

## Full Paper

### *Flavobacterium ginsengiterrae* sp. nov., isolated from a ginseng field

Sang-Rae Kim, Yeon-Ju Kim, Ngoc-Lan Nguyen, Jin-Woo Min,  
Ji-Na Jeon, Dong-Uk Yang, and Deok-Chun Yang\*

Korean Ginseng Center and Ginseng Genetic Resource Bank, Kyung Hee University,  
1 Seocheon-dong, Giheung-gu, Yongin-si, Gyeonggi-do 449-701, South Korea

(Received April 14, 2011; Accepted August 17, 2011)

A novel strain of *Flavobacterium*, DCY55<sup>T</sup>, a Gram-negative, yellow-pigmented, rod-shaped, non-spore-forming and gliding-motile bacterium, was isolated from the soil of a ginseng field in South Korea. Phylogenetic analysis, based on the 16S rRNA sequence, demonstrated that strain DCY55<sup>T</sup> belongs to the genus *Flavobacterium* within the family *Flavobacteriaceae*. Strain DCY55<sup>T</sup> showed the highest similarity with *F. johnsoniae* UW101<sup>T</sup> (97.1%), *F. ginsenosidimutans* THG 01<sup>T</sup> (96.8%), *F. defluvii* EMB 117<sup>T</sup> (96.6%), *F. banpakuense* 15F3<sup>T</sup> (96.3%) and *F. anhuiense* D3<sup>T</sup> (95.8%). Chemotaxonomic results showed that strain DCY55<sup>T</sup> predominantly contains menaquinone MK-6, that its DNA G+C content is 36.1 mol%, and that its major cellular fatty acids are iso-C<sub>15:0</sub>, summed feature 3 (comprising iso-C<sub>15:0</sub> 2-OH and/or C<sub>16:1</sub>ω7c) and C<sub>16:0</sub>. The chemotaxonomic and genotypic characteristics support the taxonomic classification of strain DCY55<sup>T</sup> to the genus *Flavobacterium*. The results of physiological and biochemical tests confirmed that strain DCY55<sup>T</sup> is distinct from previously validated species. We conclude that strain DCY55<sup>T</sup> should be classified as a novel species of the genus *Flavobacterium*, for which the name *Flavobacterium ginsengiterrae* sp. nov. is proposed, with the type strain DCY55<sup>T</sup> (=KCTC 23319<sup>T</sup> = JCM 17337<sup>T</sup>).

**Key Words**—DCY55; *Flavobacteriaceae*; *Flavobacterium ginsengiterrae*

## Introduction

The genus *Flavobacterium* was established by Bergey et al. (1923) and was subsequently amended by Bernardet et al. (1996). It belongs to the family *Flavobacteriaceae*, phylum Bacteroidetes (Bernardet et al., 2002; Ludwig and Klenk, 2001). Members of the

genus *Flavobacterium* have been isolated from a variety of environments, such as the Antarctic, soil, freshwater, seawater and wastewater treatment plants (Kim et al., 2006; Liu et al., 2008; McCammon and Bowman, 2000; Park et al., 2007). Currently, the genus *Flavobacterium* consists of 70 recognized species, including the recently described species *F. dongtanense* (Xiao et al., 2011), *F. beibuense* (Fu et al., 2011), *F. ponti* (Yoon et al., 2011) and *F. sinopsychrotolerans* (Xu et al., 2011).

We isolated a novel bacterial strain from the soil of a ginseng field in South Korea. Biochemical analyses revealed this strain to be affiliated with the genus *Flavobacterium*. The strain was examined using phenotypic, physiological, and chemotaxonomic tests and by phylogenetic analysis. We report that strain DCY55<sup>T</sup> should be classified in the genus *Flavobacterium* as a

\* Address reprint requests to: Dr. Deok-Chun Yang, Korean Ginseng Center and Ginseng Genetic Resource Bank, Kyung Hee University, 1 Seocheon-dong, Giheung-gu, Yongin-si, Gyeonggi-do 449-701, South Korea.

