



Benthic oxygen exchange in a live coralline algal bed and an adjacent sandy habitat: an eddy covariance study

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ABSTRACT: Coralline algal (maerl) beds are widespread, slow-growing, structurally complex perennial habitats that support high biodiversity, yet are significantly understudied compared to seagrass beds or kelp forests. We present the first eddy covariance (EC) study on a live maerl bed, assessing the community benthic gross primary productivity (GPP), respiration (R), and net ecosystem metabolism (NEM) derived from diel EC time series collected during 5 seasonal measurement campaigns in temperate Loch Sween, Scotland. Measurements were also carried out at an adjacent (~20 m distant) permeable sandy habitat. The O_2 exchange rate was highly dynamic, driven by light availability and the ambient tidally-driven flow velocity. Linear relationships between the EC O_2 fluxes and available light indicate that the benthic phototrophic communities were light-limited. Compensation irradiance (E_c) varied seasonally and was typically ~1.8-fold lower at the maerl bed compared to the sand. Substantial GPP was evident at both sites; however, the maerl bed and the sand habitat were net heterotrophic during each sampling campaign. Additional inputs of ~4 and ~7 mol $m^{-2} yr^{-1}$ of carbon at the maerl bed and sand site, respectively, were required to sustain the benthic O_2 demand. Thus, the 2 benthic habitats efficiently entrap organic carbon and are sinks of organic material in the coastal zone. Parallel deployment of 0.1 m^2 benthic chambers during nighttime revealed O_2 uptake rates that varied by up to ~8-fold between replicate chambers (from -0.4 to -3.0 mmol $O_2 m^{-2} h^{-1}$; $n = 4$). However, despite extensive O_2 flux variability on meter horizontal scales, mean rates of O_2 uptake as resolved in parallel by chambers and EC were typically within 20% of one another.

KEY WORDS: Benthic oxygen exchange · Benthic primary production · Coastal carbon cycling · Coralline algae · Permeable sediment · Eddy covariance · Benthic chambers

INTRODUCTION

Maerl beds are composed of dense aggregations of free-living red coralline algae (Rhodophyta, Corallinales) that accumulate on seabed surfaces with low sediment deposition rates and sufficient light avail-

ability (Martin et al. 2014). Maerl beds are slow-growing, structurally and functionally complex perennial habitats that support a rich diversity of micro- and macroalgae as well as benthic macrofauna (Barbera et al. 2003, Grall et al. 2006), and are important nursery areas for juvenile invertebrates

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and fish (Kamenos et al. 2004a,b). Maerl beds occur worldwide and are among the most widespread benthic phototrophic ecosystems in terms of areal coverage (Foster 2001). While their importance for regional biodiversity is increasingly recognized, maerl beds remain significantly understudied compared to other conspicuous benthic ecosystems such as seagrass beds, kelp forests, or shallow-water coral reefs. Coralline algal community productivity and its role in coastal carbon (C) cycling in particular have received little attention. This is surprising considering that the few existing studies on the topic indicate that maerl beds are among the most productive benthic ecosystems, with areal rates comparable to those of seagrass beds (Martin et al. 2005, 2007). Given their role in ecosystem service provision, a better understanding of maerl beds and their role in coastal C cycling is required.

The most widely used method for estimating benthic primary productivity and C turnover at the seabed is the benthic O₂ exchange rate (Glud 2008). Community rates of benthic gross primary productivity (GPP), respiration (*R*), and net ecosystem metabolism (NEM) have traditionally been estimated using benthic chamber incubations that quantify areal O₂ exchange rates. Typically, chamber incubations enclose a sediment area of 0.1 m² or less and incubations last only for a few hours. Community based diel assessments of activity therefore assume that the integrated spatial and temporal scales of the chamber incubations are representative of the whole benthic community. Application of benthic chambers in maerl beds is further complicated by the presence of the maerl structure and other large benthic macroalgae and fauna, and the enclosure process excludes the natural variations in hydrodynamics that the organisms otherwise experience. More recently, the aquatic O₂ eddy covariance (EC) method (Berg et al. 2003) was introduced, which allows benthic O₂ exchange rates to be estimated non-invasively over large (~10 to 100 m²) areas of the seabed. The EC method combines high-frequency measurements of vertical flow velocity and O₂ concentration taken at a single point (typically 10 to 30 cm above the seabed surface) to estimate the vertical turbulent exchange of O₂ within the benthic boundary layer. Under ideal conditions this exchange rate can be considered equal to the depth-integrated areal average sediment O₂ dynamics. Since EC measurements are carried out away from the seabed surface, this method is not confined to soft sediments. Importantly, EC measurements provide continuous time series of O₂ exchange rates covering one or more days, to

help elucidate the dynamics of benthic O₂ exchange under changing environmental conditions. This method has been applied to complex coastal benthic habitats such as high-latitude rocky benthic surfaces (Glud et al. 2010), seagrass beds (Hume et al. 2011), and coral reefs (Long et al. 2013, Rovelli et al. 2015), and therefore provides a new valuable tool for estimating the benthic O₂ exchange rate of maerl bed communities.

The main objective of this study was to investigate the biogeochemical functioning of a live maerl bed community and an adjacent sandy habitat, and their importance for coastal C cycling. This comparison was conducted by quantifying rates of benthic primary productivity and C degradation at both sites using the state-of-the-art EC O₂ flux method during 5 measurement campaigns across all 4 seasons. In addition, benthic chambers were deployed in parallel with the EC to resolve the benthic O₂ exchange rate using 2 independent methods.

MATERIALS AND METHODS

Study site

The study site was located in the tidal narrows of Caol Scotnish, a sheltered ~0.5 km² side-loch within the Loch Sween Marine Protected Area on the west coast of Scotland (56° 01.99' N, 05° 36.13' W; Fig. 1A). The shoreline rapidly descended to a depth of ~5 m at mean low water, and this depth remained consistent across the narrows. Closest to the shoreline, the seabed was characterised by bare sediments composed primarily of a mixture of sands, gravels, and relict shell fragments (Fig. 1B). The dominant macrofauna comprised the deposit-feeding black brittle star *Ophiocomina nigra*, occurring in densities of ~100 ind. m⁻². The permeability of the top ~10 cm of sediments measured on intact cores described by Klute & Dirksen (1986) was $1.7 \times 10^{-11} \pm 0.2 \text{ m}^{-2}$ (mean \pm SD, n = 4). Further out, in the middle of the narrows, the seabed was dominated by a live maerl bed of ~500 m length and ~40 m in width, characterised by a dense population of live maerl thalli, *Lithothamnion glaciale*, and high densities (~1000 ind. m⁻²) of the suspension-feeding common brittle star *Ophiothrix fragilis* and lower densities of the deposit-feeder *O. nigra* (Fig. 1C). The top 5 to 10 cm of the maerl bed consisted of a branched structure of live maerl thalli. This layer was underlain by a mixture of relict maerl skeletons and fine sediments that accumulated due to entrapment of finer particles

by the maerl structures. The combined permeability of the maerl thalli and underlying ~10 cm of sediments measured on intact cores as described by Klute & Dirksen (1986) was $2.2 \times 10^{-12} \pm 0.3 \text{ m}^{-2}$ (mean \pm SD, $n = 4$). The flow regime in Loch Sween is tidally-driven, with velocity magnitudes most often

within the range of 5 to 10 cm s^{-1} . However, short (~2 h) periods of intensified flow reaching 15 to 20 cm s^{-1} coincide with the incoming (flood) tide. The 2 measurement sites were located within ~20 m of one another. Surface waves were negligible (<10 cm) during the measurement campaigns.

