gorin Y, New DD, Eid A, Chelmicki T, Reed A, Choudhury GG, Parekh DJ and Abboud HE: The NADPH oxidase subunit p22phox inhibits the function of the tumor suppressor protein tuberlin. *Am J Pathol* 176: 2447-2455, 2010.
- 94 Yamaura M, Mitsushita J, Furuta S, Kuniwa Y, Ashida A, Goto Y, Shang WH, Kubodera M, Kato M, Takata M, Saida T and Kamata T: NADPH oxidase 4 contributes to transformation phenotype of melanoma cells by regulating G<sub>2</sub>-M cell cycle progression. *Cancer Res* 69: 2647-2654, 2009.
- 95 Maraldi T, Prata C, Vieceli Dalla Sega F, Caliceti C, Zamboni L, Fiorentini D and Hakim G: NAD(P)H oxidase isoform Nox2 plays a prosurvival role in human leukaemia cells. *Free Radic Res* 43: 1111-1121, 2009.
- 96 Hong J, Resnick M, Behar J, Wang LJ, Wands J, DeLellis RA, Souza RF, Spechler SJ and Cao W: Acid-induced p16 hypermethylation contributes to development of esophageal adenocarcinoma via activation of NADPH oxidase NOX5-S. *Am J Physiol Gastrointest Liver Physiol* 299: G697-706, 2010.
- 97 Lu CL, Qiu JL, Huang PZ, Zou RH, Hong J, Li BK, Chen GH and Yuan YF: NADPH oxidase DUOX1 and DUOX2 but not NOX4 are independent predictors in hepatocellular carcinoma after hepatectomy. *Tumour Biol* 32: 1173-1182, 2011.
- 98 Kim J, Koyanagi T and Mochly-Rosen D: PKC $\delta$  activation mediates angiogenesis via NADPH oxidase activity in PC-3 prostate cancer cells. *Prostate* 71: 946-954, 2011.
- 99 Wang R, Dashwood WM, Nian H, Löhr CV, Fischer KA, Tsuchiya N, Nakagama H, Ashktorab H and Dashwood RH: NADPH oxidase overexpression in human colon cancers and rat colon tumors induced by 2-amino-1-methyl-6-phenylimidazo [4,5-b]pyridine (PhIP). *Int J Cancer* 128: 2581-2590, 2011.
- 100 Hsieh CH, Shyu WC, Chiang CY, Kuo JW, Shen WC and Liu RS: NADPH oxidase subunit 4-mediated reactive oxygen species contribute to cycling hypoxia-promoted tumor progression in glioblastoma multiforme. *PLoS One* 6: e23945, 2011.
- 101 Yin L, Kosugi M and Kufe D: Inhibition of the MUC1-C oncoprotein induces multiple myeloma cell death by down-regulating TIGAR expression and depleting NADPH. *Blood* 119: 810-816, 2012.
- 102 Naoe T, Takeyama K, Yokozawa T, Kiyoi H, Seto M, Uike N, Ino T, Utsunomiya A, Maruta A, Jin-nai I, Kamada N, Kubota Y, Nakamura H, Shimazaki C, Horiike S, Kodera Y, Saito H, Ueda R, Wiemels J and Ohno R: Analysis of genetic polymorphism in NQO1, GST-M1, GST-T1, and CYP3A4 in 469 Japanese patients with therapy-related leukemia/ myelodysplastic syndrome and *de novo* acute myeloid leukemia. *Clin Cancer Res* 6: 4091-4095, 2000.
- 103 Fern L, Pallis M, Ian Carter G, Seedhouse C, Russell N and Byrne J: Clonal haemopoiesis may occur after conventional chemotherapy and is associated with accelerated telomere shortening and defects in the NQO1 pathway; possible mechanisms leading to an increased risk of t-AML/MDS. *Br J Haematol* 126: 63-71, 2004.
- 104 Hamajima N, Matsuo K, Iwata H, Shinoda M, Yamamura Y, Kato T, Hatooka S, Mitsudomi T, Suyama M, Kagami Y, Ogura M, Ando M, Sugimura Y and Tajima K: NAD(P)H: quinone oxidoreductase 1 (NQO1) C609T polymorphism and the risk of eight cancers for Japanese. *Int J Clin Oncol* 7: 103-108, 2002.
- 105 Chan EC, Lam SY, Fu KH and Kwong YL: Polymorphisms of the GSTM1, GSTP1, MPO, XRCC1, and NQO1 genes in Chinese patients with non-small cell lung cancers: relationship with aberrant promoter methylation of the CDKN2A and RARB genes. *Cancer Genet Cytogenet* 162: 10-20, 2005.
- 106 Lan Q, Zheng T, Shen M, Zhang Y, Wang SS, Zahm SH, Holford TR, Leaderer B, Boyle P and Chanock S: Genetic polymorphisms in the oxidative stress pathway and susceptibility to non-Hodgkin lymphoma. *Hum Genet* 121: 161-168, 2007.
- 107 Hoffmann M, Schirmer MA, Tzvetkov MV, Kreuz M, Ziepert M, Wojnowski L, Kube D, Pfreundschuh M, Trümper L, Loeffler M, Brockmüller J and German Study Group for High-Grade Non-

- Hodgkin Lymphoma: A functional polymorphism in the NAD(P)H oxidase subunit CYBA is related to gene expression, enzyme activity, and outcome in non-Hodgkin lymphoma. *Cancer Res* 70: 2328-2338, 2010.
- 108 Gra OA, Glotov AS, Kozhekbaeva Zh, Makarova OV and Nasedkina TV: [Genetic polymorphism in GST, NAT2, and MTRR and susceptibility to childhood acute leukemia]. *Mol Biol (Mosk)* 42: 214-225, 2008. (in Russian).
