

Review

# Are we Missing the Target? – Cancer Stem Cells and Drug Resistance in Non-small Cell Lung Cancer

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**Abstract.** *Non-small cell lung cancer (NSCLC) is the leading cause of cancer-related death in Western countries. Despite novel molecular therapies, the majority of patients with advanced or metastatic disease show rapid progression and a median survival time of not more than 18 months. In the last decade, there has been increasing evidence that cancer stem cells (CSC) play a pivotal role in drug resistance, tumour regeneration and metastasis of various cancer entities including lung cancer. In this review, we discuss the evidence for stem cells in NSCLC, their predictive and prognostic significance, their specific mechanisms of resistance and potential targets and strategies for eradication of these cells. Consideration of the specific properties of CSC in lung cancer therapy might substantially contribute to increased response and prolonged survival rates in this disease.*

Lung cancer is the leading cause of cancer-related death in Western countries (1). Non-small cell lung cancer (NSCLC) accounts for approximately 85% of all lung cancer cases and is divided into the major histological subtypes of adenocarcinoma (AC), squamous cell carcinoma (SCC) and large cell carcinoma (LCC). About 70% of all newly-diagnosed patients present with locally-advanced or metastatic disease and require systemic treatment (2). Despite novel molecular therapies, the prognosis of lung cancer is still poor and shows a median survival time of about 18 months in inoperable stages.

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**Key Words:** Non-small cell lung cancer, cancer stem cells, drug resistance, targeted therapy, review.

Lung cancer is characterized by frequent intratumoural heterogeneity (3). Up to 63% of all NSCLC were reported to contain more than one histologic subtype. This led to the assumption that lung cancer derives from a multipotent stem-like cell which is able to generate these various histological components.

The major criteria of stem cells is their ability to self-renew and differentiate into various cellular lineages via asymmetric division. Moreover, stem cells are characterized by slow division kinetics, drug and irradiation resistance and an increased capacity of invasion, metastasis, tumour formation and proliferation (4-7). Two types of stem cells are distinguished: pluripotent embryonic stem cells (ESC) which are able to differentiate into cells of all three germ layers and multipotent adult stem cells which are able to differentiate into various cells of a specific tissue or organ. Since the knowledge on ESC in lung cancer is yet scant, this review focuses on the relevance of adult stem cells in this disease.

So far, serial transplantation experiments in rodent models represent the gold standard for the approval of stem cell characteristics. Moreover, several surface antigens such as CD24, CD34, CD44, CD117 and CD133 and the expression of *aldehyde dehydrogenase 1A1* (*ALDH1A1*) are used for the identification and prospective isolation of adult stem cells. In addition, the ability to extrude dyes such as Hoechst 33342 or Rhodamine 123 and the slow division kinetics of stem cells, which results in the retention of membrane or DNA dyes such as PKH, carboxyfluorescein succinimidyl ester (CFSE) or bromdesoxyuridine (BrDU), are used to identify these cells (Figure 1).

Two general concepts of stem cell-driven tumorigenesis exist (Figure 2): In the deterministic model, all tumour cells originate from a degenerated stem cell or a de-differentiated tumour cell which acquired stem cells characteristics. This cancer stem cell (CSC) simultaneously self-renews and generates committed tumour cells *via* asymmetric division (8, 9). The committed tumour cells are thought to follow a

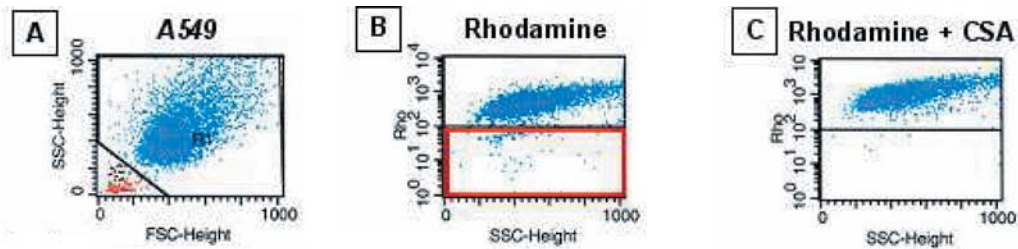


Figure 1. Side population (SP) of the adenocarcinoma cell line A549. The adenocarcinoma cell line A549 was stained with Rhodamine 123 (Rho), a dye extruded by the ABC transporter multidrug resistance-1 (MDR1). After 2 h, a small subpopulation of cells had effluxed the dye and showed a decreased fluorescence intensity (red box; picture B). When the ABC transporter was blocked with cyclosporine A (CSA), the cells expressing this drug transporter were not able to extrude the dye and maintained their initial fluorescence intensity (picture C). Dead cells were excluded from analysis (picture A). Dot plots: FSC: forward scatter, SSC: side scatter, y-axis: fluorescence intensity (log), horizontal line within the dot plots: autofluorescence cut-off.

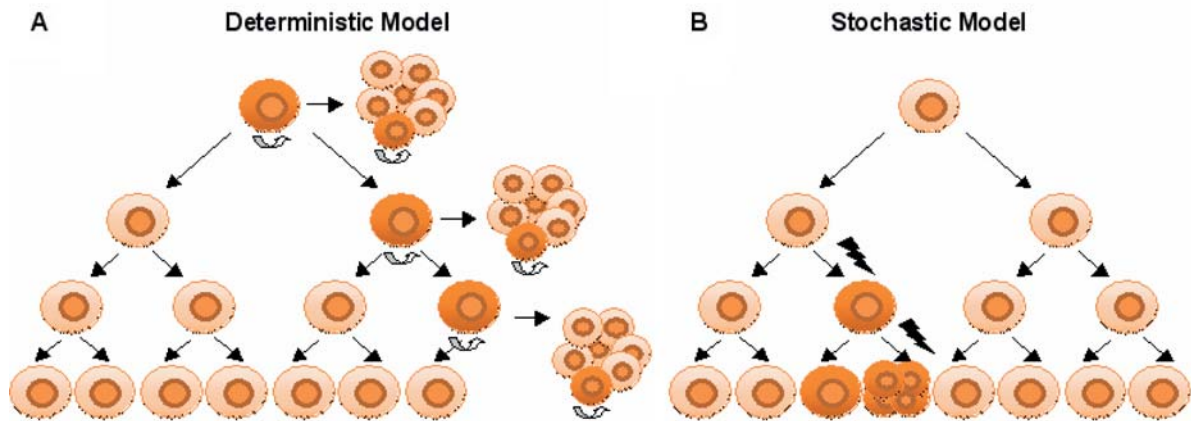


Figure 2. Models of stem cell-driven tumorigenesis. A: Deterministic model. The tumor originates from a cancer stem cell (CSC; dark brown) which simultaneously generates committed tumour cells (light brown) and CSC by asymmetric cell division. B: Stochastic model. The tumour cells (light brown) acquire genetic alterations (flashes) and undergo clonal evolution resulting in the generation of dominant clones with CSC properties (dark brown). All cells undergo symmetric division. In reality both concepts might occur in parallel or in combination.

pre-destinated and irreversible route of differentiation. The stochastic model, in contrast, assumes that one or more tumour cells randomly acquire genetic alterations which induce stem cell characteristics. Subsequent clonal selection results in the evolution of dominant clones which mainly sustain the tumour growth and proliferate *via* symmetric division. Differentiation of tumour cells occurs randomly and reversibly by events such as exposure to niche-specific factors or subsequent genetic changes. In reality both concepts might occur in parallel or in combination.

Irrespective of the way of stem cell-driven tumorigenesis, the presence of such cells in tumour tissue would require either elimination of this critical population or of all dominant clones for successful therapy. Since stem cells play a crucial role in tissue repair and homeostasis, they possess special mechanisms that protect them from lethal damage.

These mechanisms represent an elementary obstacle for the successful therapy of cancer.

In this review, we will discuss the evidence for stem cells in normal lung tissue and lung cancer, their prognostic and predictive significance and novel therapeutic options for targeting these cells.

### Evidence for Stem Cells in the Normal Lung Epithelium

The lung epithelium consists of several different cell types whose composition varies with the anatomical region (Figure 3): The epithelium of the trachea and the proximal airways consists of ciliated cells and Clara-like cells producing secretoglobins, and moreover, a small number of neuroendocrine cells and mucus-producing submucosal



Figure 3. The lung epithelium. A: Tracheobronchial epithelium. The columnar tracheobronchial epithelium contains ciliated cells, goblet cells, basal cells and submucosal glands. Clara cells and neuroendocrine cells are not shown. Basal cells and a variant of Clara cells are considered stem/progenitor cells B: Bronchioalveolar duct junction (BADJ). As shown in rodent models, the junction between the cuboidal terminal bronchiole epithelium and the alveolar epithelium contains bronchioalveolar stem cells (BASC). C: Alveolar epithelium. The alveolar epithelium consists of pneumocytes type I and II and alveolar macrophages. Pneumocytes type II are able to regenerate and differentiate into pneumocytes type I. Microphotographs: Stain: hematoxylin-eosin, magnification:  $\times 200$ , scale bar:  $50 \mu\text{m}$ .