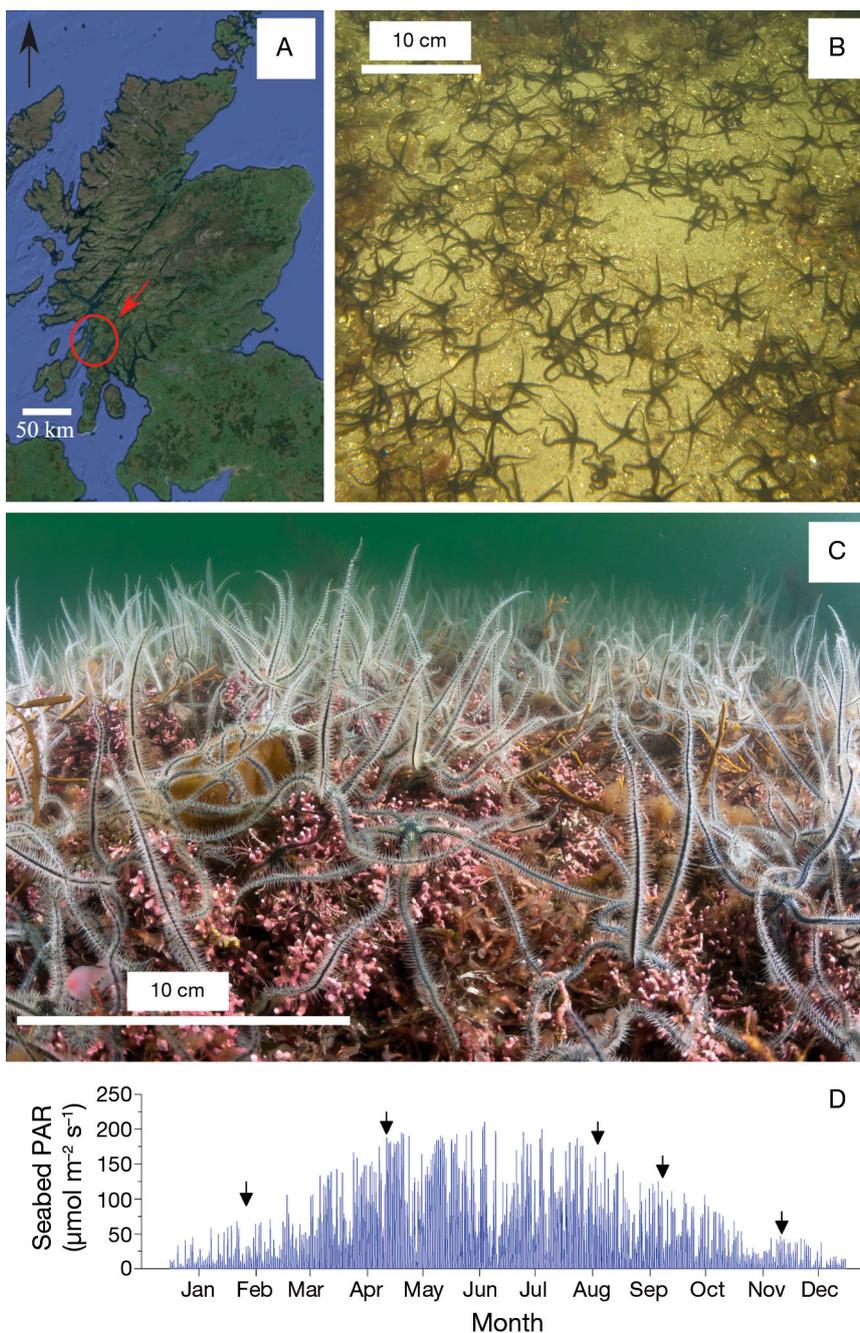


Fig. 1. (A) General location of the 2 study sites investigated in Loch Sween (red arrow) on the west coast of Scotland (map edited from Google Earth). (B) The sand/gravel site with high densities (~100 ind. m^{-2}) of the deposit-feeding brittle star *Ophiocomina nigra*, and (C) the live maerl bed with soft brown macroalgae and high densities of the suspension-feeding brittle star *Ophiothrix fragilis* (~1000 ind. m^{-2}). Maerl bed image courtesy of Rob Cook (Heriot-Watt University). (D) The 5 measurement campaigns were carried out in different seasons representing contrasting light regimes (black arrows) in Loch Sween

***In situ* instrumentation**

Eddy covariance measurements

For each of the 5 measurement campaigns at Loch Sween, we aimed to obtain EC time series of at least one complete 24 h period at each of the 2 sites. During the September 2010 and April 2012 campaigns, 2 EC systems were available. This allowed for simultaneous measurements of EC O₂ exchange rates at the 2 sites. During the other campaigns (November 2011, February 2012, August 2012) a single EC system was alternated between the 2 sites. In all cases, EC deployment was conducted by divers who used lift bags to carefully position the EC systems on the seabed. In total, 14 successful EC deployments were made at Loch Sween during the 5 measurement campaigns that altogether integrated >300 h of measuring time divided between the 2 sampling sites.

The configuration of the applied EC systems was similar to the original design described by Berg & Huettel (2008). The main components of the EC systems consisted of an acoustic Doppler velocimeter (ADV; Vector, Nortek) and a Clark-type O₂ microsensor with a 10 to 20 µm tip diameter (Revsbech 1989). Each microsensor was individually tested for its quality prior to deployment to ensure that it had a 90% response time of ≤0.5 s and a stirring sensitivity of <0.5% (Gundersen et al. 1998). The O₂ microsensor signal was relayed to the ADV via a submersible amplifier (McGinnis et al. 2011). The ADV recorded the longitudinal (*u*), traverse (*v*), and vertical (*w*) velocity components as well as the O₂ microsensor output in continuous mode at frequencies of 32 or 64 Hz, and in addition collected ancillary information such as instrument pitch and roll, flow direction, and signal strength. The equipment was mounted onto a stainless-steel tripod frame measuring 130 × 90 cm, designed to minimize hydrodynamic interference. The ADV sampling volume was located ~15 cm above the seabed surface. Visual inspection by divers ensured that the O₂ sensor tips and the ADV measurement volume stood well clear of bottom features such as stones, maerl branches, and benthic macrofauna. The O₂ microsensor tip was carefully positioned just at the edge of the ADV sampling volume (0.5 cm separation) to extract the O₂ data close to the ADV measurement point without compromising the velocity measurements. Prior to deployment, the EC O₂ microsensors were left to polarize for ~12 h to minimize sensor drift during deployments. Following polarization, the signal range and response time of each O₂ microsensor used was evaluated using water

samples of known O₂ concentration. A sodium dithionite solution was used for the zero O₂ saturation value, and collected bottom water samples were used for the *in situ* O₂ concentration. The O₂ concentrations were determined in the laboratory by Winkler titration.

Benthic chamber measurements

The square-shaped benthic chambers were constructed from semi-transparent Perspex and polyvinyl chloride, and enclosed an area of 961 cm² (0.1 m²). Each chamber had a removable lid that was fitted with a central rotating cross-shaped stirrer on the underside (Glud et al. 1996). The stirrer was powered by a small, self-contained battery-powered motor and was set to rotate at a constant frequency of 12 rpm. This rotation frequency was selected based on prior work by Glud et al. (1996) that characterised the pressure gradients within this type of chamber under controlled conditions. Several chambers were deployed by divers during the night, approximately 1 m apart from one another during periods that coincided with the EC deployments. A total of 4 chambers were deployed at each site in September, February and April; 3 were deployed in November, and 5 in August. The chambers were gently pushed ~15 cm into the sediment and the chamber lid was sealed to prevent exchange with the ambient bottom water. A sampling port on the side of the chamber wall allowed water samples from inside the chambers to be retrieved during incubation. A 100 ml water sample was retrieved by the divers at the start and at the end of each of the 3 to 6 h long incubations using a gas-tight syringe. Once extracted, water samples were transported to shore where they were fixed, stored in the dark and later analyzed in triplicate for O₂ concentration by Winkler titration. The decline in O₂ saturation between the start and the end of the incubation was <25%. The benthic chamber walls substantially attenuated the ambient light levels and were therefore not applied during daytime.

A substantial biomass of macroalgae had accumulated on the seabed surface in April. During this campaign, the divers were required to clear small areas of the seabed surface of the largest bits of algae prior to installing the benthic chambers in order to ensure a good seal between the chamber walls and the sediment. The results from this campaign are presented but are not discussed in detail since the flux measurements were likely compromised by this process.

In situ background environmental parameters

A conductivity temperature depth (CTD) sensor (XR-420, RBR) equipped with an O₂ optode (3830, Aanderaa) and a scalar photosynthetically active radiation (PAR) sensor (QSP-2200, Biospherical Instruments) was mounted onto a separate small stainless-steel frame that held the sensors 15 to 20 cm above the seabed. The system was deployed by divers at the interface between the maerl and sand sites, approximately equidistant between the 2 measurement sites. The CTD was programmed to record the environmental parameters covering the benthic chamber and EC deployments at 30 s intervals.