- 109 Seibold P, Hein R, Schmezer P, Hall P, Liu J, Dahmen N, Flesch-Janys D, Popanda O and Chang-Claude J: Polymorphisms in oxidative stress-related genes and postmenopausal breast cancer risk. *Int J Cancer* 129: 1467-1476, 2011.
- 110 Malik MA, Zargar SA and Mittal B: Role of NQO1 609C>T and NQO2-3423G>A polymorphisms in susceptibility to gastric cancer in Kashmir valley. *DNA Cell Biol* 30: 297-303, 2011.
- 111 Galdhar NR and Pawar SS: Hepatic drug metabolism and lipid peroxidation in thiamine deficient rats. *Int J Vitam Nutr Res* 46: 14-23, 1976.
- 112 Fraser DA, Hessvik NP, Nikolić N, Aas V, Hanssen KF, Bøhn SK, Thoresen GH and Rustan AC: Benfotiamine increases glucose oxidation and downregulates NADPH oxidase 4 expression in cultured human myotubes exposed to both normal and high glucose concentrations. *Genes Nutr* 7: 459-692, 2012.
- 113 Grosse W 3rd and Wade AE: The effect of thiamine consumption on liver microsomal drug-metabolizing pathways. *J Pharmacol Exp Ther* 176: 758-765, 1971
- 114 Sella T, Chodick G, Barchana M, Heymann AD, Porath A, Kokia E and Shalev V: Gestational diabetes and risk of incident primary cancer: a large historical cohort study in Israel. *Cancer Causes Control* 22: 1513-1520, 2011.
- 115 Lambe M, Wigertz A, Garmo H, Walldius G, Jungner I and Hammar N: Impaired glucose metabolism and diabetes and the risk of breast, endometrial, and ovarian cancer. *Cancer Causes Control* 22: 1163-1171, 2011.
- 116 Pelucchi C, Serraino D, Negri E, Montella M, Dellanoce C, Talamini R and La Vecchia C: The metabolic syndrome and risk of prostate cancer in Italy. *Ann Epidemiol* 21: 835-841, 2011.
- 117 Zendejdel K, Nyrén O, Ostenson CG, Adami HO, Ekblom A and Ye W: Cancer incidence in patients with type 1 diabetes mellitus: a population-based cohort study in Sweden. *J Natl Cancer Inst* 95: 1797-1800, 2003.
- 118 Swerdlow AJ, Laing SP, Qiao Z, Slater SD, Burden AC, Botha JL, Waugh NR, Morris AD, Gatling W, Gale EA, Patterson CC and Keen H: Cancer incidence and mortality in patients with insulin-treated diabetes: a UK cohort study. *Br J Cancer* 92: 2070-2075, 2005.
- 119 Huang W, Ren H, Ben Q, Cai Q, Zhu W and Li Z: Risk of esophageal cancer in diabetes mellitus: a meta-analysis of observational studies. *Cancer Causes Control* 23: 263-72, 2012.
- 120 Tian T, Zhang LQ, Ma XH, Zhou JN and Shen J: Diabetes Mellitus and Incidence and Mortality of Gastric Cancer: A Meta-Analysis. *Exp Clin Endocrinol Diabetes* 120: 217-223, 2012.
- 121 Luo W, Cao Y, Liao C and Gao F: Diabetes mellitus and the incidence and mortality of colorectal cancer: A meta-analysis of twenty four cohort studies. *Colorectal Dis* 14: 1307-1312, 2012.
- 122 Wang P, Kang D, Cao W, Wang Y and Liu Z: Diabetes mellitus and risk of hepatocellular carcinoma: a systematic review and meta-analysis. *Diabetes Metab Res Rev* 28: 109-122, 2012.
- 123 Jing W, Jin G, Zhou X, Zhou Y, Zhang Y, Shao C, Liu R and Hu X: Diabetes mellitus and increased risk of cholangiocarcinoma: a meta-analysis. *Eur J Cancer Prev* 21: 24-31, 2012.
- 124 Liao S, Li J, Wei W, Wang L, Zhang Y, Li J, Wang C and Sun S: Association between diabetes mellitus and breast cancer risk: a meta-analysis of the literature. *Asian Pac J Cancer Prev* 12: 1061-1065, 2011.
- 125 Habib SL, Prihoda TJ, Luna M and Werner SA: Diabetes and risk of renal cell carcinoma. *J Cancer* 3: 42-48, 2012.
- 126 Watson JD and Dako DY: Erythrocyte transketolase activity in adult Ghanaian subjects. *Clin Chim Acta* 59: 55-61, 1975.
- 127 Valerio G, Franzese A, Poggi V, Patrini C, C, Laforenza U and Tenore A: Lipophylic thiamine treatment in long-standing insulin-dependent diabetes mellitus. *Acta Diabetol* 36: 73-76, 1999.
- 128 Berant M, Berkovitz D, Mandel H, Zinder O and Mordohovich D: Thiamine status of the offspring of diabetic rats. *Pediatr Res* 23: 574-575, 1988.
- 129 Clark JA, Burny I, Sarnaik AP and Audhya TK: Acute thiamine deficiency in diabetic ketoacidosis: Diagnosis and management. *Pediatr Crit Care Med* 7: 595-599, 2006.
- 130 Oriot D, Wood C, Gottesman R and Huault G: Severe lactic acidosis related to acute thiamine deficiency. *JPEN J Parenter Enteral Nutr* 15: 105-109, 1991.
- 131 Saito N, Kimura M, Kuchiba A, and Itokawa Y: Blood thiamine levels in outpatients with diabetes mellitus. *J Nutr Sci Vitaminol* 33: 421-430, 1987.
- 132 Kjøsen BMS and Seim SH: The transketolase assay of thiamine in some diseases. *Am J Clin Nutr* 30: 1591-1596, 1977.
- 133 Thornalley PJ, Babaei-Jadidi R, Al Ali H, Rabbani N, Antonysunil A, Larkin J, Ahmed A, Rayman G and Bodmer CW: High prevalence of low thiamine concentration in diabetes linked to a marker of vascular disease. *Diabetologia* 50: 2164-2170, 2007.