Tel: +82-31-201-2100 Fax: +82-31-202-2687

E-mail: deokchunyang@yahoo.co.kr, yeonjukim@khu.ac.kr

The NCBI GenBank accession number for the 16S rRNA gene sequence of strain DCY55<sup>T</sup> (=KCTC 23319<sup>T</sup> = JCM 17337<sup>T</sup>) is HM776706.

Bootstrap values > 60% based on 1,000 replications are shown at branching points. Filled circles indicate that the corresponding nodes were also recovered in the tree constructed with the maximum-parsimony algorithm. *Myroides odoratus* ATCC 4651<sup>T</sup> (M58777) was used as an outgroup. Scale bar, 0.01 substitutions per nucleotide position.

maximum parsimony (Fitch, 1971) algorithms using MEGA 4 (Tamura et al., 2007) with bootstrap analyses based on 1,000 replications (Felsenstein, 1985). The evolutionary distances of the neighbor-joining method were calculated using the Kimura two-parameter model (Kimura, 1983). The resulting neighbor-joining tree showed that strain DCY55<sup>T</sup> was affiliated with the genus *Flavobacterium* (Fig. 1).

*Morphological and physiological characteristics.*

Cell morphology was observed with a Nikon light microscope (1,000× magnification). Transmission electron microscopy (TEM) was performed using the following procedure: bacteria were grown on TSA plates at 30°C for 24 h; resuspended cells were placed on carbon- and formvar-coated nickel grids for 30 s; and grids were floated on 1 drop of 0.1% (w/v) aqueous uranyl acetate, blotted dry, and then viewed with a Carl Zeiss LEO912AB electron microscope at 100 kV under standard operating conditions. Gliding motility was observed using the hanging drop technique (Bernardet et al., 2002).

Gram reactions were tested according to the non-

staining method as described by Buck (1982). Catalase activity was determined by bubble production in the presence of 3% (v/v) hydrogen peroxide solution, and oxidase activity was evaluated via the oxidation of 1% (w/v) tetramethyl-*p*-phenylenediamine (Merck, Whitehouse Station, NJ, USA). Growth on nutrient agar (NA), trypticase soy agar (TSA), and R2A agar was evaluated at 30°C. Growth at different temperatures was assessed at 4, 10, 20, 25, 30, 35, 37 and 42°C on TSA agar. Various pH values (pH 5.0–10.0 at intervals of 1.0 pH units) were assessed in trypticase soy broth (TSB), and salt tolerance was also tested in trypticase soy broth supplemented with 0, 1, 2, 3 and 5% (w/v) NaCl after 3 days' incubation. The production of flexirubin-type pigments was investigated by the method of Bernardet et al. (2002) as described previously. Hydrolysis of Tween 20 and Tween 80 was investigated according to Gerhardt et al. (1994). DNase activity and hydrolysis of casein, gelatin, tyrosine, starch and urea were also investigated as described by Cowan and Steel (1965). Hydrolysis tests of esculin and nitrate reduction were done as presented by Lanyi (1987).

Table 1. Physiological characteristics of strain DCY55<sup>T</sup> and related type strains of species of the genus *Flavobacterium*.

Characteristic	1	2	3	4	5	6
Tolerance of 2% NaCl	+	–	–	–	–	+
Oxidase activity	+	+	+	–	–	–
Gliding motility	+	+	–	+	+	+
Flexirubin-type pigments	+	+	NR	+	+	+
Nitrate reduction (NO <sub>3</sub> → NO <sub>2</sub> )	–	+	–	–	+	–
Degradation of:						
Gelatin	–	+	NR	+	+	–
Starch	+	+	+	–	+	+
DNA	+	+	–	+	NR	–
Assimilation of:						
L-Arabinose	+	+	+	(+)	+	+
N-acetyl-D-glucosamine	+	+	–	+	+	+
Enzymatic activities (API ZYM)						
α-Galactosidase	–	–	–	(+)	–	–
α-Glucosidase	+	+	NR	+	+	(+)
β-Galactosidase	(+)	+	+	(+)	+	+
β-Glucuronidase	–	+	–	+	(+)	–
β-Glucosidase	+	+	+	+	+	–
DNA G+C content (mol%)	36.1	35.2	32.1	33.5	31.1	31.4