Eddy covariance fluxes

The EC O₂ fluxes were first extracted from the raw EC dataset and then evaluated for their ‘quality’ based on a set of defined criteria as detailed in Attard et al. (2014). Flux extraction was carried out using the open source software package Sulfide Oxygen Heat Flux Eddy Analysis (SOHFEA) version 2.0 (www.dfmcginnis.com/SOHFEA). Additional data treatments that are detailed below, but not available in SOHFEA, were carried out in MATLAB® (MathWorks). The datasets were processed for flux extraction in the following order: (1) weak signals in the raw 32 or 64 Hz ADV velocity data were identified by a low beam correlation and/or low signal strength. Individual data points with beam correlations below 50 % and signal-to-noise ratios below 12 dB were discarded. (2) The 32 or 64 Hz raw EC data was averaged down to 8 Hz. This reduced the noise level, and the smaller file size allowed for easier data handling. (3) A spectral analysis was carried out on the 8 Hz vertical velocity (i.e. w) and the O₂ microsensor data (C). The spectra showed the presence of an inertial subrange, identified as the region on the pressure spectral density plot for w and C where the slope of energy cascade from large scales to the smaller scales followed the predicted $-5/3$ fit, suggesting well-developed turbulence. The spectra were furthermore used to infer that the data reduction through adjacent averaging from 64 or 32 Hz to 8 Hz did not result in a loss of signal at high frequency, since most of the turbulent contributions typically occurred at a frequency of 1 Hz or lower (Fig. 2). (4) The O₂ microsensor output was calibrated to the CTD O₂ optode data using a linear regression. (5) Spike noise in the w and C data were removed using the

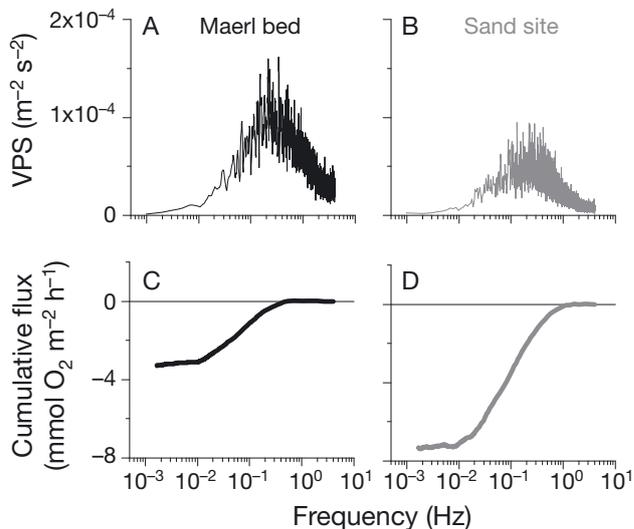


Fig. 2. Variance-preserving Power Spectra (VPS) of the vertical flow velocity for (A) the maerl bed and (B) the sand site. The data used in this example cover a 3 h dark period when 2 eddy covariance (EC) instruments were measuring in parallel at both sites. The mean (\pm SE) flow velocity magnitude (U) during this period was 6.9 ± 0.5 cm s⁻¹ at the maerl bed and 8.5 ± 0.5 cm s⁻¹ at the sand site. The average cumulative co-spectra (ogives) of the EC O₂ flux for the same period are presented in (C) and (D). The ogives indicate a dominance of flux-contributing eddies within the frequency range of 10^0 to 10^{-2} Hz, or 1 to 100s. The flux is seen to converge at lower frequencies

‘3D phase space’ method, and the excluded data points were interpolated using a cubic polynomial function (Mori et al. 2007). (6) Coordinate rotation was applied to the global 8Hz ADV velocity dataset using the ‘planar fit’ method to transform measured velocities into streamline coordinates (Lorke et al. 2013). (7) The O₂ fluxes (in mmol m⁻² h⁻¹) were extracted from the ADV w and C data as the covariance $\langle w'C' \rangle$, where w' and C' are deviations from a least-squares linear trend fitted to the measured vertical velocity and O₂ concentration respectively, and the angle bracket denotes time averaging. Following Reynolds decomposition theory, the vertical velocity vector may therefore be expressed as $w = \langle w \rangle + w'$ and the scalar quantity $C = \langle C \rangle + C'$ (Berg et al. 2003). The selected time averaging interval is a tradeoff between including as many of the flux-contributing turbulent eddies as possible while excluding low-frequency non-turbulent contributions such as advective flows (McGinnis et al. 2008, Lorke et al. 2013). To determine the optimal time interval, an analysis was carried out to investigate the effects of averaging time on the mean covariance and subsequent flux estimates for the friction velocity (u_*) and the O₂ fluxes. The u_* is directly related to the turbu-

lence regime within the benthic boundary layer (BBL). Estimates for u_* were computed from complex Reynolds stress measurements derived from the ADV velocity time series (McPhee 2008). The u , v , and w velocity components were decomposed into a mean and deviatoric velocity as $u = \langle u \rangle + u'$, $v = \langle v \rangle + v'$ and $w = \langle w \rangle + w'$. The u_* was then calculated as $u_* = (\langle u'w' \rangle^2 + \langle v'w' \rangle^2)^{1/4}$. The mean u_* and O_2 flux were computed as a function of the ensemble average. A time window of 10 min was consistently identified as the optimal interval for flux calculation at the 2 sites (Fig. 2). (8) A time-shift correction was applied to the data. Time shifting was performed for each ensemble interval by shifting the O_2 data in time relative to the velocity data to a maximum of 2 s to achieve the maximum numerical covariance (defined in terms of the maximum numerical flux) for $\langle w' C' \rangle$. This correction is necessary when the physical separation between the O_2 sensor and the ADV measurement volume, and/or the sensor response time, result in a slight misalignment in the data (McGinnis et al. 2008, Donis et al. 2015). (9) By assuming law-of-the-wall velocity profiles, the mean sediment surface roughness (z_0) was estimated as

$$z_0 = z \cdot \exp\left(-\kappa \cdot \frac{U}{u_*}\right)$$

where z is the measurement height above the benthic surface (0.15 m), κ is the von Karman constant (0.41), and U is the mean flow velocity magnitude (Berg et al. 2007).

The extracted 10 min EC fluxes were then carefully evaluated for their quality based on 3 criteria, namely by excluding periods with (1) large spikes or jumps in the O_2 microsensor data, (2) rapid changes in flow direction and/or flow velocity magnitude, and (3) suppressed exchange rates of O_2 due to insufficient turbulent mixing, which typically occurred when U dropped below $\sim 2 \text{ cm s}^{-1}$. Altogether, the screening process based on these 3 criteria typically resulted in the exclusion of <15% of the measured 10 min EC O_2 fluxes.

Dark benthic chamber O_2 fluxes

Areal rates of benthic O_2 uptake ($\text{mmol m}^{-2} \text{ h}^{-1}$) were derived from the rate of change of O_2 in the well-mixed incubated water phase over time as $(v/a) \times (\partial C/\partial t)$, where v is the volume of the water phase in the chamber (m^3), a is the area of enclosed sediment (m^2), C is the O_2 concentration (mmol m^{-3}), and t is the incubation time (h) (Glud 2008).

Rates of benthic productivity

The rates of benthic productivity were calculated from each time series of the EC O_2 exchange rates. Night-time periods were identified as the periods when PAR was $\leq 1 \text{ } \mu\text{mol quanta m}^{-2} \text{ s}^{-1}$. Daytime periods comprised the remaining intervals. The O_2 exchange during light (termed 'net daytime production', NDP) and R were then computed from the EC time series in $\text{mmol } O_2 \text{ m}^{-2} \text{ h}^{-1}$ as a bulk average of the O_2 fluxes during light and dark, respectively. Assuming a light-independent respiration rate, we also derived estimates for the benthic GPP ($\text{mmol } O_2 \text{ m}^{-2} \text{ h}^{-1}$) as $\text{GPP} = \text{NDP} + |R|$. Acknowledging that the rate of R during daytime typically is higher than that at night, we regard the GPP estimates presented herein as minimum values (Glud et al. 2009).

The autotrophic–heterotrophic balance of the benthic ecosystem (termed 'net ecosystem metabolism', NEM; $\text{mmol m}^{-2} \text{ d}^{-1}$) was derived directly from the EC time series as the 24 h integration of the O_2 flux. When more than 24 h of data was available (as was most often the case), the NEM was computed as a weighted average of the NDP and R fluxes accounting for the number of day versus night hours. The NEM indicated whether sediment O_2 production through photosynthesis balanced the various heterotrophic processes that directly or indirectly consume O_2 . Positive NEM values indicated a net O_2 release by the benthic ecosystem (autotrophy) while negative NEM values indicated a net O_2 uptake (heterotrophy) over a 24 h period.

Benthic community light response

The light dependency of the EC O_2 fluxes was evaluated using the photosynthesis versus irradiance (P – E) relationship. The screened 10 min EC O_2 fluxes covering at least one 24 h period were binned to 2 h intervals and plotted against the PAR data. This time interval was selected as a compromise between maintaining a high enough temporal resolution while minimizing the flow-induced effects and other non-steady state dynamics within the BBL that confounded the EC O_2 fluxes on shorter time scales (Holtappels et al. 2013). The relationship between EC O_2 fluxes and PAR was investigated using various fitting functions, and from regression analyses it was established that the P – E relationships were best explained by linear fits to the data. The compensation

irradiance (E_c , $\mu\text{mol quanta m}^{-2} \text{s}^{-1}$), that is, the PAR level at which GPP balances R , was subsequently derived from each linear fit as the x-intercept.