- 134 Briggs MH: Vitamin and coenzyme content of hepatomas induced by buter yellow. *Nature* 187: 249-250, 1960.
- 135 Ayre JE and Bauld WA: Thiamine Deficiency and High Estrogen Findings in Uterine Cancer and in Menorrhagia. *Science* 103: 441-445, 1945.
- 136 Seligmann H, Levi R, Konijn AM and Prokocimer M: Thiamine deficiency in patients with B-chronic lymphocytic leukaemia: a pilot study. *Postgrad Med J* 77: 582-585, 2001.
- 137 Bruce WR, Ciocco M, Giacca A, Kim YI, Marcon N and Minkin S: A pilot randomised controlled trial to reduce colorectal cancer risk markers associated with B-vitamin deficiency, insulin resistance and colonic inflammation. *Br J Cancer* 93: 639-946, 2005.
- 138 Ostrovskii IuM, Trebukhina RV and Mikhail'tsevich GN: Tumor and body interrelationships in 14C-thiamine utilization]. *Biull Eksp Biol Med* 88: 708-710, 1979. (in Russian).
- 139 Boros LG, Brandes JL, Lee WN, Cascante M, Puigjaner J, Revesz E, Bray TM, Schirmer WJ and Melvin WS: Thiamine supplementation to cancer patients: a double edged sword. *Anticancer Res* 18: 595-602, 1998.
- 140 Pamukcu AM, Yalçiner S, Price JM and Bryan GT: Effects of the coadministration of thiamine on the incidence of urinary bladder carcinomas in rats fed bracken fern. *Cancer Res* 30: 2671-2674, 1970.
- 141 Suryo Rahmanto Y, Dunn LL and Richardson DR: Identification of distinct changes in gene expression after modulation of melanoma tumor antigen p97 (melanotransferrin) in multiple models *in vitro* and *in vivo*. *Carcinogenesis* 28: 2172-2183, 2007.

- 142 Endo N, Takeda Y, Umemoto N, Kishida K, Watanabe K, Saito M, Kato Y and Hara T: Nature of linkage and mode of action of methotrexate conjugated with antitumor antibodies: implications for future preparation of conjugates. *Cancer Res* 48: 3330-3335, 1988.
- 143 Naujokaitis SA: Thiamine protection of murine L1210 leukemia cells against mechlorethamine cytotoxicity and its relation to the choline uptake system. *Res Commun Chem Pathol Pharmacol* 32: 317-327, 1981.
- 144 Trebukhina RV, Koltaniuk TA, Petushok VG, Mikhal'tsevich GN, Lashak LK and Gritsenko EA: [Metabolism of vitamins B1 and PP and their use in oncological practice]. *Vopr Med Khim* 38: 33-36, 1992. (in Russian).
- 145 Liu S, Monks NR, Hanes JW, Begley TP, Yu H and Moscow JA: Sensitivity of breast cancer cell lines to recombinant thiaminase I. *Cancer Chemother Pharmacol* 66: 171-179, 2010.
- 146 Daily A, Liu S, Bae Y, Bhatnagar S and Moscow JA: Linear chain PEGylated recombinant *Bacillus thiaminolyticus* thiaminase I enzyme has growth inhibitory activity against lymphoid leukemia cell lines. *Mol Cancer Ther* 10: 1563-1570, 2011.
- 147 Trebukhina RV, Kravchuk RI, Mikhal'tsevich GN, Petushok VG and Nikitin VS: [Effect of thiamine and its antimetabolite oxythiamine on the proliferative activity of carcinosarcoma Walker 256 cells]. *Eksp Onkol* 9: 60-63, 1987. (in Russian).
- 148 Bruce WR, Furrer R, Shangari N, O'Brien PJ, Medline A and Wang Y: Marginal dietary thiamin deficiency induces the formation of colonic aberrant crypt foci (ACF) in rats. *Cancer Lett* 202: 125-129, 2003.
- 149 Salley JJ, Eshleman JR and Morgan JH: Effect of chronic thiamine deficiency on oral carcinogenesis. *J Dent Res* 41: 1405-1413, 1962.
- 150 Sheline CT: Thiamine supplementation attenuated hepatocellular carcinoma in the Atp7b mouse model of Wilson's disease. *Anticancer Res* 31: 3395-3399, 2011.
- 151 Lee BY, Yanamandra K and Bocchini JA Jr: Thiamin deficiency: a possible major cause of some tumors? (review). *Oncol Rep* 14: 1589-1592, 2005.
- 152 Comín-Anduix B, Boren J, Martinez S, Moro C, Centelles JJ, Trebukhina R, Petushok N, Lee WN, Boros LG and Cascante M: The effect of thiamine supplementation on tumour proliferation. A metabolic control analysis study. *Eur J Biochem* 268: 4177-4182, 2001.
- 153 Schupp N, Dette EM, Schmid U, Bahner U, Winkler M, Heidland A and Stopper H: Benfotiamine reduces genomic damage in peripheral lymphocytes of hemodialysis patients. *Naunyn Schmiedebergs Arch Pharmacol* 378: 283-291, 2008.
- 154 Holtkamp N, Atallah I, Okuducu AF, Mucha J, Hartmann C, Mautner VF, Friedrich RE, Mawrin C and von Deimling A: MMP-13 and p53 in the progression of malignant peripheral nerve sheath tumors. *Neoplasia* 9: 671-677, 2007.
- 155 Zhang Y, Zhao H, Wang Y, Lin Y, Tan Y, Fang X and Zheng L: Non-small cell lung cancer invasion and metastasis promoted by MMP-26. *Mol Med Report* 4: 1201-1209, 2011.
- 156 Yan W, Zhang W, Sun L, Liu Y, You G, Wang Y, Kang C, You Y and Jiang T: Identification of MMP-9 specific microRNA expression profile as potential targets of anti-invasion therapy in glioblastoma multiforme. *Brain Res* 1411: 108-115, 2011.