Strains: 1, *Flavobacterium ginsengiterrae* DCY 55<sup>T</sup>; 2, *Flavobacterium johnsoniae* DSM 2064<sup>T</sup>; 3, *Flavobacterium ginsenosidimutans* THG 01<sup>T</sup> (data from Yang et al., 2011); 4, *Flavobacterium defluvii* DSM 17963<sup>T</sup>; 5, *Flavobacterium banpakuense* 15F3<sup>T</sup> (data from Kim et al., 2011); 6, *Flavobacterium anhuiense* KCTC 22128<sup>T</sup>. +, Positive result; (+), weakly positive; –, negative result; NR, not reported.

Hydrolysis of CM-cellulose was tested as described by Ramesh et al. (2008). Enzyme activities and additional biochemical tests were tested by determining API 20NE and API ZYM systems (bioMérieux, Marcy l'Etoile, France) according to the instructions of the manufacturer. The phenotypic characteristics of strain DCY55<sup>T</sup> are given in the species description and in Table 1.

**Chemotaxonomic characteristics.** Isoprenoidquinones were extracted and analyzed by HPLC according to Komagata and Suzuki (1987). For analysis of fatty acid composition, type strains were grown on TSA plates for 48 h at 28°C, and the cells were har-

vested. Fatty acids were extracted, methylated, and separated by gas chromatography, and the identification and quantification of methyl esters were conducted using a TSBA library (version 4.5) from Sherlock Microbial Identification Systems (MIDI, Newark, DE, USA) as described by Sasser (1990). The fatty acid profiles of strain DCY55<sup>T</sup> and related type strains are represented in Table 2.

**DNA base composition.** For G+C content analysis, the genomic DNA was extracted and purified using the Genomic DNA Isolation Kit (GeneAll Biotechnology, Seoul, Korea) according to the manufacturer's instructions. The DNA was degraded into nucleosides using P1 nuclease, and the G+C content mol% was measured using reverse-phase HPLC as described by Mesbah et al. (1989). Rather than 97.0%, a 16S rRNA gene sequence similarity threshold range of 98.7–99% is the point at which DNA–DNA reassociation experiments should be mandatory for testing the genomic uniqueness of a novel isolate (Stackerbrandt and Ebers, 2006).

## Results and Discussion

### Phylogenetic analysis

The sequence of strain DCY55<sup>T</sup> was compared with 16S rRNA sequences obtained from EzTaxonserver (<http://www.eztaxon.org>) using its BLAST program. 16S rRNA gene sequence analyses determined that strain DCY55<sup>T</sup> belongs to the genus *Flavobacterium* within the family *Flavobacteriaceae*. The highest similarity of sequence was with *F. johnsoniae* UW101<sup>T</sup> (97.1%), followed by *F. ginsenosidimutans* THG 01<sup>T</sup> (96.8%), *F. defluvii* EMB 117<sup>T</sup> (96.6%), *F. banpakuense* 15F3<sup>T</sup> (96.3%) and *F. anhuiense* D3<sup>T</sup> (95.8%), in diminishing order (Fig. 1).

### Morphological and physiological characteristics

Cells are Gram-negative, non-spore-forming, non-flagellated, aerobic rods (0.6–0.9 µm wide and 3.2–4.3 µm long) with gliding motility. Colonies grown on TSA agar are yellow, irregular, and slightly convex, with a diameter of 1–1.5 mm after 2 days of incubation on TSA agar. Growth occurs at 4–35°C (optimum, 30°C) and at pH 5–9 (optimum, pH 7.0). Growth occurs in the presence of 0–2% NaCl (optimum, 0–1%). Growth also occurs on NA and R2A. Flexirubin-type pigments are produced. Catalase and oxidase are positive. Nitrate is not reduced to nitrite. The bacteria

Table 2. Fatty acid profiles of strain DCY55<sup>T</sup> and related species of the genus *Flavobacterium*.