RESULTS

Environmental conditions

Bottom water temperature ranged from $16.6 \pm 0.4^\circ\text{C}$ in summer to $6.3 \pm 0.2^\circ\text{C}$ in winter. Bottom water salinity remained relatively constant during deployments but varied between 30.8 and 33.6 during the year. Bottom water O₂ saturation was lowest in November ($86 \pm 2\%$) and highest in August ($109 \pm 4\%$) (Table 1). Variations in the bottom water O₂ concentrations of up to $40 \mu\text{mol l}^{-1}$ (~15% saturation) during deployments coincided with the tidal advection of O₂-enriched waters from outer Loch Sween into Caol Scotnish. The daily integrated *in situ* PAR was lowest in February ($0.7 \text{ mol quanta m}^{-2} \text{d}^{-1}$) and highest in April ($5.5 \text{ mol quanta m}^{-2} \text{d}^{-1}$).

Eddy covariance measurements

The narrow (<100 m) width of the side loch in Loch Sween and the tidally-driven flow resulted in well-constrained hydrodynamics that allowed the divers to accurately align the EC instrument's longitudinal velocity component and the O₂ microsensors within the main flow direction. Therefore rotation of the traverse velocity component during data processing was minimal and typically amounted to less than $\pm 20^\circ$. Care was taken to deploy the EC instruments level with the seabed such that the ADV tilt as measured by the instrument's internal compass was less than $\pm 3^\circ$. Under unidirectional flow conditions, the time lag correction required to identify a maximum numerical flux was typically 0.2 to 0.8 s, which is in

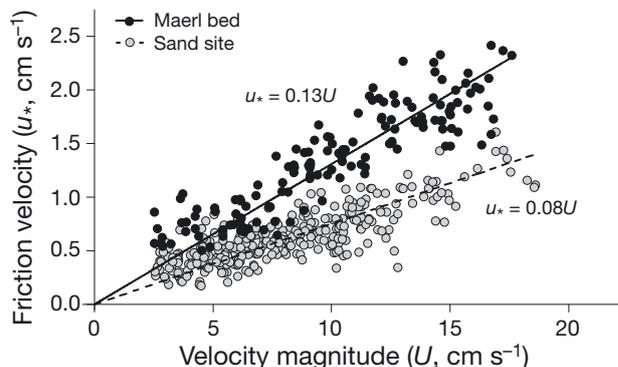


Fig. 3. Friction velocity (u_*) as a function of flow velocity magnitude (U) from parallel eddy covariance (EC) deployments over a 24 h period in April. Despite very similar flow magnitudes at both sites, u_* was consistently higher at the maerl bed, indicating more vigorous turbulent mixing due to higher benthic surface roughness

good agreement with the 90% response time of the O₂ microsensor. Linear relationships between the mean flow velocity magnitude (i.e. U) and u_* indicate that a well-developed turbulent BBL was present during ~90% of the sampling time at the 2 sites (Fig. 3). At low flow velocities (<5 cm s^{-1}) the relationship between the flow velocity and u_* deviated from linearity due to diminishing turbulent transport. When the flow velocity decreased below $\sim 2 \text{ cm s}^{-1}$ the O₂ fluxes were highly reduced due to insufficient turbulent mixing (Brand et al. 2008, Attard et al. 2014). Fluxes that fell within these periods amounted to <10% of the total time series, and were excluded from further analyses.

We evaluated the sensitivity of the EC O₂ fluxes to coordinate transformation by comparing the flux estimates derived following coordinate rotation by the 'double' and 'planar' rotation methods, which gave the same results. Furthermore, we followed the procedure described by Holtappels et al. (2015) to estimate the artificial flux caused by the stirring-sensi-

Table 1. Data from the CTD deployments for the 5 measurement campaigns. Values for depth, temperature, salinity, and O₂ are presented as a bulk average (\pm SD) of the entire deployment. Daytime hours are defined as periods when seabed photosynthetically active radiation (PAR) >1 $\mu\text{mol m}^{-2} \text{s}^{-1}$. sat. = saturation

Measurement campaign	Depth (m)	Temperature ($^\circ\text{C}$)	Salinity	O ₂ ($\mu\text{mol l}^{-1}$)	O ₂ (% sat.)	PAR ($\text{mol m}^{-2} \text{d}^{-1}$)	Hours of day/night
Sep	4.7 ± 0.3	14.6 ± 0.2	32.3 ± 0.3	245 ± 20	96 ± 8	0.95	12.8/11.2
Nov	5.0 ± 0.4	10.1 ± 0.3	30.8 ± 0.5	244 ± 7	86 ± 2	1.17	11.2/12.8
Feb	4.7 ± 0.4	6.3 ± 0.2	31.6 ± 0.2	310 ± 6	101 ± 2	0.70	8.5/15.5
Apr	4.6 ± 0.3	9.6 ± 0.4	33.6 ± 0.2	302 ± 11	107 ± 6	5.48	14.3/9.7
Aug	4.7 ± 0.3	16.6 ± 0.4	31.8 ± 0.7	270 ± 9	109 ± 4	3.29	14.5/9.5

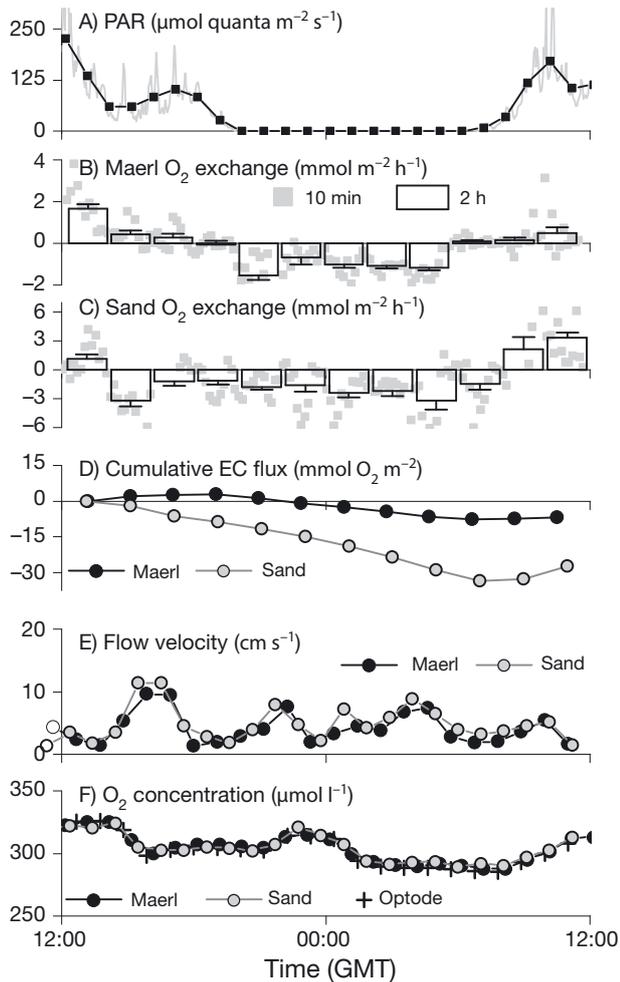


Fig. 4. A 24 h time series of eddy covariance (EC) O_2 fluxes measured in parallel at (B) the maerl bed and (C) the adjacent (~20 m apart) sand site in April. Positive fluxes indicate a net release of O_2 from the seabed and negative fluxes indicate net O_2 consumption. (A) The photosynthetically active radiation (PAR) availability was the main driver of O_2 production, with O_2 fluxes typically being positive during peak irradiance. (D) The 24 h integration of the O_2 fluxes represents the net ecosystem metabolism (NEM). (E) The mean flow velocity was the second main control on the EC fluxes. (F) Flow velocity and O_2 concentration show very good agreement between the 2 sites

tive O_2 microsensors. To determine the theoretical upper limit of the flux bias due to stirring sensitivity, we selected periods of nighttime EC data that fulfilled the conditions required to induce a maximum flux bias, that is (1) the O_2 microsensor being aligned with the main flow direction at an inclination of $\sim 60^\circ$, (2) high bottom-water O_2 concentrations, and (3) vigorous turbulent mixing due to high flow velocity and/or high z_0 (Holtappels et al. 2015). Using MODEL 2 and a sensor stirring sensitivity of 0.5%, we estimated the theoretical maximum flux bias to be

on the order of 10 to 15% of the measured EC O_2 fluxes, which is well within the error margin of our flux estimates.