#### Driver Mutations in Lung Adenocarcinoma

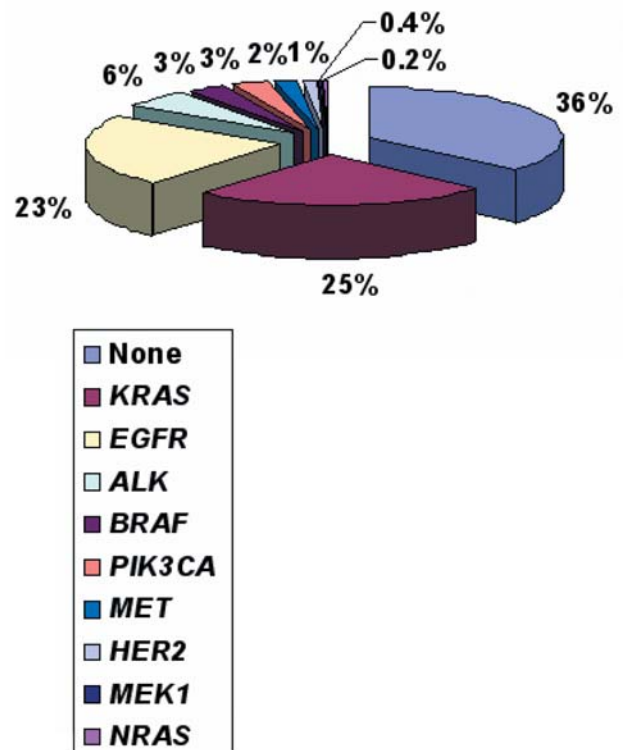


Figure 4. Driver mutations in lung adenocarcinoma. Mutation analysis of 422 patients with lung adenocarcinoma revealed driver mutations in 60% of the cases. *K-ras* (Kirsten rat sarcoma) and *EGFR* (epidermal growth factor receptor-1) mutations represent the most prominent alterations. Other mutations, *ALK* (anaplastic lymphoma kinase) rearrangements and *MET* (Met proto-oncogene) amplifications are present in  $<10\%$  of the tumours. *BRAF*: B-Raf proto-oncogene serine/threonine-protein kinase, *PIK3CA*: phosphatidylinositol-3-kinase, catalytic, alpha polypeptide, *HER2*: epidermal growth factor receptor-2, *MEK1*: dual specificity mitogen-activated protein kinase kinase-1, *NRAS*: neuroblastoma RAS viral oncogene homolog. Adapted from M.G. Kris *et al.*, *J Clin Oncol* 2011 (May 20 Supplement).

glands containing ciliated and basal cells (7). In the more distal airways, Clara cells predominate over ciliated cells and there are more neuroendocrine cells, but no basal cells. The alveolar epithelium consists of pneumocytes type I and pneumocytes type II, producing surfactant factor.

The first evidence for the presence of regenerating cells in the lung epithelium came from rodent injury models in the 1970s (7). Selective damage of specific cell types by inhaled oxidants such as  $\text{NO}_2$  or ozone or intravenous administration of bleomycin revealed a regenerative potential of Clara cells and pneumocytes type II. In the nineties and 2000s, intraperitoneal injection of naphthalene showed that the

regenerative potential of Clara cells is restricted to a specific subpopulation, so called variant Clara cells, which are lacking expression of cytochrome *P450F2*. Additional genetic labeling studies showed that cytokeratin 14- positive basal cells also possess regenerative potential in the proximal airways and give rise to basal cells, Clara cells and ciliated cells (10).

The distal lung epithelium appears to be regenerated by variant Clara cells and *calcitonin gene-related peptide* (*CGRP*)-expressing pulmonary neuroendocrine cells, both located in the neuroendocrine bodies (11). Moreover, in 2005 Bender Kim *et al.* showed that the junction between the cuboidal epithelium of the terminal bronchiole and the alveolar epithelium contains a population of stem cell *antigen-1* (*Sca-1*)<sup>+</sup>, CD31<sup>-</sup>, CD34<sup>+</sup>, CD45<sup>-</sup>, Clara cell antigen (CCA)<sup>+</sup> cells which maintain the terminal bronchiolar epithelium (12). These so-called bronchioalveolar stem cells (BASC) gave rise to Clara-like, pneumocyte type II-like and pneumocyte type I-like cells *in vitro* and showed expansion and differentiation towards the alveolar lineage upon induction of a *Kirsten rat sarcoma* (*k-ras*) mutation *in vivo*.

So far, it is unclear whether these stem and progenitor cells are organized in a hierarchical structure, as known from the hematopoietic system, or represent different types of region-specific stem cells. An analysis of Qualter *et al.* in *epithelial cell adhesion molecule* (*EpCAM*)<sup>+</sup>, *Sca-1*<sup>+</sup>, CD24<sup>dim</sup>, CD31<sup>-</sup>, CD45<sup>-</sup>, CD49<sup>+</sup>, CD104<sup>+</sup> murine lung epithelial cells provided first evidence for a stem cell hierarchy in the lung (13). These cells were able to self-renew in cytokine-supplemented, stroma-free cultures and generated colonies of airway, alveolar and mixed lung epithelial lineages upon co-culture with pulmonary mesenchymal cells.

Although the use of different stem and progenitor cell markers in the various studies hamper an uniform view so far, there is strong evidence that the lung contains various stem and progenitor cells that regenerate specific regions of the pulmonary epithelium and might be organized in a hierarchical structure (14). The characteristics of these putative stem and progenitor cells are summarized in Table I.

## Evidence for Stem Cells in NSCLC

First evidence for the presence of stem cells in NSCLC came from a mouse strain with inducible *k-ras* *G12D* mutation developed by Bender Kim *et al.* (12). In 2005 this group demonstrated an increased expansion and reduced differentiation of BASC upon activation of *k-ras* resulting in an atypical adenomatous hyperplasia. Combination of *k-ras* activation and naphthalene treatment led to an increase in tumour number and areas, strongly indicating that BASC might represent the progenitor cells of pulmonary adenocarcinoma. Subsequent analyses of Xu *et al.* demonstrated that not only BASC, but also pneumocytes

type II and Clara cells in the terminal bronchioles proliferate in response to *k-ras* activation. In this study, only pneumocytes type II progressed to adenocarcinoma (15).

As for human lung cancer cell lines and primary cells derived from lung cancer samples, Ho *et al.* demonstrated the presence of a Hoechst dye 33342 extruding subpopulation, the so-called side population (SP), which accounted for 0.03%-6.1% of the tumour cells (16). In contrast to non-SP cells, this small subpopulation was able to regenerate SP and non-SP cells and showed an increased tumorigenicity in mice. In line with their resistance to multiple cytostatic drugs and enhanced proliferation capacity, these cells showed an elevated expression of various ATP-binding cassette transporters and *human telomerase reverse transcriptase* (*hTERT*).

Eramo *et al.* used the common stem cell antigen CD133 to prospectively isolate human lung cancer cells with stem cell properties. These cells were able to form tumour spheres, showed co-expression of the ATP-binding cassette transporter *breast cancer resistance protein-1* (*BCRP1*) and the ESC transcription factors *octamer binding protein* (*Oct*) 3/4 and *Nanog*. Moreover, these cells were highly tumorigenic in mice, able to regenerate the original tumour histology and resistant to various cytostatic drugs (17). Chen *et al.* reported very similar results for CD133-positive NSCLC cells and moreover showed that these cells lost their stem cell characteristics when the expression of *Oct4* was knocked down by siRNA (18).

Levina *et al.* who used a reverse approach to isolate lung cancer stem cells also reported corresponding findings (19). Exposure of the NSCLC cell line *H460* to cytostatic drugs enriched a Hoechst dye 33342-extruding SP. The antigen profile and functional properties of this SP was basically consistent with the stem cell population described by Eramo *et al.* and Chen *et al.* In addition to previous analyses, Levina *et al.* demonstrated that these cells had an activated  $\beta$ -*Catenin* pathway known to be involved in stem cell self-renewal, and moreover, that tumours originating from these cells show a high expression of angiogenic cytokines and growth factors.