- 157 Garamszegi N, Garamszegi SP and Scully SP: Matrix metalloproteinase-1 contribution to sarcoma cell invasion. *J Cell Mol Med* 16: 1331-1341, 2012.
- 158 Hong SW, Kang YK, Lee B, Lee WY, Jang YG, Paik IW and Lee H: Matrix metalloproteinase-2 and -7 expression in colorectal cancer. *J Korean Soc Coloproctol* 27: 133-139, 2011.
- 159 Shin YJ and Kim JH: The Role of EZH2 in the Regulation of the Activity of Matrix Metalloproteinases in Prostate Cancer Cells. *PLoS One* 7: e30393, 2012.
- 160 Ghosh S, Basu M and Roy SS: ETS-1 regulates vascular endothelial growth factor-induced matrix metalloproteinase-9 and matrix metalloproteinase-13 expression in human ovarian carcinoma cell SKOV-3. *J Biol Chem* 287: 15001-15015, 2012.
- 161 Ghilardi G, Biondi ML, Caputo M, Leviti S, DeMonti M, Guagnellini E and Scorza R: A single nucleotide polymorphism in the matrix metalloproteinase-3 promoter enhances breast cancer susceptibility. *Clin Cancer Res* 8: 3820-3823, 2002.
- 162 Xu E, Xia X, Lü B, Xing X, Huang Q, Ma Y, Wang W and Lai M: Association of matrix metalloproteinase-2 and -9 promoter polymorphisms with colorectal cancer in Chinese. *Mol Carcinog* 46: 924-929, 2007.
- 163 Hirata H, Okayama N, Naito K, Inoue R, Yoshihiro S, Matsuyama H, Suehiro Y, Hamanaka Y and Hinoda Y: Association of a haplotype of matrix metalloproteinase (MMP)-1 and MMP-3 polymorphisms with renal cell carcinoma. *Carcinogenesis* 25: 2379-2384, 2004.
- 164 Lu Z, Wang Y, Zhang Q, Zhang X, Wang S, Xie H, Li Y, Jiao B and Zhang J: Association between the functional polymorphism in the matrix metalloproteinase-7 promoter and susceptibility to adult astrocytoma. *Brain Res* 1118: 6-12, 2006.
- 165 Sugimoto M, Yoshida S, Kennedy S, Deguchi M, Ohara N and Maruo T: Matrix metalloproteinase-1 and -9 promoter polymorphisms and endometrial carcinoma risk in a Japanese population. *J Soc Gynecol Investig* 13: 523-529, 2006.
- 166 Kader AK, Shao L, Dinney CP, Schabath MB, Wang Y, Liu J, Gu J, Grossman HB and Wu X: Matrix metalloproteinase polymorphisms and bladder cancer risk. *Cancer Res* 66: 11644-11648, 2006.
- 167 Li Y, Jin X, Kang S, Wang Y, Du H, Zhang J, Guo W, Wang N and Fang S: Polymorphisms in the promoter regions of the matrix metalloproteinases-1, -3, -7, and -9 and the risk of epithelial ovarian cancer in China. *Gynecol Oncol* 101: 92-96, 2006.
- 168 Vairaktaris E, Yapjakis C, Vasiliou S, Derka S, Nkenke E, Serefoglou Z, Vorriss E, Vylliotis A, Ragos V, Neukam FW and Patsouris E: Association of -1171 promoter polymorphism of matrix metalloproteinase-3 with increased risk for oral cancer. *Anticancer Res* 27: 4095-4100, 2007.
- 169 Singh H, Jain M and Mittal B: MMP-7 (-181A>G) promoter polymorphisms and risk for cervical cancer. *Gynecol Oncol* 110: 71-75, 2008.
- 170 Wu J, Zhang L, Luo H, Zhu Z, Zhang C and Hou Y: Association of matrix metalloproteinases-9 gene polymorphisms with genetic susceptibility to esophageal squamous cell carcinoma. *DNA Cell Biol* 27: 553-557, 2008.
- 171 Alakus H, Afriani N, Warnecke-Eberz U, Bollschweiler E, Fetzner U, Drebber U, Metzger R, Hölscher AH and Mönig SP: Clinical impact of MMP and TIMP gene polymorphisms in gastric cancer. *World J Surg* 34: 2853-2859, 2010.
- 172 Chaudhary AK, Pandya S, Mehrotra R, Bharti AC, Jain S and Singh M: Functional polymorphism of the MMP-1 promoter (-1607 1G/2G) in potentially malignant and malignant head and neck lesions in an Indian population. *Biomarkers* 15: 684-692, 2010.

- 173 Altaş M, Bayrak OF, Ayan E, Bolukbasi F, Silav G, Coskun KK, Culha M, Sahin F, Sevli S and Elmaci I: The effect of polymorphisms in the promoter region of the MMP-1 gene on the occurrence and invasiveness of hypophyseal adenoma. *Acta Neurochir (Wien)* 152: 1611-1617, 2010.
- 174 Gil Ugarteburu R, Rivas del Fresno M, González Rodríguez I, González Arriaga P, López Cima F, Fernández Samoano A, Fernández García I, Benito García P, Muruamendiáraz Fernández V and Tardón A: Matrix metalloproteinase-9 polymorphisms in the diagnosis of prostate cancer. A preliminary experience. *Arch Esp Urol* 63: 125-132, 2010.
- 175 Malik N, Kumar R, Prasad KN, Kawal P, Srivastava A and Mahapatra AK: Association of matrix metalloproteinase-1 gene polymorphism with glioblastoma multiforme in a northern Indian population. *J Neurooncol* 102: 347-352, 2011.