The well-developed turbulent BBL in Loch Sween maintained a steady benthic flux signal and allowed for detailed inferences to be made about the dynamics of the benthic O_2 exchange rate in response to changing environmental conditions. Parallel deployment of 2 EC systems located ~ 20 m apart from one another documented similar O_2 flux dynamics, but substantial differences in the absolute exchange rates between the 2 sites that were exposed to very similar PAR, flow velocity magnitude, and bottom water O_2 concentrations (Fig. 4). Typically, the EC O_2 fluxes showed a distinct diel response to the availability of PAR. During peak irradiance the EC O_2 fluxes were positive, indicating net autotrophy. During the night, fluxes were negative. The diel signal in the data was overlain by extensive short-term variability, as seen in the 10 min fluxes in Fig. 4. This variability decreased substantially when averaged over 1 h or more. However, even on hourly time scales, the flow velocity magnitude was a major control on the EC fluxes. This was particularly evident in the nighttime data, where rates of EC O_2 uptake were enhanced by up to ~ 10 -fold at the sand site and by ~ 4 -fold at the maerl bed (Fig. 5). A seasonal trend in the slope of the regression between the nighttime O_2 uptake rate and the mean flow velocity was noticeable, especially at the sand site. The magnitude of the slope increased from months with low to high benthic activity (Fig. 5). This result could reflect

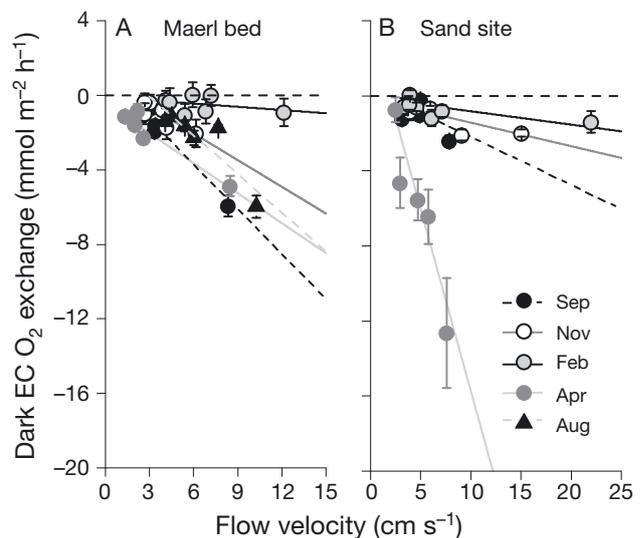


Fig. 5. Eddy covariance (EC) O_2 exchange rates in darkness (2 h bins) as a function of the mean flow velocity at (A) the maerl bed and (B) the sand site. Error bars are SE ($n = 12$)

the reduced state of the sediment, where reduced products from anaerobic decay processes accumulate in the surface sediments under low flow velocity, and are subsequently oxidized when the surface sediments are flushed with O₂ under high flow velocities.

The 2 h EC O₂ fluxes showed a good correlation with the *in situ* PAR during all seasons (Fig. 6, Table 2). There was no evidence of light saturation or photoinhibition in any of the 14 datasets (Fig. 6). Little light ($\sim 15 \mu\text{mol quanta m}^{-2} \text{s}^{-1}$) was required to drive a net autotrophic community response in winter. In the other campaigns the E_c was much higher, with peak values in September indicating a substantial heterotrophic component (Table 2). With the exception of February, the E_c at the sand site was consistently ~ 2 -fold higher than that at the maerl bed, suggesting that less light was required to drive a net community autotrophic response at the maerl bed.

Rates of EC O₂ exchange

We derived the first estimates of NDP, R , NEM, and GPP for a live maerl bed community using the non-invasive EC method as evidence for an active and productive benthic habitat. The maerl bed was a net source of O₂ to the surrounding waters during daytime in February, April, and August, up to $0.6 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$. There was a net community consumption of O₂ during daytime in September and November (Table 3). Mean EC O₂ exchange rates in the dark ranged from $-0.6 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$ in February to $-2.2 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$ in September. By contrast, the sand site was on average heterotrophic or at metabolic balance during the daytime (range from 0.04 to $-1.0 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$). The mean EC O₂ exchange during the night at the sand site ranged from $-0.5 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$ in February to $-3.7 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$ in April. When integrated over 24 h, both the maerl bed and the sand site were heterotrophic year-round, indicating that the benthic communities required additional inputs of organic matter originating from external sources. The maerl bed NEM ranged from -3.1 to $-33.1 \text{ mmol O}_2 \text{ m}^{-2} \text{ d}^{-1}$ and at the sand site it was from -3.1 to $-41.9 \text{ mmol O}_2 \text{ m}^{-2} \text{ d}^{-1}$.

The benthic phototrophic communities in Loch Sween were active during all measurement campaigns. The highest mean rates of GPP were measured in April (up to $3.3 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$) when the integrated daily PAR was at its highest ($5.5 \text{ mol quanta m}^{-2} \text{ d}^{-1}$). However, even in winter when the

duration of the daytime period was ~ 1.7 times shorter and daily PAR was ~ 8 -fold lower, the benthic phototrophs were still active. The highest mean rate of GPP for this period was measured at the maerl bed at $1.2 \text{ mmol O}_2 \text{ m}^{-2} \text{ h}^{-1}$, which is just ~ 1.5 -fold lower than the April GPP estimate for the same site

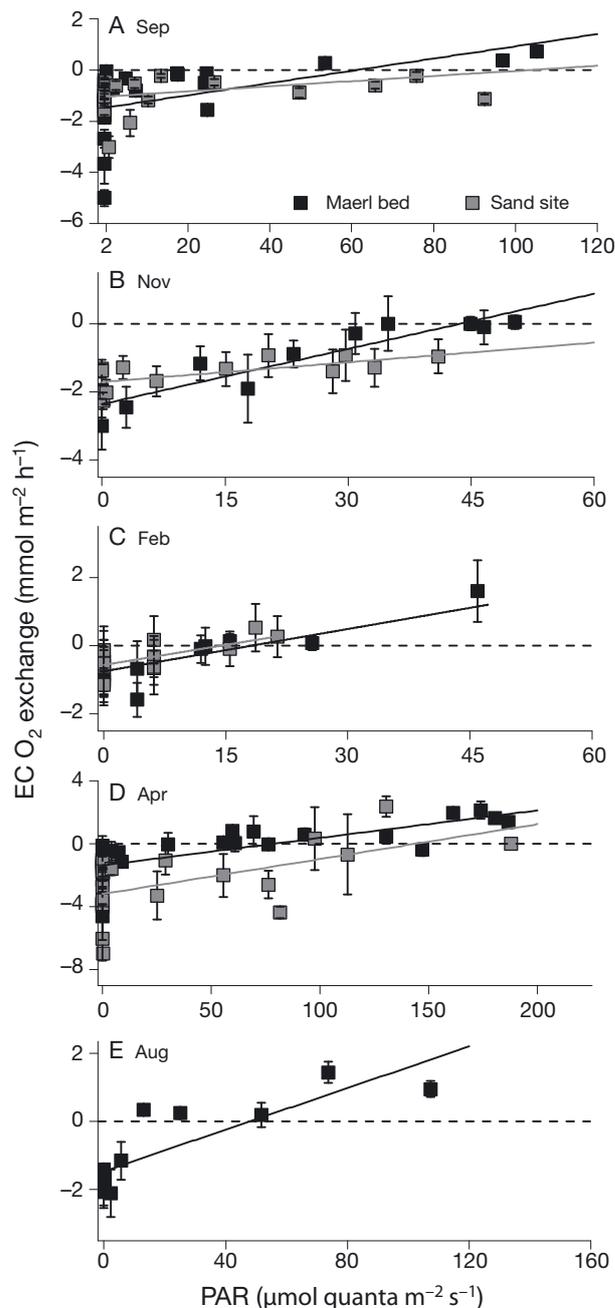


Fig. 6. Photosynthesis–irradiance (P – E) relationships for both sites over the year. Linear regressions were fitted to 2 h binned eddy covariance (EC) O₂ exchange rates ($n = 12$). With the exclusion of the February datasets, the compensation irradiance at the maerl site was consistently ~ 2 -fold lower than that at the sand site

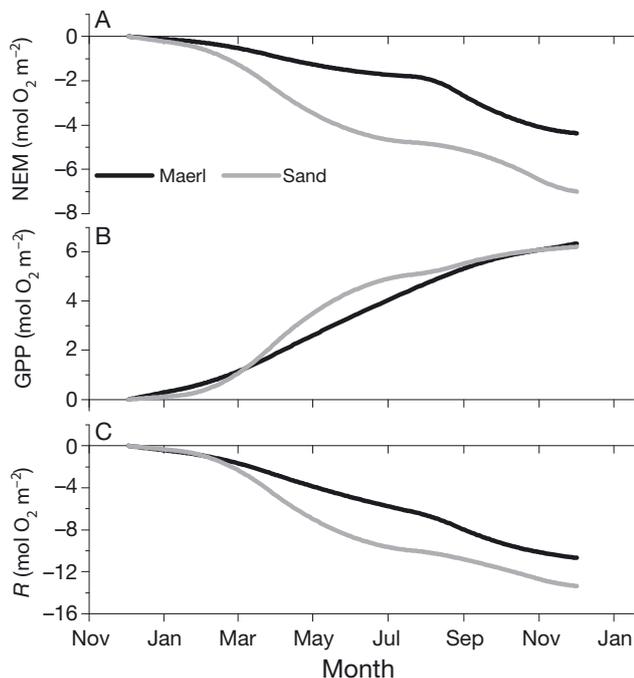


Fig. 7. Annual estimates of net (A) ecosystem metabolism (NEM), (B) gross primary productivity (GPP), and (C) respiration (R) for the 2 sites as derived from seasonal eddy covariance (EC) measurements. The difference between the curves reflects the difference in the response of the benthic community between sites

(Table 3). Interestingly, despite the low light levels, the highest mean rate of net O_2 release during day-time was measured at the maerl bed in February (NDP of $0.6 \text{ mmol } O_2 \text{ m}^{-2} \text{ h}^{-1}$).