Bertolini *et al.* confirmed an enrichment of CSC after cytostatic treatment *in vivo* (20). When mice carrying lung cancer xenografts were treated with cisplatin, the tumours showed an enrichment of epithelial cells expressing *CD133* and *BCRP1*. These cells shared several functional and molecular properties with the CSC described in previous studies including an extraordinary high tumorigenicity in mice and an increased expression of the transcription factors *Oct3/4* and *Nanog*, adhesion and motility genes and various ATP-binding cassette transporters.

As for human pulmonary CSC, CD133 and *BCRP1* appear to be the most appropriate markers so far. However, a recent study of Curtis *et al.* demonstrated that the phenotype of lung cancer propagating cells varies with the type of genetic alteration present in the tumour (21). Since recent analyses in

Table I. Putative pulmonary stem and progenitor cells. The table shows the characteristics of putative stem/progenitor cells identified in the lung epithelium of rodent models. CK: Cytokeratin, CCSP: Clara cell secretory protein, CYP2F2: cytochrome P450, family 2, subfamily F, polypeptide 2, NEB: neuroendocrine body, PNEC: pulmonary neuroendocrine cells, CGRP: calcitonin gene-related peptide, SP-A, -C, -D: surfactant protein-A, -C, -D, AQP3: aquaporin 3, BASC: bronchioalveolar stem cell, Sca-1: stem cell antigen-1, BADJ: bronchioalveolar duct junction, y: yes, n: no.

Putative stem/progenitor cell	Marker	Niche	Descendant	Self-Renewal	Multi-lineage differentiation	References
Basal cell	CK5 <sup>+</sup> , CK14 <sup>+</sup> , p63 <sup>+</sup>	Conducting airway epithelium	Basal cells, Clara cells, ciliated cells	y	y	Hong <i>et al.</i> , 2004
Variant Clara cell	CCSP <sup>+</sup> , CYP2F2 <sup>-</sup>	NEB	Variant Clara cells, Clara cells, PNEC, ciliated cells	y	y	Hong <i>et al.</i> , 2001
PNEC	CGRP <sup>+</sup>	NEB	PNEC	y	n	
Type II pneumocyte	SP-A <sup>+</sup> , SP-C <sup>+</sup> , SP-D <sup>+</sup> , CD44v6 <sup>+</sup> , AQP3 <sup>+</sup>	Alveoli	Type II pneumocytes, Type I pneumocytes	y	n	Evans <i>et al.</i> , 1975
BASC	CCSP <sup>+</sup> , SP-C <sup>+</sup> , CD34 <sup>+</sup> , Sca-1 <sup>+</sup> , CD45 <sup>-</sup> , CD31 <sup>-</sup>	BADJ	Clara cells, type II pneumocytes	y	y	Bender Kim <i>et al.</i> , 2005

lung adenocarcinoma revealed the presence of driver mutations in up to 87% of all cases, several phenotypes of pulmonary CSC might exist. This suggests that rather functional properties than surface antigens represent appropriate targets for specific therapies against CSC (22, 23).

### Predictive and Prognostic Significance of CSC in Lung Cancer

In recent years, several immunohistochemical and a few microarray studies analyzed the prognostic and predictive significance of CSC antigens and gene signatures in NSCLC.

As for the most common human pulmonary CSC antigens CD133 and *BCRP1*, expression frequencies between 15%-have been 63% and 15%-51%, respectively, were reported (24-29). While our group and Salnikov *et al.* found no prognostic significance of CD133 and *BCRP1* expression in 133 stage I/II and 88 stage I-III patients who underwent complete pulmonary resection, two Asian studies in 145 stage I NSCLCs and 177 stage I AC patients found an up to 9.3-fold increased risk of postoperative disease recurrence in CD133/*BCRP1* and CD133-positive cases, respectively (25, 28-30).

In patients with locally-advanced or metastatic disease who received platinum-based therapy studies of Yoh *et al.* and Ota *et al.* demonstrated a predictive and prognostic significance of *BCRP1* (26, 27): while Yoh *et al.* reported a higher response rate (44% vs. 24%,  $p=0.08$ ) of patients with *BCRP1* negative tumours and a 2.8-fold ( $p=0.001$ ) and 1.8-fold ( $p=0.08$ ) reduced risk of relapse and disease-related mortality, respectively, Ota *et al.* found no association between *BCRP1* expression and response to therapy or progression-free survival (PFS), but an association with a significantly reduced median survival time (7.8 vs. 14.9 months,  $p=0.03$ ) in *BCRP1*-positive cases.

Also in neoadjuvant treatment, a recent study in 50 patients who received preoperative chemoradiotherapy, demonstrated that the 5-year overall survival (OS) was worse in patients with CD133 or *ALDH1* expression (45% vs. 90%,  $p=0.042$ ) (31).

Authors who used other stem cell markers such as the basal cell antigen cytokeratin 14 or the ESC transcription factor *Oct4*, found a 1.6-fold increased risk of disease-related mortality in two large datasets of 399 ( $p=0.003$ ) and 505 stage I-IV patients ( $p=0.004$ ) and a 2.2-fold increased risk of disease-related mortality in a collective of 112 operated stage IB-IIIa AC patients ( $p=0.015$ ), respectively (32, 33).

Onaitis *et al.* applied a 10-gene progenitor cell signature derived from a transgenic mouse model to a dataset of resected human AC patients and found a 3.5-fold increased risk of disease-related mortality ( $p=0.0004$ ) when this gene signature was expressed (34).

All in all, these studies provide strong evidence that the presence of tumour cells expressing antigens or gene signatures associated with stem cells might play a prognostic and predictive role in NSCLC.

### Significance of Drug Resistance in Current Lung Cancer Therapy

In recent years, increasing efforts were undertaken to identify therapy responders in advance. The presence of specific targets and the absence of resistance mechanisms represent the two pillars of tailored therapy.

Overexpression of key enzymes of the folate synthesis, particularly *thymidylate synthase* (*TS*), and components of the mitotic spindle apparatus such as *tubulin beta III* (*TUBB3*) show a high level of evidence for an association with response to folate antagonist such as pemetrexed and microtubule-inhibiting taxanes, respectively (35-37).

Particularly targeted therapies with tyrosine kinase inhibitors (TKI) such as erlotinib, gefitinib and crizotinib rendered impressive results when response predictors were considered. While conventional chemotherapy shows response rates of maximal 30% in the first-line treatment and about 10% in subsequent therapy lines, treatment with the *epidermal growth factor receptor (EGFR)*-TKI erlotinib or gefitinib or the *echinoderm microtubule-associated protein-like 4-anaplastic lymphoma kinase (EML4-ALK)*-TKI crizotinib in patients with activating *EGFR* mutations or *EML4-ALK* rearrangements shows response rates of  $\geq 70\%$  and 57%, respectively (38, 39).

Also DNA repair proteins such as *excision repair cross-complementation group 1 (ERCC1)*, *breast cancer 1, early onset (BRCA1)* and *ribonucleotide reductase 1 (RRM1)* gained growing significance for tailoring cisplatin or gemcitabine-containing therapy regimens. Operated patients who received cisplatin-based, adjuvant treatment showed a significantly longer survival, when their tumours were *ERCC1*-negative. Also in advanced disease, lower *ERCC1* mRNA levels were highly predictive for longer survival after cisplatin-based treatment (40-42). In metastatic disease, low *BRCA1* mRNA levels predicted a higher response rate, longer PFS and median OS towards platinum-containing second-line therapies (43). As for gemcitabine, studies of Bepler *et al.* demonstrated that patients with low *RRM1* mRNA levels experienced better response and longer PFS and OS to this antimetabolite (44, 45). However, none of these candidate biomarkers have been established to date since large, prospective trials are just getting under way.

So far, the association between these putative biomarkers and CSC is widely unknown. The findings of Ota *et al.* who demonstrated a correlation between protein expression of *ERCC1* and the stem cell antigen *BCRP1* ( $p=0.012$ ) and the results of Salnikov *et al.* who demonstrated a correlation between protein expression of *TS* and the stem cell antigen CD133 ( $p<0.01$ ), indicate that tumour cells with predominant expression of these enzymes might bear stem cell characteristics (27, 29).

## Mechanisms of Drug Resistance in CSC

Hitherto, little heed has been taken of the specific drug resistance mechanisms of CSC in antineoplastic treatment. The ability of asymmetric cell division, the slow division kinetics or even quiescence and the expression of several drug efflux pumps and DNA repair proteins represent the major escape mechanisms of stem cells (46-48).