Fatty acid	1	2	3	4	5	6
<b>Saturated</b>						
C <sub>14:0</sub>	1.6	1.5	tr	1.9	1.2	1.2
C <sub>16:0</sub>	14.4	13.4	6.1	10.8	4.7	5.3
<b>Unsaturated</b>						
C <sub>15:1</sub> ω6c	tr	tr	tr	1.5	3.0	2.8
C <sub>16:1</sub> ω5c	2.1	1.5	tr	1.0	tr	tr
C <sub>17:1</sub> ω6c	tr	tr	tr	tr	1.4	1.2
C <sub>17:1</sub> ω8c	tr	tr	tr	tr	1.2	tr
<b>Branched</b>						
iso-C <sub>15:0</sub>	24.1	23.6	31.2	25.7	24.7	31.4
iso-C <sub>15:0</sub> 3-OH	5.1	4.8	8.9	5.4	8.3	7.3
iso-C <sub>15:1</sub> G	2.2	1.7	1.2	2.7	5.2	3.5
anteiso-C <sub>15:0</sub>	3.0	2.6	1.5	1.7	1.5	1.6
iso-C <sub>16:0</sub>	tr	tr	1.2	tr	tr	tr
iso-C <sub>16:0</sub> 3-OH	tr	tr	tr	tr	1.0	tr
iso-C <sub>17:0</sub>	tr	tr	1.5	tr	tr	tr
iso-C <sub>17:0</sub> 3-OH	5.2	5.5	10.3	4.3	8.2	6.2
iso-C <sub>17:1</sub> ω9c	2.0	2.6	6.5	2.0	tr	2.3
<b>Hydroxy</b>						
C <sub>15:0</sub> 3-OH	—	—	—	1.18	1.7	1.3
C <sub>16:0</sub> 3-OH	5.4	5.4	2.6	6.9	5.7	3.8
Summed feature 3 <sup>a</sup>	19.9	22.7	12.1	22.9	14.6	14.0

<sup>a</sup>Summed feature 3 comprises iso-C<sub>15:0</sub> 2-OH and/or C<sub>16:1</sub> ω7c that could not be separated by GLC with the MIDI system.

Strains: 1, *Flavobacterium ginsengiterrae* DCY55<sup>T</sup>; 2, *Flavobacterium johnsoniae* DSM 2064<sup>T</sup>; 3, *Flavobacterium ginsenosidimutans* THG 01<sup>T</sup> (nutrient agar at 28°C for 2 days, data from Yang et al., 2011); 4, *Flavobacterium defluvii* DSM 17963<sup>T</sup>; 5, *Flavobacterium banpakuense* 15F3<sup>T</sup> (tryptic soy agar at 30°C for 3 days, data from Kim et al., 2011); 6, *Flavobacterium anhuiense* KCTC 22128<sup>T</sup>. All data are from the present study except for column 3 and 5. Fatty acids amounting to <1% in all strains tested are not listed. tr, Trace amount (<1.0%); —, not detected.



degrade esculin, casein, CM-cellulose, DNA, starch and Tween 20, but not gelatin, tyrosine, urea or Tween 80. Acid production from carbohydrates is not present. The physiological characteristics of strain DCY55<sup>T</sup> and related type strains are summarized in Table 1.

#### Chemotaxonomic characteristics

The predominant quinone was MK-6; other members of the family *Flavobacteriaceae*, including the genus *Flavobacterium*, contain MK-6 as the predominant quinone (Bernardet et al., 2002). The cellular fatty acid profiles of strain DCY55<sup>T</sup> and other species are shown in Table 2. Strain DCY55<sup>T</sup> mainly contains fatty acids (>10% of the total composition) of iso-C<sub>15:0</sub> (24.1%), summed feature 3 (comprising iso-C<sub>15:0</sub> 2-OH and/or C<sub>16:1ω7c</sub>, 19.9%) and C<sub>16:0</sub> (14.4%). The major fatty acid profiles of strain DCY55<sup>T</sup> are highly similar to its closest type strain, *F. johnsoniae* DSM 2046<sup>T</sup>.