- 176 Chen TY, Li YC, Liu YF, Tsai CM, Hsieh YH, Lin CW, Yang SF and Weng CJ: Role of MMP14 gene polymorphisms in susceptibility and pathological development to hepatocellular carcinoma. *Ann Surg Oncol* 18: 2348-2356, 2011.
- 177 Lei Z, Liu R, Chen J, Zhou Q and Zhang H: [Meta Analysis of Association between Polymorphisms in Promoter Region of MMPs gene and Risk of Lung Cancer.]. *Zhongguo Fei Ai Za Zhi* 12: 381-386, 2009. (in Chinese).
- 178 Zhou P, Du LF, Lv GQ, Yu XM, Gu YL, Li JP and Zhang C: Current evidence on the relationship between four polymorphisms in the matrix metalloproteinases (MMP) gene and breast cancer risk: a meta-analysis. *Breast Cancer Res Treat* 127: 813-818, 2011.
- 179 Zhang LY and Ren KW: Meta-analysis of MMP2 -1306T allele as a protective factor in digestive cancer. *Arch Med Res* 42: 239-243, 2011.
- 180 Peng B, Cao L, Wang W, Xian L, Jiang D, Zhao J, Zhang Z, Wang X and Yu L: Polymorphisms in the promoter regions of matrix metalloproteinases 1 and 3 and cancer risk: a meta-analysis of 50 case-control studies. *Mutagenesis* 25: 41-48, 2010.
- 181 Peng B, Cao L, Ma X, Wang W, Wang D and Yu L: Meta-analysis of association between matrix metalloproteinases 2, 7 and 9 promoter polymorphisms and cancer risk. *Mutagenesis* 25: 371-379, 2010.
- 182 Calingasan NY and Gibson GE: Dietary restriction attenuates the neuronal loss, induction of heme oxygenase-1 and blood-brain barrier breakdown induced by impaired oxidative metabolism. *Brain Res* 885: 62-69, 2000.
- 183 Beauchesne É, Desjardins P, Hazell AS and Butterworth RF: eNOS gene deletion restores blood-brain barrier integrity and attenuates neurodegeneration in the thiamine-deficient mouse brain. *J Neurochem* 111: 452-459, 2009.
- 184 Kohda Y, Shirakawa H, Yamane K, Otsuka K, Kono T, Terasaki F and Tanaka T: Prevention of incipient diabetic cardiomyopathy by high-dose thiamine. *J Toxicol Sci* 33: 459-472, 2008.
- 185 Tarallo S, Beltramo E, Berrone E, Dentelli P and Porta M: Effects of high glucose and thiamine on the balance between matrix metalloproteinases and their tissue inhibitors in vascular cells. *Acta Diabetol* 47: 105-111, 2010.
- 186 Gu B, Desjardins P and Butterworth RF: Selective increase of neuronal cyclooxygenase-2 (COX-2) expression in vulvulnerable brain regions of rats with experimental Wernicke's encephalopathy: effect of nimesulide. *Metab Brain Dis* 23: 175-187, 2008.
- 187 Liu S, Stromberg A, Tai HH and Moscow JA: Thiamine transporter gene expression and exogenous thiamine modulate the expression of genes involved in drug and prostaglandin metabolism in breast cancer cells. *Mol Cancer Res* 2: 477-487, 2004.
- 188 Tai HH, Tong M and Ding Y: 15-hydroxyprostaglandin dehydrogenase (15-PGDH) and lung cancer. *Prostaglandins Other Lipid Mediat* 83: 203-208, 2007.
- 189 Yaday UC, Subramanyam S and Ramana KV: Prevention of endotoxin-induced uveitis in rats by benfotiamine, a lipophilic analogue of vitamin B1. *Invest Ophthalmol Vis Sci* 50: 2276-2282, 2009.
- 190 Yaday UC, Kalariya NM, Srivastava SK and Ramana KV: Protective role of benfotiamine, a fat-soluble vitamin B1 analogue, in lipopolysaccharide-induced cytotoxic signals in murine macrophages. *Free Radic Biol Med* 48: 1423-1434, 2010.
- 191 Shoeb M and Ramana KV: Anti-inflammatory effects of benfotiamine are mediated through the regulation of the arachidonic acid pathway in macrophages. *Free Radic Biol Med* 52: 182-190, 2012.
- 192 Jabbour HN, Milne SA, Williams AR, Anderson RA and Boddy SC: Expression of COX-2 and PGE synthase and synthesis of PGE(2) in endometrial adenocarcinoma: a possible autocrine/paracrine regulation of neoplastic cell function via EP2/EP4 receptors. *Br J Cancer* 85: 1023-1031, 2001.
- 193 Kundu N, Yang Q, Dorsey R and Fulton AM: Increased cyclooxygenase-2 (cox-2) expression and activity in a murine model of metastatic breast cancer. *Int J Cancer* 93: 681-686, 2001.
- 194 Gatalica Z, Lilleberg SL, Koul MS, Vanecek T, Hes O, Wang B and Michal M: COX-2 gene polymorphisms and protein expression in renomedullary interstitial cell tumors. *Hum Pathol* 39: 1495-1504, 2008.
- 195 Okawa T, Yoshinaga K, Uetake H, Sato T, Higuchi T, Nemoto T and Sugihara K: Cyclooxygenase-2 overexpression is related to polypoid growth and K-ras gene mutation in T1 colorectal carcinomas. *Dis Colon Rectum* 47: 1915-1921, 2004.
- 196 Leung WK, To KF, Ng YP, Lee TL, Lau JY, Chan FK, Ng EK, Chung SC and Sung JJ: Association between cyclo-oxygenase-2 overexpression and missense p53 mutations in gastric cancer. *Br J Cancer* 84: 335-339, 2001.
- 197 Sales KJ, Katz AA, Davis M, Hinz S, Soeters RP, Hofmeyr MD, Millar RP and Jabbour HN: Cyclooxygenase-2 expression and prostaglandin E(2) synthesis are up-regulated in carcinomas of the cervix: a possible autocrine/paracrine regulation of neoplastic cell function via EP2/EP4 receptors. *J Clin Endocrinol Metab* 86: 2243-2249, 2001.