In 2010 Pine *et al.* demonstrated for the first time that a small subpopulation of lung cancer cells which were present in commercially available cell lines and primary tumour cell cultures was able to undergo asymmetric cell division: While the differentiation markers *prosurfactant protein C* and pan-cytokeratins were passed to one daughter cell, the stem cell

marker CD133 was passed to the other and exclusively characterized cells with tumour repopulating capability (49). A subsequent study of Dey-Guha *et al.* in breast cancer cell lines showed that asymmetric division generated slowly cycling  $G_0$ -like cells which were enriched in tumour tissue after neoadjuvant therapy (50). The presence of such a quiescent and drug-resistant subpopulation has also been shown in lung cancer (51). Several pathways and molecules such as *v-akt murine thymoma viral oncogene homolog 1 (AKT1)*, *insulin-like growth factor receptor-1 (IGF-1R)*, *protein kinase C (PKC)*, *aurora kinase A (AURAKA)*, *Notch*, *phosphatidylinositol 3-kinase (PI3K)*, *p53* or *phosphatase and tensin homolog deleted on chromosome 10 (PTEN)* were found to be involved in the regulation of division symmetry and quiescence of stem cells (50-53). As most classical cytostatics require cell division for being effective, quiescence or slow division kinetics protects tumour cells against lethal DNA damage. Moreover, novel molecular drugs such as AKT-TKI might rather support than suppress CSC as they favor asymmetric cell division and the generation of slowly-cycling, drug-resistant tumour cells (50, 54).

Besides division symmetry and slow division kinetics, CSC show an increased expression of DNA repair proteins and drug efflux transporters. Salnikov *et al.* and Ota *et al.* demonstrated that overexpression of *TS* and *ERCC1* which might predict resistance against pemetrexed and cisplatin, respectively, correlated with the expression of the stem cell antigens CD133 and *BCRP1* (27, 29). Moreover, several other proteins involved in detoxification and drug resistance such as *glutathione-S-transferase*, the drug transporter *ABCC2 (multidrug resistance 1 [MDR1], p-glycoprotein [p-gp])*, *metallothionein*, *catalase* or *O<sup>6</sup>-methylguanine-DNA methyltransferase* showed an overexpression in CD133-positive cells. Ho *et al.* reported an increased expression of the drug transporters *ABCG2 (BCRP1)*, *ABCA2*, *MDR1* and *ABCC1 (multidrug resistance protein-1 [MRP1])* in the SP of various NSCLC cell lines (16). In other tumour entities, *glutathione-S-transferase* and *MRP1* were shown to confer resistance to gemcitabine, while *MDR1* was associated with resistance to paclitaxel and *BCRP1* with resistance to the *EGFR*-TKI gefitinib (55-57).

All in all, CSC possess several mechanisms to overcome irreversible damage by cytostatic drugs. These mechanisms have neither been studied sufficiently nor have they been considered in cancer therapy so far.

## Potential Targets and Approaches for CSC-directed Therapy

The ability of asymmetric cell division and the generation of slowly cycling or quiescent progenies represent important escape mechanism of stem cells. Thus, pivotal molecules and mechanisms involved in the regulation of division symmetry might provide one of the Achilles' heels to target these cells in NSCLC.

Table II. Potential targets and therapeutic agents against cancer stem cells (CSC). The table shows a selection of substances which are currently tested in clinical trials including lung cancer (ClinicalTrials.gov). The listed target molecules and pathways are involved in the regulation of CSC. PARP1: poly(ADP-ribose)-polymerase 1, MDR1: multidrug-resistance 1, Akt: v-akt murine thymoma viral oncogene, PI3K: phosphatidylinositol 3-kinase, mTOR: mechanistic target of rapamycin (serine/threonine kinase), MAPK: mitogen-activated protein kinase, \*: selection.

Target	Function	Agent	Phase	ClinicalTrials.gov Identifier
<i>PARP1</i>	Single strand break damage repair	Iniparib (BSI-201)	III, II	NCT01082549, NCT01086254
<i>Telomerase</i>	Self-renewal	GV 1001 telomerase peptide, Imetelstat (GRN163L)	II, III II	NCT00509457, NCT01579188 NCT01137968
<i>MDR1</i>	Drug resistance	Tariquidar (XR9576)	III, III	NCT00042315, NCT00042302
<i>Wnt/β-Catenin</i>	Self-renewal	PRI-724	I	NCT01302405
<i>Hedgehog</i>	Self-renewal	Vismodegib (GDC-0449)	I, I	NCT00968981, NCT00607724
		Saridegib (IPI-926)	I	NCT00761696
		LEQ506	I	NCT01106508
		BMS-833923	I, I	NCT00670189, NCT01413906
		PF-04449913	I	NCT01286467
<i>Notch</i>	Self-renewal	RO4929097	I, II, I	NCT01193881, NCT01193868, NCT01131234
		LY2606368	I	NCT01115790
<i>Akt</i>	Self-renewal	MK-2206	I, II	NCT01147211, NCT01294306*
<i>PI3K</i>	Self-renewal	BKM-120	I/II, I, II, II	NCT01487265, NCT01570296, NCT01297491, NCT01501604
<i>mTOR</i>	Self-renewal	Everolimus (RAD001)	I, I	NCT00457119, NCT00456833*
<i>MAPK</i>	Self-renewal	Selumetinib (AZD6244)	I, II	NCT01586624, NCT01229150*
		PD-0325901	I, II	NCT01347866, NCT00174369
		MSC1936369B	I, I	NCT01357330, NCT01390818*
		GSK1120212	I, II	NCT00955773, NCT01362296
		LY2228820	I	NCT1393990
		LY3007113	I	NCT01463631

The *Notch*, *Wnt* and *Hedgehog* pathway represent key mechanisms of stem cell regulation (58). In 2008 Tsao *et al.* demonstrated that disruption of *Notch* signaling by inhibition of gamma-secretase cleavage of *Notch* receptors dramatically expanded the population of distal pulmonary progenitors and prevented formation of proximal structures in murine embryonic lungs (59). Two years later, Sullivan *et al.* showed that the proliferation and clonogenicity of self-renewing *ALDH*-positive CSC in primary human NSCLC cell lines was significantly decreased by treatment with either a  $\gamma$ -secretase inhibitor or stable expression of shRNA against *Notch3* (60). A large study of Westhoff *et al.* in tumour tissue of more than 400 NSCLC patients demonstrated an activated *Notch* pathway in about one third of the cases and a significantly worse survival of these patients when concomitant *p53* mutations were lacking (61). Altogether, these studies indicate not only a pivotal pathogenetic role, but also a prognostic relevance of *Notch* signaling in NSCLC.

*Wnt* signaling represents another pathway that is critically involved in the regulation of CSC (62). Levina *et al.* demonstrated that NSCLC cells surviving cytostatic treatment show stem cell properties and an activated *Wnt* pathway (18). Similar findings were reported by Teng *et al.*

in cisplatin-selected A549 NSCLC cells: Activation of *Wnt* signaling led to a significantly enhanced proliferation, colony formation, migration and drug resistance (63). In a murine model, Zhang *et al.* demonstrated that *Gata6*-regulated *Wnt* signaling controls the balance between progenitor cell expansion and epithelial differentiation required for both, lung development and regeneration (64).

As for *Hedgehog* signaling, recent studies of Shi *et al.* and Tian *et al.* in a pulmonary LCC and an AC cell line showed that the proliferation of the SP was significantly impaired by inhibition of the *Hedgehog* pathway with Cyclopamine or Vismodegib (GDC-0449), respectively (65, 66).

Another molecule which has repeatedly been reported to be involved in the regulation of CSC in NSCLC is the tumour suppressor gene *PTEN*. In a murine model with constitutive loss of *PTEN*, epithelial cells residing in the bronchioalveolar duct junction underwent proliferation and formed uniform masses (67). Similar results were derived from a previous study analyzing a murine model with bronchioalveolar epithelium-specific null mutation of *PTEN*: The numbers of BASC, the putative progenitors of pulmonary adenocarcinoma, were likewise increased (68). In addition, *PTEN* is an inhibitor of the *PI3K/AKT/mTOR* (mammalian target of rapamycin) signal transduction axis

and loss of function leads to an activation of these molecules which have all been shown to be involved in the maintenance of CSC (50, 52, 69).

Currently, several phase I and II studies testing inhibitors of the *Wnt*, *Notch* and *Hedgehog* pathway as well as the *PI3K/AKT/mTOR* signal transduction axis are in progress (Table II). However, it remains to be elucidated whether targeting these pathways and molecules will lead to an increased therapeutic efficacy or even have an opposite effect in lung cancer (54).