#### DNA base composition

The DNA G+C content of strain DCY55<sup>T</sup> was 36.1%. The DNA G+C content is in the range of 30–52 mol% (Liu et al., 2008), supporting that strain DCY55<sup>T</sup> is within the genus *Flavobacterium*.

Based on the results of polyphasic analysis, strain DCY55<sup>T</sup> should be classified as a novel species of the genus *Flavobacterium*, for which the name *Flavobacterium ginsengiterrae* sp. nov. has been proposed.

#### Description of *Flavobacterium ginsengiterrae* sp. nov.

*Flavobacterium ginsengiterrae* (gin.sen. gi. ter'rae. N. L. n. *ginsengum* ginseng; L. n. *terrae* soil; N.L. gen. n. *ginsengiterrae* of soil of a ginseng field, the source of the type strain). Cells are Gram-negative, non-spore-forming, non-flagellated, aerobic rods (0.6–0.9 µm wide and 3.2–4.3 µm long) with gliding motility. Colonies on TSA agar are yellow, circular, and slightly convex with a diameter of 1–1.5 mm after 2 days of incubation on TSA agar. Growth occurs at 4–35°C (optimum, 28–30°C), at pH 5–9 (optimum, pH 6.0–8.0) and in the presence of 0–2% NaCl (optimum, 0–1%). Growth also occurs on NA and R2A. Flexirubin-type pigments are produced. Catalase and oxidase are positive. Indole and H<sub>2</sub>S production are negative. The bacteria degrade esculin, casein, CM-cellulose, DNA, Starch and Tween 20, but not gelatin, tyrosine, urea or Tween 80. Acid production from carbohydrates is not present. In the API 20NE kit, esculin hydrolysis and β-galactosidase were positive, but reduction of nitrate to nitrite, indole

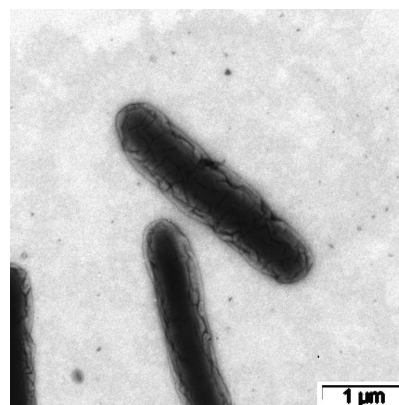


Fig. 2. Transmission electron micrograph of *Flavobacterium ginsengiterrae* DCY55<sup>T</sup>.

Bar, 1 µm.

production, glucose acidification, and arginine dihydrolase were negative. The species displayed assimilation of D-glucose, L-arabinose, D-mannose, *N*-acetyl-D-glucosamine and D-maltose but not D-mannitol, potassium gluconate, capric acid, adipic acid, malic acid, trisodium citrate or phenylacetic acid. In the API ZYM kit, alkaline phosphatase, esterase (C4), esterase lipase (C8), leucine arylamidase, valine arylamidase, cystine arylamidase, acid phosphatase, naphthol-AS-BI-phosphohydrolase, β-galactosidase (weak), α-glucosidase, β-glucosidase and *N*-acetyl-β-glucosaminidase activities were detected. But lipase (C14), trypsin, α-chymotrypsin, α-galactosidase, β-glucuronidase, α-mannosidase and α-fucosidase activities were not detected. The major cellular fatty acids are iso-C<sub>15:0</sub>, summed feature 3 (iso-C<sub>15:0</sub> 2-OH and/or C<sub>16:1ω7c</sub>) and C<sub>16:0</sub>. The major isoprenoid quinone is menaquinone-6 (MK-6), and the DNA G+C content is 36.1 mol%.

The type strain, DCY55<sup>T</sup> (=KCTC 23319<sup>T</sup> = JCM 17337<sup>T</sup>), was isolated from a soil sample from a ginseng field in South Korea.