- 198 Brosens LA, Iacobuzio-Donahue CA, Keller JJ, Hustinx SR, Carvalho R, Morsink FH, Hyland LM, Offerhaus GJ, Giardiello FM and Goggins M: Increased cyclooxygenase-2 expression in duodenal compared with colonic tissues in familial adenomatous polyposis and relationship to the -765G -> C COX-2 polymorphism. *Clin Cancer Res* 11: 4090-4096, 2005.
- 199 Markosyan N, Chen EP, Ndong VN, Yao Y, Sterner CJ, Chodosh LA, Lawson JA, Fitzgerald GA and Smyth EM: Deletion of cyclooxygenase 2 in mouse mammary epithelial cells delays breast cancer onset through augmentation of type 1 immune responses in tumors. *Carcinogenesis* 32: 1441-1449, 2011.

- 200 Santander S, Cebrián C, Esquivias P, Conde B, Esteva F, Jiménez P, Ortego J, Lanas A and Piazzuelo E: Cyclooxygenase inhibitors decrease the growth and induce regression of human esophageal adenocarcinoma xenografts in nude mice. *Int J Oncol* 40: 527-534, 2012.
- 201 Rozic JG, Chakraborty C and Lala PK: Cyclooxygenase inhibitors retard murine mammary tumor progression by reducing tumor cell migration, invasiveness and angiogenesis. *Int J Cancer* 93: 497-506, 2001.
- 202 Kang S, Kim YB, Kim MH, Yoon KS, Kim JW, Park NH, Song YS, Kang D, Yoo KY, Kang SB and Lee HP: Polymorphism in the nuclear factor kappa-B binding promoter region of cyclooxygenase-2 is associated with an increased risk of bladder cancer. *Cancer Lett* 217: 11-16, 2005.
- 203 Sakoda LC, Gao YT, Chen BE, Chen J, Rosenberg PS, Rashid A, Deng J, Shen MC, Wang BS, Han TQ, Zhang BH, Cohen-Webb H, Yeager M, Welch R, Chanock S, Fraumeni JF Jr and Hsing AW: Prostaglandin-endoperoxide synthase 2 (PTGS2) gene polymorphisms and risk of biliary tract cancer and gallstones: a population-based study in Shanghai, China. *Carcinogenesis* 27: 1251-1256, 2006.
- 204 Park JM, Choi JE, Chae MH, Lee WK, Cha SI, Son JW, Kim CH, Kam S, Kang YM, Jung TH and Park JY: Relationship between cyclooxygenase 8473T>C polymorphism and the risk of lung cancer: a case-control study. *BMC Cancer* 6: 70, 2006.
- 205 Lira MG, Mazzola S, Tessari G, Malerba G, Ortombina M, Naldi L, Remuzzi G, Boschiero L, Forni A, Rugiu C, Piaserico S, Girolomoni G and Turco A: Association of functional gene variants in the regulatory regions of COX-2 gene (PTGS2) with nonmelanoma skin cancer after organ transplantation. *Br J Dermatol* 157: 49-57, 2007.
- 206 Upadhyay R, Jain M, Kumar S, Ghoshal UC and Mittal B: Functional polymorphisms of cyclooxygenase-2 (COX-2) gene and risk for esophageal squamous cell carcinoma. *Mutat Res* 663: 52-59, 2009.
- 207 Ben Nasr H, Chahed K, Bouaouina N and Chouchane L: PTGS2 (COX-2) -765 G > C functional promoter polymorphism and its association with risk and lymph node metastasis in nasopharyngeal carcinoma. *Mol Biol Rep* 36: 193-200, 2009.
- 208 Zhao D, Xu D, Zhang X, Wang L, Tan W, Guo Y, Yu D, Li H, Zhao P and Lin D: Interaction of cyclooxygenase-2 variants and smoking in pancreatic cancer: a possible role of nucleophosmin. *Gastroenterology* 136: 1659-1668, 2009.
- 209 Lurie G, Terry KL, Wilkens LR, Thompson PJ, McDuffie KE, Carney ME, Palmieri RT, Cramer DW and Goodman MT: Pooled analysis of the association of PTGS2 rs5275 polymorphism and NSAID use with invasive ovarian carcinoma risk. *Cancer Causes Control* 21: 1731-1741, 2010.
- 210 Abraham JE, Harrington P, Driver KE, Tyrer J, Easton DF, Dunning AM and Pharoah PD: Common polymorphisms in the prostaglandin pathway genes and their association with breast cancer susceptibility and survival. *Clin Cancer Res* 15: 2181-2191, 2009.
- 211 Li Y, He W, Liu T and Zhang Q: A new cyclo-oxygenase-2 gene variant in the Han Chinese population is associated with an increased risk of gastric carcinoma. *Mol Diagn Ther* 14: 351-355, 2010.
- 212 Zheng J, Chen S, Jiang L, You Y, Wu D and Zhou Y: Functional genetic variations of cyclooxygenase-2 and susceptibility to acute myeloid leukemia in a Chinese population. *Eur J Haematol* 87: 486-493, 2011.
- 213 Amirian ES, Ittmann MM and Scheurer ME: Associations between arachidonic acid metabolism gene polymorphisms and prostate cancer risk. *Prostate* 71: 1382-1389, 2011.
- 214 Peters WH, Lacko M, Te Morsche RH, Voogd AC, Oude Ophuis MB and Manni JJ: COX-2 polymorphisms and the risk for head and neck cancer in white patients. *Head Neck* 31: 938-943, 2009.
- 215 Gong Z, Bostick RM, Xie D, Hurley TG, Deng Z, Dixon DA, Zhang J and Hebert JR: Genetic polymorphisms in the cyclooxygenase-1 and cyclooxygenase-2 genes and risk of colorectal adenoma. *Int J Colorectal Dis* 24: 647-654, 2009.