As for drug efflux transporters, pharmacodynamic studies with the *MDR1* inhibitors CBT-1(R) and tariquidar showed an increased uptake of the radionuclid  $^{99m}\text{Tc}$ -sestamibi, a substrate of *MDR1* (70-72). However, two phase III studies in stage IIIB/IV NSCLC combining tariquidar with first line single-agent vinorelbine or a carboplatin/paclitaxel doublet showed no advantage and had to be terminated prematurely (ClinicalTrials.gov ID NCT00042315, NCT00042302). Recent *in vitro* and *in vivo* studies analyzing the effect of TKI such as axitinib, gefitinib, vandetanib, pelitinib or crizotinib on the function of *ABCG2* (*BCRP1*) and *ABCB1* (*MDR1*) demonstrated an inhibition of these drug efflux transporters and an enhanced efficacy of cytostatic drugs in multidrug-resistant cell lines and murine xenograft models (51, 73, 74). These TKI might be worthwhile combination partners for specific cytostatics such as anthracyclines, *Vinca* alkaloids, taxanes or epipodophyllotoxins in subsequent clinical trials.

A recent analysis of Bartucci *et al.* in CSC derived from primary NSCLC cultures demonstrated a strong activation of the *DNA damage checkpoint protein kinase 1* (*Chk1*) upon exposure to cisplatin, gemcitabine or paclitaxel (75). In contrast, differentiated, chemosensitive NSCLC cells showed only a weak activation of *Chk1*. When the *Chk1* inhibitor AZD7762 and chemotherapy were co-administered tumour growth *in vivo* was suppressed, whereas chemotherapy alone was scarcely effective. Thus, AZD7761 might represent an interesting agent for future clinical trials directing CSC in NSCLC.

*Poly [ADP-ribose] polymerase 1* (*PARP1*), *telomerase* and *mitogen-activated protein kinases* (*MAPK*) represent other promising targets for the eradication of CSC. In patients with squamous NSCLC, low expression of the DNA repair protein *PARP1* showed a significant association with an increased chemotherapeutic efficacy (76). Although analyses on *PARP1* expression in pulmonary CSC are so far lacking, recent data on the pivotal significance of this enzyme in the regulation of pluripotency and differentiation of murine ESC suggest a general role in stem cells (77). As for human telomerase, Ho *et al.* demonstrated an increased expression of *hTERT* in the SP of various NSCLC cell lines and primary NSCLC cells corresponding to the enhanced proliferation capacity of this subpopulation (19, 78, 79). Also *MAPK* is involved in the regulation of pulmonary stem cell proliferation (80).

However, the efficacy of *MAPK* inhibitors might depend on the type of *MAPK* that is blocked: While inhibition of *rat fibrosarcoma/mitogen-activated protein kinase kinase 1/2* (*Raf/ERK1/2*)-mediated downstream signaling might impair the self-renewal of NSCLC cells, blocking of the *p38 MAPK $\alpha$*  might have the opposite effect: In a murine model of Ventura *et al.*, inhibition of *p38 MAPK $\alpha$*  generated an immature and hyperproliferative lung epithelium which was highly susceptible to *k-ras*-induced tumorigenesis (80-82).

Presently, several early phase studies analyzing the efficacy of *MAPK* inhibitors in lung cancer and various phase II and III trials with the *PARP1* inhibitor Iniparib and agents against telomerase are in progress (Table II). High throughput screening methods such as those developed by Gupta *et al.* for breast cancer stem cells, might represent an effective approach for the identification of novel compounds against CSC (83).

All in all, consideration of CSC and their specific mechanisms of drug resistance might pave novel ways for more effective treatment and long-lasting therapeutic success in lung cancer.

## Future Perspectives

Although analyses in lung cancer and various other tumour entities provide strong evidence for a pivotal role of CSC in drug resistance and relapse, several questions remain to be answered.

So far, various markers and methods have been used to identify and isolate cancer cells with self-renewing and multilineage differentiation capability. However, comparative functional and molecular analyses of these differentially isolated cells are so far lacking. It is unclear, whether they represent the same cell population, a hierarchy of stem and progenitor cells or different cell types which share some common functional and molecular properties. Moreover, the origin and development of these cells is not yet understood: Do CSC derive from a neoplastic stem cell or a neoplastic tumour cell which acquired stem cell characteristics? Does their development require specific mutations that occur in an orchestrated manner or does it represent a clonal selection of randomly mutated cells by microenvironmental cues? The identification of specific markers and gene signatures of various differentiation stages might represent a first step towards better understanding of pulmonary stem cells. However, established differentiation assays are the prerequisite for such analyses and are yet lacking.

Moreover, several fundamental results on lung stem cells derive from murine models. So far, it is uncertain, whether these findings can be transferred onto human conditions. The common murine stem cell marker *Sca-1*, for instance, has no human homologue. In addition, the life expectancy of mice and men is very different and might affect the proliferation

capacities of murine and human stem cells. As for transplantation studies, the growth behaviour of human xenografts strongly varies under the murine background and thus, results from *in vivo* studies might in a large part reflect the experimental conditions and less the genuine behaviour of the human tumour cells.

The development of novel imaging techniques to identify and track stem cells in their natural niche might help to better understand their significance in drug resistance, tumour regeneration and metastasis (84-86). However, these methods are still in early stages of development.

Altogether, several *in vitro* and *in vivo* studies provide strong evidence for the presence of drug-resistant and highly regenerative tumour cells in NSCLC. Eradication of these cells might contribute to better response and prolonged survival in lung cancer.

## References

- 1 American Cancer Society: Cancer Statistics 2008. [http://www.cancer.org/docroot/STT/stt\\_0.asp](http://www.cancer.org/docroot/STT/stt_0.asp).
- 2 Goeckenjan G, Sitter H, Thomas M, Branscheid D, Flentje M, Griesinger F, Niederle N, Stuschke M, Blum T, Deppermann KM, Ficker JH, Freitag L, Lübke AS, Reinhold T, Späth-Schwalbe E, Ukena D, Wickert M, Wolf M, Andreas S, Auberger T, Baum RP, Baysal B, Beuth J, Bickeböller H, Böcking A, Bohle RM, Brüske I, Burghuber O, Dickgreber N, Diederich S, Dienemann H, Eberhardt W, Eggeling S, Fink T, Fischer B, Franke M, Friedel G, Gauler T, Gütz S, Hautmann H, Hellmann A, Hellwig D, Herth F, Heussel CP, Hilbe W, Hoffmeyer F, Horneber M, Huber RM, Hübner J, Kauczor HU, Kirchbacher K, Kirsten D, Kraus T, Lang SM, Martens U, Mohn-Staudner A, Müller KM, Müller-Nordhorn J, Nowak D, Ochmann U, Passlick B, Petersen I, Pirker R, Pokrajac B, Reck M, Riha S, Rube C, Schmittel A, Schönfeld N, Schütte W, Serke M, Stamatis G, Steingraber M, Steins M, Stoelben E, Swoboda L, Teschler H, Tessen HW, Weber M, Werner A, Wichmann HE, Irlinger Wimmer E, Witt C, Worth H; German Respiratory Society; German Cancer Society: Prevention, diagnosis, therapy, and follow-up of lung cancer: interdisciplinary guideline of the German Respiratory Society and the German Cancer Society. *Pneumologie* 65: 39-59, 2011.
- 3 Fraire AE, Roggli VL, Vollmer RT, Greenberg SD, McGavran MH, Spjut HJ and Yesner R: Lung cancer heterogeneity. Prognostic implications. *Cancer* 60: 3701-375, 1987.
- 4 Reya T, Morrison SJ, Clarke MF, Weissman IL: Stem cells, cancer, and cancer stem cells. *Nature* 414: 105-111, 2001.
- 5 Sullivan JP, Minna JD and Shay JW: Evidence for self-renewing lung cancer stem cells and their implications in tumour initiation, progression, and targeted therapy. *Cancer Metastasis Rev* 29: 61-72, 2010.
- 6 Otto WR: Lung epithelial stem cells. *J Pathol* 197: 527-535, 2002.
- 7 Rawlins EL and Hogan BLM: Epithelial stem cells of the lung: privileged few or opportunities for many? *Development* 133: 2455-2465, 2006.
- 8 Maenhaut C, Dumont JE, Roger PP and van Staveren WCG: Cancer stem cells: a reality, a myth, a fuzzy concept or a misnomer? An analysis. *Carcinogenesis* 31: 149-158, 2010.
- 9 Adams JM and Strasser A: Is tumor growth sustained by rare cancer stem cells or dominant clones? *Cancer Res* 68: 4018-4021, 2008.
- 10 Hong KU, Reynolds SD, Watkins S, Fuchs E and Stripp BR: Basal cells are a multipotent progenitor capable of renewing the bronchial epithelium. *Am J Pathol* 164: 577-588, 2004.
- 11 Reynolds SD, Giangreco A, Power JHT and Stripp BR: Neuroepithelial bodies of pulmonary airways serve as a reservoir of progenitor cells capable of epithelial regeneration. *Am J Pathol* 156: 269-278, 2000.
- 12 Bender Kim CF, Jackson EL, Woolfenden AE, Lawrence S, Babar I, Vogel S, Crowley D, Bronson RT and Jacks T: Identification of bronchioalveolar stem cells in normal lung and lung cancer. *Cell* 121: 823-835, 2005.
- 13 McQuilter JL, Yuen K, Williams B and Bertoncello I: Evidence of an epithelial stem/progenitor cell hierarchy in the adult mouse lung. *Proc Natl Acad Sci USA* 107: 1414-1419, 2010.
- 14 Chen H, Matsumoto K, Brockway BL, Rackley CR, Liang J, Lee JH, Jiang D, Noble PW, Randell SH, Kim CF and Stripp BR: Airway epithelial progenitors are region-specific and show differential response to bleomycin-induced lung injury. *Stem Cells* 2012 Jun 13. doi: 10.1002/stem.1150. [Epub ahead of print]
- 15 Xu X, Rock JR, Lu Y, Futtner C, Schwab B, Guinney J, Hogan BLM and Onaitis MW: Evidence for type II cells as cells of origin of K-Ras-induced distal lung adenocarcinoma. *Proc Natl Acad Sci USA* 109: 4910-4915, 2012.
- 16 Ho MM, Ng AV, Lam S and Hung JY: Side population in human lung cancer cell lines and tumors is enriched with stem-like cancer cells. *Cancer Res* 67: 4827-4833, 2007.
- 17 Eramo A, Lotti F, Sette G, Pilozzi E, Biffoni M, Di Virgilio A, Conticello C, Ruco L, Peschle C and De Maria R: Identification and expansion of the tumorigenic lung cancer stem cell population. *Cell Death Differ* 15: 504-514, 2008.
- 18 Chen Y-C, Hsu H-S, Chen Y-W, Tsai T-H, How C-K, Wang C-Y, Hung S-C, Chang Y-L, Tsai M-L, Lee Y-Y, Ku H-H and Chiou S-H: Oct-4 expression maintained cancer stem-like properties in lung cancer-derived CD133-positive cells. *PLoS One* 3: e2637, 2008.
- 19 Levina V, Marrangoni AM, DeMarco R, Gorelik E and Lokshin AE: Drug-selected human lung cancer stem cells: cytokine network, tumorigenic and metastatic properties. *PLoS One* 3: e3077, 2008.
- 20 Bertolini G, Roz L, Perego P, Tortoreto M, Fontanella E, Gatti L, Pratesi G, Fabbri A, Andriani F, Tinelli S, Roz E, Caserini U, Lo Vullo S, Camerini T, Mariani L, Delia D, Calabro E, Pastorino U and Sozzi G: Highly tumorigenic lung cancer CD133+ cells display stem-like features and are spared by cisplatin treatment. *Proc Natl Acad Sci USA* 106: 16281-16286, 2009.
- 21 Curtis SJ, Sinkevicius KW, Li D, Lau AN, Roach RR, Zamponi R, Woolfenden AE, Kirsch DG, Wong K-K and Kim CF: Primary tumor genotype is an important determinant in identification of lung cancer propagating cells. *Cell Stem Cell* 7: 127-133, 2010.
- 22 Kris MG, Johnson BE, Kwiatkowski DJ, Iafrate AJ, Wistuba II, Aronson SL, Engelman JA, Shyr Y, Khuri FR, Rudin CM, Garon EB, Pao W, Schiller JH, Haura EB, Shirai K, Giaccone G, Berry LD, Kugler K, Minna JD and Bunn PA: Identification of driver mutations in tumor specimens from 1,000 patients with lung adenocarcinoma: The NCI's Lung Cancer Mutation Consortium (LCMC). *J Clin Oncol* 29 Suppl. (May 20 Supplement): CRA7506, 2011.