#### Acknowledgments

This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (#20110015122).

#### References

- Bergey, D. H., Harrison, F. C., Breed, R. S., Hammer, B. W., and Huntoon, F. M. (1923) Genus II. *Flavobacterium* gen. nov. In *Bergey's Manual of Determinative Bacteriology*, Williams &

- Wilkins, Baltimore, pp. 97–117.
- Bernardet, J.-F., Nakagawa, Y., and Holmes, B. (2002) Proposed minimal standards for describing new taxa of the family *Flavobacteriaceae* and emended description of the family. *Int. J. Syst. Evol. Microbiol.*, **52**, 1049–1070.
- Bernardet, J.-F., Segers, P., Vancanneyt, M., Berthe, F., Kersters, K., and Vandamme, P. (1996) Cutting a Gordian knot: Emended classification and description of the genus *Flavobacterium*, emended description of the family *Flavobacteriaceae*, and proposal of *Flavobacterium hydatidis* nom. nov. (basonym, *Cytophaga aquatilis* Strohl and Tait 1978). *Int. J. Syst. Bacteriol.*, **46**, 128–148.
- Buck, J. D. (1982) Nonstaining (KOH) method for determination of Gram reactions of marine bacteria. *Appl. Environ. Microbiol.*, **44**, 992–993.
- Chun, J., Lee, J. H., Jung, Y., Kim, M., Kim, S., Kim, B. K., and Lim, Y. W. (2007) EzTaxon: A web-based tool for the identification of prokaryotes based on 16S ribosomal RNA gene sequence. *Int. J. Syst. Evol. Microbiol.*, **57**, 2259–2261.
- Cowan, S. T. and Steel, K. J. (1965) Manual for the Identification of Medical Bacteria, Cambridge University Press, London.
- Felsenstein, J. (1985) Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, **39**, 783–791.
- Fitch, W. M. (1971) Toward defining the course of evolution: Minimum change for a 222 specific tree topology. *Syst. Zool.*, **20**, 406–416.
- Fu, Y., Tang, X., Lai, Q., Zhang, C., Zhong, H., Li, W., Liu, Y., Chen, L., Sun, F., and Shao, Z. (2011) *Flavobacterium beibuense* sp. nov., isolated from marine sediment. *Int. J. Syst. Evol. Microbiol.*, **61**, 205–209.
- Gerhardt, P., Murray, R. G. E., Wood, W. A., and Krieg, N. R. (1994) Phenotypic characterization. In *Methods for General and Molecular Bacteriology*, ed. by Gerhardt, P., American Society for Microbiology, Washington, DC, pp. 607–654.
- Hall, T. A. (1999) BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp. Ser.*, **41**, 95–98.
- Kim, B.-Y., Weon, H.-Y., Cousin, S., Yoo, S.-H., Kwon, S.-W., Go, S.-J., and Stackebrandt, E. (2006) *Flavobacterium daejeonense* sp. nov. and *Flavobacterium suncheonense* sp. nov., isolated from greenhouse soils in Korea. *Int. J. Syst. Evol. Microbiol.*, **56**, 1645–1649.
- Kim, J.-J., Jin, H. M., Lee, H. J., Jeon, C. O., Kanaya, E., Koga, Y., Takano, K., and Kanaya, S. (2011) *Flavobacterium banpakuense* sp. nov., isolated from leaf-and-branch compost. *Int. J. Syst. Evol. Microbiol.*, **61**, 1595–1600.
- Kimura, M. (1983) The Neutral Theory of Molecular Evolution, Cambridge University Press, Cambridge.
- Komagata, K. and Suzuki, K. I. (1987) Lipid and cell wall analysis in bacterial systematics. *Methods Microbiol.*, **19**, 161–207.
- Lanyi, B. (1987) Classical and rapid identification methods for medically important bacteria. *Methods Microbiol.*, **19**, 1–67.