- 216 Akkız H, Bayram S, Bekar A, Akgöllü E and Ülger Y: Functional polymorphisms of cyclooxygenase-2 gene and risk for hepatocellular carcinoma. *Mol Cell Biochem* 347: 201-208, 2011.
- 217 Dong J, Dai J, Zhang M, Hu Z and Shen H: Potentially functional COX-2-1195G>A polymorphism increases the risk of digestive system cancers: a meta-analysis. *J Gastroenterol Hepatol* 25: 1042-1050, 2010.
- 218 Zhu W, Wei BB, Shan X and Liu P: -765G>C and 8473T>C polymorphisms of COX-2 and cancer risk: a meta-analysis based on 33 case-control studies. *Mol Biol Rep* 37: 277-288, 2010.
- 219 Shrubsole MJ, Cai Q, Wen W, Milne G, Smalley WE, Chen Z, Ness RM and Zheng W: Urinary prostaglandin E2 Metabolite and Risk for Colorectal Adenoma. *Cancer Prev Res (Phila)* 5: 336-342, 2012.
- 220 Hogendorf P, Durczyński A, Kumor A and Strzelczyk J: Prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) in Portal Blood in Patients with Pancreatic Tumor-A Single Institution Series. *J Invest Surg* 25: 8-13, 2012.
- 221 Rasmuson A, Kock A, Fuskevåg OM, Kruspig B, Simón-Santamaría J, Gogvadze V, Johnsen JI, Kogner P and Sveinbjörnsson B: Autocrine prostaglandin e(2) signaling promotes tumor cell survival and proliferation in childhood neuroblastoma. *PLoS One* 7: e29331, 2012.
- 222 Han S and Roman J: Suppression of prostaglandin E2 receptor subtype EP2 by PPARgamma ligands inhibits human lung carcinoma cell growth. *Biochem Biophys Res Commun* 314: 1093-1099, 2004.
- 223 Gupta SC, Hevia D, Patchva S, Park B, Koh W and Aggarwal BB: Upsides and Downsides of Reactive Oxygen Species for Cancer: The Roles of Reactive Oxygen Species in Tumorigenesis, Prevention, and Therapy. *Antioxid Redox Signal* 16: 1295-1322, 2012.
- 224 Marra M, Sordelli IM, Lombardi A, Lamberti M, Tarantino L, Giudice A, Stiuso P, Abbruzzese A, Sperlongano R, Accardo M, Agresti M, Caraglia M and Sperlongano P: Molecular targets and oxidative stress biomarkers in hepatocellular carcinoma: an overview. *J Transl Med* 9: 171, 2011.
- 225 Zhang Q, Ma Y, Cheng YF, Li WJ, Zhang Z and Chen SY: Involvement of reactive oxygen species in 2-methoxyestradiol-induced apoptosis in human neuroblastoma cells. *Cancer Lett* 313: 201-210, 2011.
- 226 Wang HC and Choudhary S: Reactive oxygen species-mediated therapeutic control of bladder cancer. *Nat Rev Urol* 8: 608-616, 2011.
- 227 Rogalska A, Gajek A, Szwed M, Józwiak Z and Marczak A: The role of reactive oxygen species in WP 631-induced death of human ovarian cancer cells: a comparison with the effect of doxorubicin. *Toxicol In Vitro* 25: 1712-1720, 2011.

- 228 Gupta-Elera G, Garrett AR, Robison RA and O'Neill KL: The role of oxidative stress in prostate cancer. *Eur J Cancer Prev* 21: 155-162, 2012.
- 229 Hong CC, Ambrosone CB, Ahn J, Choi JY, McCullough ML, Stevens VL, Rodriguez C, Thun MJ and Calle EE: Genetic variability in iron-related oxidative stress pathways (Nrf2, NQO1, NOS3, and HO-1), iron intake, and risk of postmenopausal breast cancer. *Cancer Epidemiol Biomarkers Prev* 16: 1784-1794, 2007.
- 230 Woodson K, Tangrea JA, Lehman TA, Modali R, Taylor KM, Snyder K, Taylor PR, Virtamo J and Albanes D: Manganese superoxide dismutase (MnSOD) polymorphism, alpha-tocopherol supplementation and prostate cancer risk in the alpha-tocopherol, beta-carotene cancer prevention study (Finland). *Cancer Causes Control* 14: 513-518, 2003.
- 231 Wang S, Wang F, Shi X, Dai J, Peng Y, Guo X, Wang X, Shen H and Hu Z: Association between manganese superoxide dismutase (MnSOD) Val-9Ala polymorphism and cancer risk - A meta-analysis. *Eur J Cancer* 45: 2874-2881, 2009.
- 232 Mao C, Qiu LX, Zhan P, Xue K, Ding H, Du FB, Li J and Chen Q: MnSOD Val16Ala polymorphism and prostate cancer susceptibility: a meta-analysis involving 8,962 subjects. *J Cancer Res Clin Oncol* 136: 975-979, 2010.
- 233 Ma X, Chen C, Xiong H, Fan J, Li Y, Lin H, Xu R, Huang G and Xu B: No association between SOD2 Val16Ala polymorphism and breast cancer susceptibility: a meta-analysis based on 9,710 cases and 11,041 controls. *Breast Cancer Res Treat* 122: 509-514, 2010.
- 234 Calingasan NY, Chun WJ, Park LC and Gibson GE: Oxidative stress is associated with region-specific neuronal death during thiamine deficiency. *J Neuropathol Exp Neurol* 58: 946-958, 1999.
- 235 Lukienko PI, Mel'nichenko NG, Zverinskii IV and Zabrodskaya SV: Antioxidant properties of thiamine. *Bull Exp Biol Med* 130: 874-876, 2000.