The seasonally measured EC rates of NEM, GPP, and R (Table 3) were used to derive cumulative estimates of these parameters over the year (Fig. 7). This analysis assumes that the rates measured during the individual measurement campaigns are representative of each season. Annual estimates of GPP were

Table 2. The fitting statistics of the linear photosynthesis versus irradiance ($P-E$) relationships as fitted to 2 h averages of the eddy covariance (EC) O_2 exchange rates. The slope is in $\text{mmol } O_2 \text{ m}^{-2} \text{ h}^{-1} (\mu\text{mol quanta m}^{-2} \text{ s}^{-1})^{-1}$, the intercept is in $\text{mmol } O_2 \text{ m}^{-2} \text{ h}^{-1}$, and the compensation irradiance (E_c) is in $\mu\text{mol quanta m}^{-2} \text{ s}^{-1}$. No $P-E$ relationship is available for the sand site in August 2012 due to sensor damage incurred after ~6 h of deployment

Measurement campaign	Maerl site				Sand site			
	Slope	Intercept	E_c	R^2	Slope	Intercept	E_c	R^2
Sep	0.02	-1.5	61	0.26	0.01	-1.0	103	0.37
Nov	0.05	-2.4	43	0.83	0.02	-1.7	89	0.40
Feb	0.05	-0.8	16	0.84	0.04	-0.5	13	0.47
Apr	0.02	-1.4	78	0.52	0.02	-3.2	144	0.29
Aug	0.03	-1.5	48	0.68	-	-	-	-

~ $6 \text{ mol } O_2 \text{ m}^{-2} \text{ yr}^{-1}$ for both sites. However, the sand site was overall more net heterotrophic, with a NEM of $-7 \text{ mol } O_2 \text{ m}^{-2} \text{ yr}^{-1}$ and a R rate of $-13 \text{ mol } O_2 \text{ m}^{-2} \text{ yr}^{-1}$. At the maerl bed NEM was $-4 \text{ mol } O_2 \text{ m}^{-2} \text{ yr}^{-1}$ and R was $-11 \text{ mol } O_2 \text{ m}^{-2} \text{ yr}^{-1}$.

Hydrodynamics and EC footprint characteristics

The z_0 and u_* estimates computed from the ADV velocity time series were variable between sites and between seasons. However, the measurements consistently indicated more intensified turbulent mixing at the maerl bed compared to the sand site (Fig. 3). Mean (\pm SD) z_0 for the 5 campaigns at the sand site was $0.3 \pm 0.6 \text{ cm}$ ($n = 5$), whereas at the maerl site it was $1.2 \pm 1.2 \text{ cm}$ ($n = 5$). The characteristics of the EC flux footprint for the maerl bed calculated according to the parameterisation by Berg et al. (2007) suggest that the footprint was typically ~15 m long with a width of ~1 m and a region of maximum flux located ~0.5 m upstream from the instrument. For the sand site, the flux footprint had an average length of 30 m with a width of 1 m and the region of maximum flux was 1.3 m upstream from the instrument. Due to the tidal reversal and therefore the reversal of the upstream location of the EC footprint, the EC O_2 exchange estimates for GPP, R , and NEM actually integrate an area of about twice the size of the footprint dimensions estimated above. Analyses of the O_2 flux as a function of the flow direction suggested no substantial direction-dependent variations under the same light and flow conditions. The spatial extent of the maerl bed and the sand site was on the order of 100s of m upstream of the EC instrument in either direction of flow, and therefore we can be confident that the flux estimates presented here integrate the activities of each respective benthic ecosystem.

Shallow-water benthic habitats are frequently characterised by spatial heterogeneity in the distribution of benthic organisms such as fauna and algae that may cause an uneven distribution of the flux signal within the EC footprint area. For flux measurements to be representative of the total benthic community activities, the spatial scales integrated by the EC measurements must be larger than the spatial scales of variation at the seabed. The former can be estimated with knowledge of the measurement height at which the EC measure-

Table 3. Eddy covariance (EC) O₂ exchange rates (mean ± SE). The values for light fluxes (i.e. net daytime production, NDP), dark fluxes (i.e. respiration, R), gross primary productivity (GPP), and the dark chambers are in mmol m⁻² h⁻¹ whereas net ecosystem metabolism (NEM) values are in mmol m⁻² d⁻¹. MC = measurement campaign

MC	EC maerl bed					Maerl chambers (R)	EC sand site					Sand chambers (R)
	Dataset length (h)	NDP	R	GPP	NEM		Dataset length (h)	NDP	R	GPP	NEM	
Sep	44	-0.69 ± 0.4	-2.16 ± 0.4	1.49	-33.1	-2.36 ± 0.19	39	0.04 ± 0.3	-1.09 ± 0.5	1.13	-11.7	-1.03 ± 0.05
Nov	24	-0.30 ± 0.3	-0.95 ± 0.2	0.65	-15.5	-1.05 ± 0.13	26	-0.98 ± 0.2	-1.42 ± 0.5	0.44	-29.2	-1.07 ± 0.28
Feb	26	0.58 ± 0.3	-0.58 ± 0.1	1.16	-4.1	-1.05 ± 0.38	24	-0.03 ± 0.1	-0.48 ± 0.2	0.45	-7.8	-0.56 ± 0.18
Apr	51	0.15 ± 0.3	-1.63 ± 0.3	1.78	-13.7	-1.06 ± 0.19 ^a	43	-0.42 ± 1.5	-3.69 ± 1.3	3.27	-41.9	-1.07 ± 0.64 ^a
Aug	24	0.47 ± 0.2	-1.04 ± 0.2	1.51	-3.1	-1.11 ± 0.17	6	-0.08 ± 0.6	-	0.37 ^b	-3.1	-0.45 ± 0.13

^aCompromised measurements; excluded from yearly average
^bEstimated using the R-value from the chamber incubations

ments were performed (i.e. 0.15 m) and the average z_0 derived from the ADV measurements (Rheuban & Berg 2013). The heterogeneity of the seabed surface can be approximated from high-resolution images of the seabed. In short, a low measurement height and a high z_0 would result in a rapid transfer of the benthic flux signal from the seabed to the EC sensors, and would thus integrate small spatial scales. Following the procedure described by Rheuban & Berg (2013), we deduced that the spatial scales that our EC measurements integrated ('patch sizes' of ~30 cm at the maerl bed and ~50 cm at the sand site) were large relative to the spatial scales of variation within the EC footprint area. Therefore, errors induced due to spatial variability were small (<5%), and our EC setup was optimal for deriving O₂ exchange rates that are representative of the total benthic communities at both sites.

Benthic chamber measurements

The benthic O₂ exchange rates from the chamber deployments during the night revealed O₂ uptake rates at the maerl bed ranging from a mean (±SE) of -1.0 ± 0.1 mmol m⁻² h⁻¹ (n = 3) in November to -2.4 ± 0.2 mmol m⁻² h⁻¹ (n = 4) in September (Table 3). At the sand site, the range was from -0.5 ± 0.4 mmol O₂ m⁻² h⁻¹ (n = 4) in February to -1.1 ± 0.2 mmol O₂ m⁻² h⁻¹ (n = 4) in April. Variations in the resolved O₂ exchange rate between the replicate 0.1 m² chambers were less than ~2-fold at both sites in September and November. However in February, April, and August the replicate chambers resolved highly variable O₂ exchange rates that varied by up to ~6-fold at the maerl bed in February (from -0.3 to -1.9 mmol m⁻² h⁻¹) and by a factor of ~8 at the sand site in April (from -0.4 to -3.0 mmol m⁻² h⁻¹).