- Liu, H., Liu, R., Yang, S.-Y., Gao, W.-K., Zhang, C.-X., Zhang, K.-Y., and Lai, R. (2008) *Flavobacterium anhuiense* sp. nov., isolated from field soil. *Int. J. Syst. Evol. Microbiol.*, **58**, 756–760.
- Ludwig, W. and Klenk, H.-P. (2001) Overview: A phylogenetic backbone and taxonomic framework for procaryotic systematics. In *Bergey's Manual of Systematic Bacteriology*, 2nd ed., Vol. 1, ed. by Boone, D. R., Castenholz, R. W., and Garrity, G. M., Springer, New York, pp. 49–65.
- McCammon, S. A. and Bowman, J. P. (2000) Taxonomy of Antarctic *Flavobacterium* species: Description of *Flavobacterium gillisiae* sp. nov., *Flavobacterium tegetincola* sp. nov. and *Flavobacterium xanthum* sp. nov., nom. rev. and reclassification of [*Flavobacterium*] *salegens* as *Salegentibacter salegens* gen. nov., comb. nov. *Int. J. Syst. Evol. Microbiol.*, **50**, 1055–1063.
- Mesbah, M., Premachandran, U., and Whiteman, W. B. (1989) Precise measurement of the G+C content of deoxyribonucleic acid by high-performance liquid chromatography. *Int. J. Syst. Evol. Bacteriol.*, **39**, 159–167.
- Park, M., Ryu, S. H., Thi Vu, T.-H., Ro, H.-S., Yun, P.-Y., and Jeon, C. O. (2007) *Flavobacterium defluvii* sp. nov., isolated from activated sludge. *Int. J. Syst. Evol. Microbiol.*, **57**, 233–237.
- Ramesh, C. K., Richa, S., Hena, D., Som, D., and Arvind, G. (2008) A rapid and easy method for the detection of microbial cellulases on agar plates using Gram's iodine. *Curr. Microbiol.*, **57**, 503–507.
- Saitou, N. and Nei, M. (1987) The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.*, **4**, 406–425.
- Sasser, M. (1990) Identification of bacteria by gas chromatography of cellular fatty acids. MIDI Technical Note 101, MIDI, Inc., Newark, DE.
- Stackebrandt, E. and Ebers, J. (2006) Taxonomic parameters revisited: Tarnished gold standards. *Microbiol. Today*, **33**, 152–155.
- Tamura, K., Dudley, J., Nei, M., and Kumar, S. (2007) MEGA4: Molecular evolutionary genetics analysis (MEGA) software version 4.0. *Mol. Biol. Evol.*, **24**, 1596–1599.
- Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F., and Higgins, D. G. (1997) The CLUSTAL\_X Windows interface: Flexible strategies for multiple sequence alignment aided by quality analysis tools. *Nucleic Acids Res.*, **25**, 4876–4882.
- Xiao, Y. P., Hui, W., Lee, J. S., Lee, K. C., and Quan, Z. X. (2011) *Flavobacterium dongtanense* sp. nov., isolated from the rhizosphere of a wetland reed. *Int. J. Syst. Evol. Microbiol.*, **61**, 343–346.
- Xu, M., Xin, Y., Tian, J., Dong, K., Yu, Y., Zhang, J., Liu, H., and Zhou, Y. (2011) *Flavobacterium sino psychrotolerans* sp. nov., isolated from a glacier. *Int. J. Syst. Evol. Microbiol.*, **61**, 20–24.
- Yang, J.-E., Kim, S.-Y., Im, W.-T., and Yi, T.-H. (2001) *Flavobacterium ginsenosidimutans* sp. nov., a bacterium with ginsenoside converting activity isolated from soil of a ginseng field. *Int. J. Syst. Evol. Microbiol.*, **61**, 1408–1412.
- Yoon, J. H., Park, S., Kang, S. J., Oh, S. J., Myung, S. C., and Kim, W. (2011) *Flavobacterium ponti* sp. nov., isolated from seawater. *Int. J. Syst. Evol. Microbiol.*, **61**, 81–85.