- 23 Ding L, Getz G, Wheeler DA, Mardis ER, McLellan MD, Cibulskis K, Sougnez C, Greulich H, Muzny DM, Morgan MB, Fulton L, Fulton RS, Zhang Q, Wendl MC, Lawrence MS, Larson DE, Chen K, Dooling DJ, Sabo A, Hawes AC, Shen H, Jhangiani SN, Lewis LR, Hall O, Zhu Y, Mathew T, Ren Y, Yao J, Scherer SE, Clerc K, Metcalf GA, Ng B, Milosavljevic A, Gonzalez-Garay ML, Osborne JR, Meyer R, Shi X, Tang Y, Koboldt DC, Lin L, Abbott R, Miner TL, Pohl C, Fewell G, Haiepek C, Schmidt H, Dunford-Shore BH, Kraja A, Crosby SD, Sawyer CS, Vickery T, Sander S, Robinson J, Winckler W, Baldwin J, Chirieac LR, Dutt A, Fennell T, Hanna M, Johnson BE, Onofrio RC, Thomas RK, Tonon G, Weir BA, Zhao X, Ziaugra L, Zody MC, Giordano T, Orringer MB, Roth JA, Spitz MR, Wistuba II, Ozenberger B, Good PJ, Chang AC, Beer DG, Watson MA, Ladanyi M, Broderick S, Yoshizawa A, Travis WD, Pao W, Province MA, Weinstock GM, Varmus HE, Gabriel SB, Lander ES, Gibbs RA, Meyerson M and Wilson RK: Somatic mutations affect key pathways in lung adenocarcinoma. *Nature* 455: 1069-1075, 2008.
- 24 Moreira AL, Gonen M, Reikhtman N, Downey RJ: Progenitor stem cell marker expression by pulmonary carcinomas. *Mod Pathol* 23: 889-895, 2010.
- 25 Herpel E, Jensen K, Muley T, Warth A, Schnabel PA, Meister M, Herth FJF, Dienemann H, Thomas M and Gottschling S: The cancer stem cell antigens CD133, BCRP1/ABCG2 and CD117/c-KIT are not associated with prognosis in resected early-stage non-small cell lung cancer. *Anticancer Res* 31: 4491-4500, 2011.
- 26 Yoh K, Ishii G, Yokose T, Minegishi Y, Tsuta K, Goto K, Nishiwaki Y, Kodama T, Suga M and Ochiai A: Breast cancer resistance protein impacts clinical outcome in platinum-based chemotherapy for advanced non-small cell lung cancer. *Clin Cancer Res* 10: 1691-1697, 2004.
- 27 Ota S, Ishii G, Goto K, Kubota K, Kim YH, Kojika M, Murata Y, Yamazaki M, Nishiwaki Y, Eguchi K and Ochiai A: Immunohistochemical expression of BCRP1 and ERCC1 in biopsy specimen predicts survival in advanced non-small-cell lung cancer treated with cisplatin-based chemotherapy. *Lung Cancer* 64: 98-104, 2009.
- 28 Li F, Zeng H and Ying K: The combination of stem cell markers CD133 and ABCG2 predicts relapse in stage I non-small cell lung carcinomas. *Med Oncol* 28: 1458-1462, 2011.
- 29 Salnikow AV, Gladkich J, Moldenhauer G, Volm M, Mattern J and Herr I: CD133 is indicative for a resistance phenotype but does not represent a prognostic marker for survival of non-small cell lung cancer patients. *Int J Cancer* 126: 950-958, 2010.
- 30 Woo T, Okudela K, Mitsui H, Yazawa T, Ogawa N, Tajiri M, Yamamoto T, Rino Y, Kitamura H and Masuda M: Prognostic value of CD133 expression in stage I lung adenocarcinomas. *Int J Clin Exp Pathol* 4: 32-42, 2010.
- 31 Shien K, Toyooka S, Ichimura K, Soh J, Furukawa M, Maki Y, Muraoka T, Tanaka N, Ueno T, Asano H, Tsukuda K, Yamane M, Oto T, Kiura K and Miyoshi S: Prognostic impact of cancer stem cell-related markers in non-small cell lung cancer patients treated with induction chemoradiotherapy. *Lung Cancer* 77: 162-167, 2012.
- 32 Ooi AT, Mah V, Nickerson DW, Gilbert JL, Ha VL, Hegab AE, Horvath S, Alavi M, Maresh EL, Chia D, Gower AC, Lenburg ME, Spira A, Solis LM, Wistuba II, Walser TC, Wallace WD, Dubinett SM, Goodglick L and Gomperts BN: Presence of a putative tumor-initiating progenitor cell population predicts poor prognosis in smokers with non-small cell lung cancer. *Cancer Res* 70: 6639-6648, 2010.
- 33 Zhang X, Han B, Huang J, Zheng B, Geng Q, Aziz F and Dong Q: Prognostic significance of OCT4 expression in adenocarcinoma of the lung. *Jpn J Clin Oncol* 40: 961-966, 2010.
- 34 Onaitis M, D'Amico TA, Clark CP, Guinney J, Harpole DH and Rawlins EL: A 10-gene progenitor cell signature predicts poor prognosis in lung adenocarcinoma. *Ann Thorac Surg* 91: 1046-1050, 2011.
- 35 Eismann U, Oberschmidt O, Ehnert M, Fleeth J, Lüdtkke FE, Struck S, Schulz L, Blatter J, Lahn MM, Ma D, Niyikiza C, Paoletti P and Hanauske AR: Pemetrexed: mRNA expression of the target genes TS, GARFT and DHFR correlates with the *in vitro* chemosensitivity of human solid tumors. *Int J Clin Pharmacol Ther* 43: 567-569, 2005.
- 36 Reiman T, Lai R, Veillard AS, Paris E, Soria JC, Rosell R, Taron M, Graziano S, Kratzke R, Seymour L, Shepherd FA, Pignon JP, Sève P; LACE-Bio Group: Cross-validation study of class III beta-tubulin as a predictive marker for benefit from adjuvant chemotherapy in resected non-small-cell lung cancer: analysis of four randomized trials. *Ann Oncol* 23: 86-93, 2012.
- 37 Kaira K, Takahashi T, Murakami H, Shukuya T, Kenmotsu H, Ono A, Naito T, Tsuya A, Nakamura Y, Endo M, Kondo H, Nakajima T and Yamamoto N: The role of  $\beta$ III-tubulin in non-small cell lung cancer patients treated by taxane-based chemotherapy. *Int J Clin Oncol* 2012 Feb 23. doi: 10.1007/s10147-012-0386-8. [Epub ahead of print]
- 38 Rosell R, Moran T, Queralt C, Porta R, Cardenal F, Camps C, Majem M, Lopez-Vivanco G, Isla D, Provencio M, Insa A, Massuti B, Gonzalez-Larriba JL, Paz-Ares L, Bover I, Garcia-Campelo R, Moreno MA, Catot S, Rolfo C, Reguart N, Palmero R, Sánchez JM, Bastus R, Mayo C, Bertran-Alamillo J, Molina M, Sanchez JJ, Taron M; Spanish Lung Cancer Group: Screening for epidermal growth factor receptor mutations in lung cancer. *N Engl J Med* 361: 958-967, 2009.
- 39 Kwak EL, Bang YJ, Camidge DR, Shaw AT, Solomon B, Maki RG, Ou SH, Dezube BJ, Jänne PA, Costa DB, Varella-Garcia M, Kim WH, Lynch TJ, Fidias P, Stubbs H, Engelman JA, Sequist LV, Tan W, Gandhi L, Mino-Kenudson M, Wei GC, Shreeve SM, Ratain MJ, Settleman J, Christensen JG, Haber DA, Wilner K, Salgia R, Shapiro GI, Clark JW and Iafrate AJ: Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 363: 1693-1703, 2010.
- 40 Olaussen KA, Dunant A, Fouret P, Brambilla E, André F, Haddad V, Taranchon E, Filipits M, Pirker R, Popper HH, Stahel R, Sabatier L, Pignon JP, Tursz T, Le Chevalier T, Soria JC; IALT Bio Investigators: DNA repair by ERCC1 in non-small-cell lung cancer and cisplatin-based adjuvant chemotherapy. *N Engl J Med* 355: 983-991, 2006.
- 41 Ceppi P, Volante M, Novello S, Rapa I, Danenberg KD, Danenberg PV, Cambieri A, Selvaggi G, Saviozzi S, Calogero R, Papotti M and Scagliotti GV: ERCC1 and RRM1 gene expressions but not EGFR are predictive of shorter survival in advanced non-small-cell lung cancer treated with cisplatin and gemcitabine. *Ann Oncol* 17: 1818-1825, 2006.
- 42 Lord RV, Brabender J, Gandara D, Alberola V, Camps C, Domine M, Cardenal F, Sánchez JM, Gumerlock PH, Tarón M, Sánchez JJ, Danenberg PV and Rosell R: Low ERCC1 expression correlates with prolonged survival after cisplatin plus gemcitabine chemotherapy in non-small cell lung cancer. *Clin Cancer Res* 8: 2286-2291, 2002.