DISCUSSION

Metabolism of a live maerl bed

Detailed studies on the metabolism of isolated red coralline algae indicate low rates of photosynthesis and respiration in line with their slow growth rate of <1 mm yr⁻¹ (Roberts et al. 2002, Blake & Maggs 2003, Martin et al. 2013). Growth rates of *Lithothamnion glaciale* in Loch Sween have been estimated at around 0.2 mm yr⁻¹ (Burdett et al. 2011). However, live maerl beds increase the substratum heterogeneity and provide structurally complex habitats that are colonized by a large diversity of both autotrophic and heterotrophic organisms (Barbera et al. 2003, Teichert 2014). Therefore, it is likely that the respiration and primary production of the maerl themselves only represent a small fraction of the total benthic community C turnover rates. The highly porous maerl framework is ventilated by the turbulent water flow, and particles, nutrients, and plankton can easily flow through the complex cavities and sustain community metabolism. The ventilation process, the complex structure of the maerl bed, and the spatiotemporal variability in O₂ production and consumption introduce various assumptions when quantifying the GPP, R, and NEM using benthic chambers that could potentially compromise the estimates. In contrast to this, the EC method is non-invasive and the measurements integrate a large footprint area. This study is the first of its kind to apply this method to document community O₂ dynamics and quantify benthic primary productivity and C turnover in such environments.

The non-invasive EC measurements revealed substantial benthic primary production in the maerl bed year-round. To our knowledge, there exists just one other seasonal study on benthic O₂ exchange in a

maerl bed community, conducted in the temperate Bay of Brest, France, using *in situ* benthic chambers (Martin et al. 2007). The reported GPP values for that site range from ~4 to 134 mmol O₂ m⁻² d⁻¹ between winter and summer, with an annual rate of 24.5 mol O₂ m⁻² yr⁻¹. This rate is ~4-fold higher than the yearly GPP estimate for the maerl bed community in Loch Sween of ~6 mol O₂ m⁻² yr⁻¹. However, the PAR available for benthic photosynthesis at our study site was up to ~8-fold lower than that reported by Martin et al. (2007) for the Bay of Brest. This difference could explain much of the variations in the GPP between the 2 temperate sites. In contrast with phytoplankton productivity, benthic primary productivity is typically limited by PAR rather than nutrient availability (Barranguet et al. 1998, Gattuso et al. 2006). This inference is supported by our *in situ* EC measurements, which document a linear increase of the EC O₂ fluxes in response to increasing light levels at the seabed. Therefore, a maximum rate of benthic O₂ production was not observed, even when seabed PAR reached ~300 μmol quanta m⁻² s⁻¹ in summer (Fig. 6, Table 2).

Despite substantial benthic primary production, the benthic O₂ consumption exceeded O₂ production in the maerl bed over a diel period, and the maerl bed community was net heterotrophic during each sampling campaign. Even though the biomass of live maerl thalli can be substantial, its contribution to the food web is thought to be low due to the very low organic content of its tissues (<6%; N. A. Kamenos pers. obs.) and the limited number of grazers that are able to feed directly on live maerl fragments (Graham 1988, Grall et al. 2006, Martin et al. 2007). Most of the C required by the benthic communities was therefore likely synthesized by epiphytic macroalgae and microphytobenthos, but the communities required an additional input of C from the surrounding environment amounting to ~4 mol C m⁻² yr⁻¹ (assuming a respiration quotient of 1.0). These inferences are consistent with the seasonal study in the Bay of Brest, where despite a substantial annual benthic GPP, the maerl community was net heterotrophic and required an additional C input of 14 mol m⁻² yr⁻¹ (167 g C m⁻² yr⁻¹) (Martin et al. 2007).

The annual C deficit in Loch Sween was likely a result of sedimentation and entrapment of phytoplankton, and to a lesser extent, terrestrially derived material from the catchment area. The potential importance of external inputs of organic matter to the maerl bed is substantiated by the high densities of the suspension-feeding brittle stars *Ophiothrix fragilis* associated with the live maerl in Loch Sween

(Fig. 1C). The heterogeneous substrate of live maerl thalli in combination with the hydrodynamics at the seabed and the high concentration of particles in the water column make this habitat ideal for suspension-feeding brittle stars (Davoult & Gounin 1995). This may explain the high densities of *O. fragilis* at the maerl bed compared to the adjacent sandy habitat that was instead dominated by the deposit feeding brittle star *Ophiocomina nigra* (Fig. 1).

Carbon turnover in maerl versus sandy sediments

To date, studies comparing maerl beds with adjacent bare sediments have focused on the importance of the maerl structure for local benthic biodiversity (Barbera et al. 2003, Kamenos et al. 2003, Teichert 2014). No previous studies have compared the biogeochemical functioning of the maerl bed to adjacent bare sediments. The 2 habitats we investigated were located at the same depth and within ~20 m of one another, and therefore experienced the same environmental conditions of PAR, flow velocity, O₂ concentrations, and temperature. This provided an ideal opportunity for detailed comparisons of community based C turnover and their controls at these 2 contrasting sites.

The EC measurements revealed very similar annual GPP for both sites: ~6 mol O₂ m⁻² yr⁻¹ (Fig. 7). These rates are comparable to the annual depth-integrated phytoplankton productivity in the <35 m deep photic zone measured at 3 nearby locations using ¹⁴C-labelled bicarbonate incubations (range from 10 to 17 mol C m⁻² yr⁻¹; Rees et al. 1995). Despite a net light under-saturation of the benthic communities, the seabed is an important component for primary production in the shallow water ecosystems of the Loch Sween Marine Protected Area.

The benthic GPP rates presented above must be regarded as minimum estimates, since computing the GPP from O₂ exchange measurements invokes the assumption that O₂ consumption is equal during day and night (Glud et al. 2009). During daytime, the rate of O₂ consumption is typically 1.4 to 1.8-fold higher as a result of (1) deeper O₂ penetration and hence a larger volume of sediment that can support O₂ consumption, and (2) elevated turnover of leaching labile photosynthates (Epping & Jørgensen 1996, Fenchel & Glud 2000). Evidence of this process within EC O₂ exchange time series is often inferred from high rates of O₂ consumption at dusk, when photosynthetic activity ceases and the leftover labile pool of photosynthates is efficiently turned over

within the first hours of the night (Glud et al. 2010, Long et al. 2013, Rheuban et al. 2014). This feature was also evident in the EC datasets from both the maerl bed and the sand site (Fig. 4B), where the rates of O₂ uptake at dusk frequently were ~2-fold higher than the corresponding values later in the night. No changes in flow velocity, flow direction, bottom water temperature, or O₂ concentration could otherwise explain this difference. However, we cannot exclude that temporal shifts in fauna activity also could have contributed to the elevated rates of O₂ consumption at sunset, a process that has been observed in other shallow water habitats (Wenzhöfer & Glud 2004).

The availability of PAR at the seabed was the main driver of benthic O₂ production at both sites (Figs. 4 & 6). The community E_c , that is, the PAR level at which GPP balances R , exhibited a distinct seasonal trend (Table 2). At both sites, E_c was lowest in winter (~15 $\mu\text{mol quanta m}^{-2} \text{s}^{-1}$). During the remaining seasons the E_c at the sand site was consistently ~2 times higher than that at the maerl bed. The higher E_c at the sand site was largely a result of flow-induced stimulation of O₂ uptake during the day. At the maerl bed, daytime EC measurements were largely unaffected by flow velocity. However at the more permeable sand site, high mean flow velocities (>5 cm s^{-1}) induced a negative O₂ exchange rate that deviated substantially from O₂ exchange rates measured at similar irradiances under lower flow velocities (<5 cm s^{-1}) (Fig. 8). Therefore, the net sediment O₂ release from the photic zone within the sediments under high irradiances was offset by a higher flow-induced uptake rate of O₂ as a result of deep and rapid pore-water advection (Berg et al. 2013).