- 236 de Souza Pinto R, Castilho G, Paim BA, Machado-Lima A, Inada NM, Nakandakare ER, Vercesi AE and Passarelli M: Inhibition of macrophage oxidative stress prevents the reduction of ABCA-1 transporter induced by advanced glycosylated albumin. *Lipids* 47: 443-450, 2012.
- 237 Kihm LP, Müller-Krebs S, Klein J, Ehrlich G, Mertes L and Gross ML, Adaikalakoteswari A, Thornalley PJ, Hammes HP, Nawroth PP, Zeier M, Schwenger V: Benfotiamine protects against peritoneal and kidney damage in peritoneal dialysis. *J Am Soc Nephrol* 22: 914-926, 2011.
- 238 Mehta R, Dedina L and O'Brien PJ: Rescuing hepatocytes from iron-catalyzed oxidative stress using vitamins B1 and B6. *Toxicol In Vitro* 25: 1114-1122, 2011.
- 239 Le Pennec S, Mirebeau-Prunier D, Boutet-Bouzamondo N, Jacques C, Guillotin D, Lauret E, Houlgatte R, Malthiery Y and Savagner F: Nitric oxide and calcium participate in the fine regulation of mitochondrial biogenesis in follicular thyroid carcinoma cells 286: 18229-18239, 2011.
- 240 Bentz BG, Chandra R, Haines GK 3rd, Robinson AM, Shah P and Radosевич JA: Nitric oxide and apoptosis during human head and neck squamous cell carcinoma development. *Am J Otolaryngol* 23: 4-11, 2002.
- 241 Siripin D, Fucharoen S and Tanyong DI: Nitric oxide and caspase 3 mediated cytokine induced apoptosis in acute leukemia. *Asian Pac J Allergy Immunol* 29: 102-111, 2011.
- 242 Saleem W, Suzuki Y, Mobaraki A, Yoshida Y, Noda S, Saitoh JI and Nakano T: Reduction of nitric oxide level enhances the radiosensitivity of hypoxic non-small cell lung cancer. *Cancer Sci* 102: 2150-2156, 2011.
- 243 Guler E, Baspinar O, Cekmen M, Kilinc M and Balat A: Nitric oxide: a new biomarker of Doxorubicin toxicity in children? *Pediatr Hemiatr Oncol* 28: 395-402, 2011.
- 244 Chin MP and Deen WM: Prediction of nitric oxide concentrations in melanomas. *Nitric Oxide* 23: 319-326, 2010.
- 245 Ellie E, Loiseau H, Lafond F, Arsaut J and Demotes-Mainard J: Differential expression of inducible nitric oxide synthase mRNA in human brain tumours. *Neuroreport* 7: 294-296, 1995.
- 246 Zhang W, He XJ, Ma YY, Wang HJ, Xia YJ, Zhao ZS, Ye ZY and Tao HQ: Inducible nitric oxide synthase expression correlates with angiogenesis, lymphangiogenesis, and poor prognosis in gastric cancer patients. *Hum Pathol* 42: 1275-1282, 2011.
- 247 Glynn SA, Boersma BJ, Dorsey TH, Yi M, Yfantis HG, Ridnour LA, Martin DN, Switzer CH, Hudson RS, Wink DA, Lee DH, Stephens RM, and Ambs S: Increased NOS2 predicts poor survival in estrogen receptor-negative breast cancer patients. *J Clin Invest* 120: 3843-3854, 2010.
- 248 Cardnell RJ and Mikkelsen RB: Nitric oxide synthase inhibition enhances the antitumor effect of radiation in the treatment of squamous carcinoma xenografts. *Plos One* 6: e20147, 2011.
- 249 Ryk C, Steineck G, Wiklund NP, Nyberg T and de Verdier PJ: The (CCTT)n microsatellite polymorphism in the nitric oxide synthase 2 gene may influence bladder cancer pathogenesis. *J Urol* 184: 2150-2157, 2010.
- 250 Shen CH, Wang YH, Wang WC, Jou YC, Hsu HS, Hsieh HY and Chiou HY: Inducible nitric oxide synthase promoter polymorphism, cigarette smoking, and urothelial carcinoma. *Urology* 69: 1001-1006, 2007.
- 251 Kaise M, Miwa J, Suzuki N, Mishiro S, Ohta Y, Yamasaki T and Tajiri H: Inducible nitric oxide synthase gene promoter polymorphism is associated with increased gastric mRNA expression of inducible nitric oxide synthase and increased risk of gastric carcinoma. *Eur J Gastroenterol Hepatol* 19: 139-145, 2007.
- 252 Arkan S, Cacina C, Guler E, Culcu S, Tuna G and Yaylım-Eraltan I: The effects of NOS3 Glu298Asp variant on colorectal cancer risk and progression in Turkish population. *Mol Biol Rep* 39: 3245-3249, 2012.
- 253 Han X, Zheng T, Lan Q, Zhang Y, Kilfoy BA, Qin Q, Rothman N, Zahm SH, Holford TR, Leaderer B and Zhang Y: Genetic polymorphisms in nitric oxide synthase genes modify the relationship between vegetable and fruit intake and risk of non-Hodgkin lymphoma. *Cancer Epidemiol Biomarkers Prev* 18: 1429-1438, 2009.
- 254 Hao Y, Montiel R and Huang Y: Endothelial nitric oxide synthase (eNOS) 894 G>T polymorphism is associated with breast cancer risk: a meta-analysis. *Breast Cancer Res Treat* 124: 809-813, 2010.
- 255 Kruse M, Navarro D, Desjardins P and Butterworth RF: Increased brain endothelial nitric oxide synthase expression in thiamine deficiency: relationship to selective vulnerability. *Neurochem Int* 45: 49-56, 2004.
- 256 Luong K and Nguyen L: The impact of thiamine treatment on generalized anxiety disorder. *Int J Clin Med* 2: 439-443, 2011.

Received April 12, 2013

Revised May 3 2013

Accepted May 9, 2013