- 43 Papadaki C, Sfakianaki M, Ioannidis G, Lagoudaki E, Trypaki M, Tryfonidis K, Mavroudis D, Stathopoulos E, Georgoulas V and Souglakos J: ERCC1 and BRCA1 mRNA expression levels in the primary tumor could predict the effectiveness of the second-line cisplatin-based chemotherapy in pretreated patients with metastatic non-small cell lung cancer. *J Thorac Oncol* 7: 663-671, 2012.
- 44 Bepler G, Kusmartseva I, Sharma S, Gautam A, Cantor A, Sharma A and Simon G: RRM1 modulated *in vitro* and *in vivo* efficacy of gemcitabine and platinum in non-small-cell lung cancer. *J Clin Oncol* 24: 4731-4737, 2006.
- 45 Simon GR, Schell MJ, Begum M, Kim J, Chiappori A, Haura E, Antonia S and Bepler G: Preliminary indication of survival benefit from ERCC1 and RRM1-tailored chemotherapy in patients with advanced nonsmall cell lung cancer: Evidence from an individual patient analysis. *Cancer* 118: 2525-2531, 2012.
- 46 Tomasetti C and Levy D: Role of symmetric and asymmetric division of stem cells in developing drug resistance. *Proc Natl Acad Sci USA* 107: 16766-16771, 2010.
- 47 Lord CJ and Ashworth A: The DNA damage response and cancer therapy. *Nature* 481: 287-294, 2012.
- 48 Saini V and Shoemaker RH: Potential for therapeutic targeting of tumor stem cells. *Cancer Sci* 101: 16-21, 2010.
- 49 Pine SR, Ryan BM, Vartikovsky L, Robles AI and Harris CC: Microenvironmental modulation of asymmetric cell division in human lung cancer cells. *Proc Natl Acad Sci USA* 107: 2195-2200, 2010.
- 50 Dey-Guha I, Wolfer A, Yeh AC, Albeck JG, Darp R, Leon E, Wulfeuhle J, Petricoin III EF, Wittner BS and Ramaswamy S: Asymmetric cancer cell division regulated by AKT. *Proc Natl Acad Sci USA* 108: 12845-12850, 2011.
- 51 Sharma SV, Lee DY, Li B, Quinlan MP, Takahashi F, Maheswaran S, McDermott U, Azizia N, Zou L, Fischbach MA, Wong KK, Brandstetter K, Wittner B, Ramaswamy S, Classon M and Settleman J: A chromatin-mediated reversible drug tolerant state in cancer cell subpopulations. *Cell* 141: 69-80, 2010.
- 52 Knoblich JA: Asymmetric cell division: recent developments and their implications for tumour biology. *Nat Rev Mol Cell Biol* 11: 849-860, 2010.
- 53 Sherley JL: Asymmetric cell kinetics genes: the key to expansion of adult stem cells in culture. *Stem Cells* 20: 561-572, 2002.
- 54 Saywers CL: Will kinase inhibitors have a dark side? *New Engl J Med* 355: 313-315, 2006.
- 55 Bai J, Sata N and Nagai H: Gene expression analysis for predicting gemcitabine sensitivity in pancreatic cancer patients. *HPB* 9: 150-155, 2007.
- 56 Gréen H, Söderkvist P, Rosenberg P, Horvath G and Peterson C: Mdr-1 single nucleotide polymorphisms in ovarian cancer tissue: G2677T/A correlates with response to paclitaxel chemotherapy. *Clin Cancer Res* 12: 854-859, 2006.
- 57 Hegedüs C, Truta-Feles K, Antalffy G, Várady G, Németh K, Ozvegy-Laczka C, Kéri G, Orfi L, Szakács G, Settleman J, Váradi A and Sarkadi B: Interaction of the EGFR inhibitors gefitinib, vandetanib, pelitinib and neratinib with the ABCG2 multidrug transporter: Implications for the emergence and reversal of cancer drug resistance. *Biochem Pharmacol* 84: 260-267, 2012.
- 58 García Campelo MR, Alonso Curbera G, Aparicio Gallego G, Grande Pulido E and Antón Aparicio LM: Stem cell and lung cancer development: blaming the Wnt, Hh and Notch signalling pathway. *Clin Transl Oncol* 13: 77-83, 2011 Review.
- 59 Tsao PN, Chen F, Izvolsky KI, Walker J, Kukuruzinska MA, Lu J and Cardoso WV: Gamma-secretase activation of notch signaling regulates the balance of proximal and distal fates in progenitor cells of the developing lung. *J Biol Chem* 283: 29532-29544, 2008.
- 60 Sullivan JP, Spinola M, Dodge M, Raso MG, Behrens C, Gao B, Schuster K, Shao C, Larsen JE, Sullivan LA, Honorio S, Xie Y, Scaglioni PP, DiMaio JM, Gazdar AF, Shay JW, Wistuba II and Minna JD: Aldehyde dehydrogenase activity selects for lung adenocarcinoma stem cells dependent on notch signaling. *Cancer Res* 70: 9937-9948, 2010.
- 61 Westhoff B, Colaluca IN, D'Ario G, Donzelli M, Tosoni D, Volorio S, Pelosi G, Spaggiari L, Mazzarol G, Viale G, Pece S and Di Fiore PP: Alterations of the Notch pathway in lung cancer. *Proc Natl Acad Sci USA* 106: 22293-22298, 2009.
- 62 He B, Barg RN, You L, Xu Z, Reguart N, Mikami I, Batra S, Rosell R and Jablons DM: Wnt signaling in stem cells and non-small-cell lung cancer. *Clin Lung Cancer* 7: 54-60, 2005.
- 63 Teng Y, Wang X, Wang Y and Ma D: Wnt/beta-catenin signaling regulates cancer stem cells in lung cancer A549 cells. *Biochem Biophys Res Comm* 392: 373-379, 2010.
- 64 Zhang Y, Goss AM, Cohen ED, Kadzik R, Lepore JJ, Muthukumaraswamy K, Yang J, DeMayo FJ, Whitsett JA, Parmacek MS, Morrissey EE: A Gata6-Wnt pathway required for epithelial stem cell development and airway regeneration. *Nat Genet* 40: 862-870, 2008.
- 65 Shi Y, Fu X, Hua Y, Han Y, Lu Y and Wang J: The side population in human lung cancer cell line NCI-H460 is enriched in stem-like cancer cells. *PLoS One* 7: e33358, 2012.
- 66 Tian F, Schrödl K, Kiefl R, Huber RM and Bergner A: The hedgehog pathway inhibitor alters intracellular Ca<sup>2+</sup> homeostasis and inhibits cell growth in cisplatin-resistant lung cancer cells. *Anticancer Res* 32: 89-94, 2012.
- 67 Tiozzo C, De Langhe S, Yu M, Londhe VA, Carraro G, Li M, Li C, Xing Y, Anderson S, Borok Z, Bellusci S and Minoo P: Deletion of Pten expands lung epithelial progenitor pools and confers resistance to airway injury. *Am J Respir Crit Care Med* 180: 701-712, 2009.
- 68 Yanagi S, Kishimoto H, Kawahara K, Sasaki T, Sasaki M, Nishio M, Yajima N, Hamada K, Horie Y, Kubo H, Whitsett JA, Mak TW, Nakano T, Nakazato M and Suzuki A: Pten controls lung morphogenesis, bronchioalveolar stem cells, and onset of lung adenocarcinoma in mice. *J Clin Invest* 117: 2929-2940, 2007.
- 69 Matsumoto K, Arao T, Tanaka K, Kaneda H, Kudo K, Fujita Y, Tamura D, Aomatsu K, Tamura T, Yamada Y, Saijo N and Nishio K: mTOR signal and hypoxia-inducible factor-1 alpha regulate CD133 expression in cancer cells. *Cancer Res* 69: 7160-7164, 2009.
- 70 Agrawal M, Abraham J, Balis FM, Edgerly M, Stein WD, Bates S, Fojo T and Chen CC: Increased <sup>99m</sup>Tc-sestamibi accumulation in normal liver and drug-resistant tumors after the administration of the glycoprotein inhibitor, XR9576 *Clin Cancer Res* 9: 650-656, 2003.
- 71 Kelly RJ, Draper D, Chen CC, Robey RW, Figg WD, Piekarczyk RL, Chen X, Gardner ER, Balis FM, Venkatesan AM, Steinberg SM, Fojo T and Bates SE: A pharmacodynamic study of docetaxel in combination with the P-glycoprotein antagonist tariquidar (XR9576) in patients with lung, ovarian, and cervical cancer. *Clin Cancer Res* 17: 569-580, 2011.

- 72 Kelly RJ, Robey RW, Chen CC, Draper D, Luchenko V, Barnett D, Oldham RK, Caluag Z, Frye AR, Steinberg SM, Fojo T and Bates SE: A pharmacodynamic study of the p-glycoprotein antagonist CBT-1(R) in combination with paclitaxel in solid tumors. *Oncologist* 17: 512, 2012.
- 73 Wang F, Mi YJ, Chen XG, Wu XP, Liu Z, Chen SP, Liang YJ, Cheng C and Fu LW: Axitinib targeted cancer stem-like cells to enhance efficacy of chemotherapeutic drug via inhibiting the drug transport function of ABCG2. *Mol Med* 2012 Apr 25. doi: 10.2119/molmed.2011.00444. [Epub ahead of print]
- 74 Zhou WJ, Zhang X, Cheng C, Wang F, Wang XK, Liang YJ, Kin Wah To K, Zhou W, Huang HB and Fu LW: Crizotinib (PF-02341066) reverses multidrug resistance in cancer cells by inhibiting the function of p-glycoprotein. *Br J Pharmacol*. 2012 Jan 10. doi: 10.1111/j.1476-5381.2012.01849.x. [Epub ahead of print]
- 75 Bartucci M, Svensson S, Romania P, Dattilo R, Patrizi M, Signore M, Navarra S, Lotti F, Biffoni M, Pilozzi E, Duranti E, Martinelli S, Rinaldo C, Zeuner A, Maugeri-Saccia M, Eramo A and De Maria R: Therapeutic targeting of Chk1 in NSCLC stem cells during chemotherapy. *Cell Death Diff* 19: 768-778, 2012.
- 76 Pierceall WE, Olaussen KA, Rousseau V, Brambilla E, Sprott KM, Andre F, Pignon JP, Le Chevalier T, Pirker R, Jiang C, Filipits M, Chen Y, Kutok JL, Weaver DT, Ward BE and Soria JC: Cisplatin benefit is predicted by immunohistochemical analysis of DNA repair proteins in squamous cell carcinoma but not adenocarcinoma: theranostic modeling by NSCLC constituent histological subclasses. *Ann Oncol* 2012 Jan 23. doi: 10.1093/annonc/mdr624. [Epub ahead of print].
- 77 Lai YS, Chang CW, Pawlik KM, Zhou D, Renfrow MB and Townes TM: SRY (sex determining region Y)-box2 (Sox2)/poly ADP-ribose polymerase 1 (Parp1) complexes regulate pluripotency. *Proc Natl Acad Sci USA* 109: 3772-3777, 2012.
- 78 Shay JW and Wright WE: Telomeres and telomerase in normal and cancer stem cells. *FEBS Lett* 584: 3819-3825, 2010.
- 79 Serrano D, Bleau AM, Fernandez-Garcia I, Fernandez-Marcelo T, Iniesta P, Ortiz-de-Solorzano C and Calvo A: Inhibition of telomerase activity preferentially targets aldehyde dehydrogenase-positive cancer stem-like cells in lung cancer. *Mol Cancer* 10: 96, 2011.
- 80 Kim CB: MAPK-ing out the pathways in lung stem cell regulation. *Cell Stem Cell* 1: 11-13, 2007.
- 81 Ventura JJ, Tenbaum S, Perdiguero E, Huth M, Guerra C, Barbacid M, Pasparakis M and Nebreda AR: p38alpha MAP kinase is essential in lung stem and progenitor cell proliferation and differentiation. *Nature Genet* 39: 750-758, 2007.
- 82 Reeves ME, Baldwin ML, Aragon R, Baldwin S, Chen ST, Li X, Mohan S and Amaar YG: RASSF1C modulates the expression of a stem cell renewal gene, PIWIL1. *BMC Res Notes* 5: 239, 2012.
- 83 Gupta PB, Onder TT, Jiang G, Tao K, Kuperwasser C, Weinberg RA and Lander ES: Identification of selective inhibitors of cancer stem cells by high-throughput screening. *Cell* 138: 645-659, 2009.
- 84 Dizdarevic S and Peters AM: Imaging of multidrug resistance in cancer. *Cancer Imaging* 11: 1-8, 2011.
- 85 Wang C, Cheng L, Xu H and Liu Z: Towards whole-body imaging at the single cell level using ultra-sensitive stem cell labeling with oligo-arginine modified upconversion nanoparticles. *Biomaterials* 33: 4872-4881, 2012.
- 86 Krasznai ZT, Tóth A, Mikecz P, Fodor Z, Szabó G, Galuska L, Hernádi Z and Goda K: Pgp inhibition by UIC2 antibody can be followed *in vitro* by using tumor-diagnostic radiotracers, <sup>99m</sup>Tc-MIBI and 18FDG. *Eur J Pharm Sci* 41: 665-669, 2010.

Received July 10, 2012  
Accepted July 18, 2012