A key component of the biogeochemical functioning of benthic environments is the entrapment and degradation of newly settled organic material by sediments and fauna (Middelburg et al. 2005). Both of these processes are included within the EC flux measurements. Interestingly, even though the maerl bed had high densities of suspension-feeding brittle stars and a highly porous upper framework, the C degradation at the sand site still exceeded that of the maerl bed (C deficit of ~4 $\text{mol m}^{-2} \text{yr}^{-1}$ at the maerl bed and ~7 $\text{mol m}^{-2} \text{yr}^{-1}$ at the sand site). This result could reflect a higher entrapment efficiency of organic material at the more permeable sand site. Highly permeable sediments have been termed 'biocatalytic filters' for their ability to effectively entrap and decompose organic matter (Huettel et al. 2014, McGinnis et al. 2014). However, for hydraulic connectivity in the surface sediments to persist, incorporation of fine sediments into the sand matrix must be

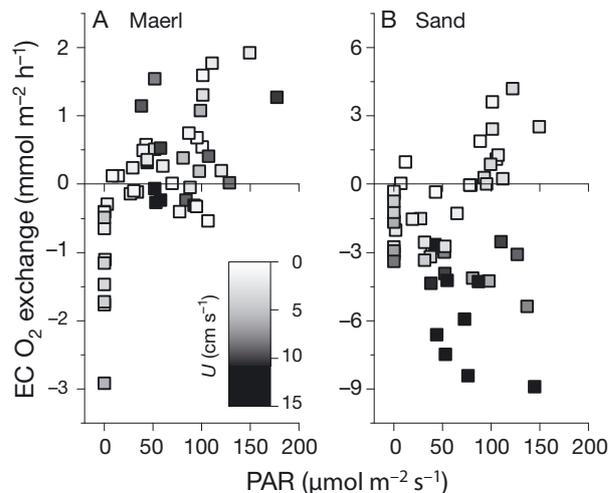


Fig. 8. Photosynthesis–irradiance ($P-E$) relationship for both sites showing the effect of flow velocity on discrete 10 min eddy covariance (EC) O₂ exchange rates over an 8 h period in April. The flow had little to no effect on the exchange rates at the maerl bed during the day, but substantially stimulated an O₂ uptake that offset the net photosynthetic O₂ release at the more permeable sandy site

kept low due to low deposition rates or by repeated cycles of current- and wave-induced sediment resuspension and winnowing. Erosion of sands occurs above a critical u_* threshold of ~0.7 cm s^{-1} (Wiberg & Smith 1987), which corresponds to a flow velocity of ~5 cm s^{-1} at the maerl bed and ~9 cm s^{-1} at the sand site. This is well below the maximal u_* values observed at both sites during a typical tidal cycle in Loch Sween (Fig. 3).

Comparison to global benthic primary productivity estimates

Globally, complex light-exposed benthic ecosystems such as maerl beds and permeable sediments cover large areas of the inner shelf region and contribute substantially to local and regional primary production (Gattuso et al. 2006). Our range of GPP estimates for the 2 sites in Loch Sween (from 0.7 to 1.8 $\text{mmol O}_2 \text{m}^{-2} \text{h}^{-1}$ for the maerl bed and from 0.4 to 3.3 $\text{mmol O}_2 \text{m}^{-2} \text{h}^{-1}$ for the sand site) exceed the global average estimates for benthic microalgal productivity for the 0 to 5 m depth range in temperate regions (~0.8 $\text{mmol C m}^{-2} \text{h}^{-1}$; Cahoon 1999). However, the Loch Sween GPP estimates are at the lower end of those for the maerl bed in the Bay of Brest, France (range from 0.2 to 5.6 $\text{mmol O}_2 \text{m}^{-2} \text{h}^{-1}$; Martin et al. 2007), as well as EC-derived GPP estimates for shallow (3 to 8 m deep) benthic habitats in a

southwest Greenland fjord (range from 0.1 to 5.7 mmol O₂ m⁻² h⁻¹; Glud et al. 2010, Attard et al. 2014). EC-derived GPP estimates in temperate seagrass beds are substantially higher (range from 2 to 20 mmol O₂ m⁻² h⁻¹; Rheuban et al. 2014), as are those for tropical coral reefs (up to 40 mmol O₂ m⁻² h⁻¹; Long et al. 2013). Therefore, while the benthic communities of Loch Sween contribute substantially to local and regional primary production, the GPP rates are somewhat modest when compared to other shallow benthic habitats.

Benthic chamber versus EC measurements

There are a growing number of studies within the literature comparing the benthic community O₂ exchange rates as resolved using chamber incubations versus the EC method. The 2 approaches have been reported to provide similar rates for cohesive sediments that lack large conspicuous macrofaunal species, giving confidence in the 2 measurement techniques (Berg et al. 2003, 2009). Many of the studies that employed both methods in more complex benthic systems such as permeable sands and in those dominated by large macrofauna also suggest good agreement between the average rates (Table 4). The largest discrepancies between the 2 methods (up to a factor of ~4) are reported for highly

permeable sediments under dynamic flow conditions (Berg et al. 2013). For coarse-grained sediments, the interaction between the boundary-layer flows and the sediment topography results in small lateral pressure gradients that drive an advective-dominated O₂ flux into the permeable seabed (Huettel & Gust 1992, Huettel et al. 2003). Benthic chambers attempt to artificially recreate these conditions but may not fully capture the *in situ* dynamics. Therefore, we could expect a wide range of reported values for the EC-versus chamber-resolved O₂ fluxes in permeable environments (EC:chamber range from 0.5 to 4.1; Table 4).

In this study, we obtained the benthic O₂ uptake rate from parallel measurements using multiple benthic chambers and EC in the dark at 2 contrasting shallow-water sites and in different seasons. To date, these measurements constitute the most extensive benthic chamber versus EC comparisons. The benthic ecosystem of Loch Sween is comprised principally of permeable sediments and a patchy distribution of large epifauna and macroalgae (Fig. 1B,C), which suggests substantial horizontal spatial heterogeneity in the O₂-consuming processes. This is reflected in the large variations between replicate 0.1 m² chambers by up to a factor of ~6 at the maerl bed and ~8 at the sand site. Despite large variability between individual chambers, the mean rates of O₂ uptake in the dark estimated from the chamber

Table 4. Studies in various benthic settings that employed both chamber and eddy covariance (EC) measurements to estimate the benthic O₂ uptake rate for non-photoc sediments, classified as 'muds' and 'sands'. Flow velocity is measured by the EC instrument 10 to 15 cm above the seabed surface. Units for benthic chamber surface area are in m². Whenever seasonal measurements are available, the EC:chamber (EC:Ch) is presented as a range of values obtained, and the overall mean is presented in brackets. The April measurements in Loch Sween were excluded from the overall average due to the benthic chamber measurements being compromised during this campaign (see 'Materials and methods: Benthic chamber measurements')

Sediment type	EC:Ch	Depth (m)	Flow velocity (cm s ⁻¹)	Chambers (no. of replicates, surface area)	Location	Study
Muds	1.4	12	2	4, 0.1	Aarhus Bay, Denmark	Berg et al. (2003)
	1.1	8	2–4	6, 0.1	Limfjord, Denmark	Berg et al. (2003)
	0.9	1450	1–3	2, 0.1	Sagami Bay, Japan	Berg et al. (2009)
	0.9	50	1–13	4, 0.1	Loch Etive, Scotland	Glud et al. unpubl. data
Sands	4.1	3	31	5, 0.1	Wakulla River, USA	Berg et al. (2013)
	2.2	1	0–5	5, 0.1	Florida, USA	Berg et al. (2015) ^b
	1.5	74	1–10	1, 0.1	Tommeliten, North Sea	McGinnis et al. (2014)
	1.1	107	3–10	1, 0.2	Half Moon Bay, USA	Johnson et al. (2011)
	0.9–1.3 (1.2)	5	2–20	10, 0.1	Loch Sween, Scotland	This study ^a
	0.7–0.8 (0.8)	95	3–17	3, 0.2	Monterey Bay, USA	Johnson et al. (2011) ^a
	0.5	80	0–13	2, 0.04	Oregon Shelf, USA	Reimers et al. (2012)
Maerl bed	0.6–0.9 (0.8)	5	2–20	16, 0.1	Loch Sween, Scotland	This study ^a

^aSeasonal measurements
^bRevised EC rates from Berg & Huettel (2008)

deployments in most cases compared well to those resolved in parallel by EC (Table 3). At the maerl bed, the ratio of the mean (\pm SD) benthic O₂ uptake rate as resolved by EC to the chambers ranged from 0.6 ± 0.1 ($n = 4$) to 0.9 ± 0.3 ($n = 5$) with an overall average of 0.8 ± 0.2 ($n = 16$). At the sand site, the range was from 0.9 ± 0.1 ($n = 4$) to 1.3 ± 0.5 ($n = 2$) with an overall average of 1.2 ± 0.2 ($n = 10$). These values are well within the range of the ratios typically reported for coastal aphotic benthic systems (between 0.8 and 1.5; Table 4).

The agreement between the EC- and chamber-resolved measurements presented here lends credibility to the application of both methods in complex environments. However, the large variation between replicate chambers indicates that replication is critical to integrate larger spatial scales and resolve an O₂ uptake rate that is representative of the total benthic community. The number of replicate chambers required varies between sites and depends on the amount of heterogeneity present. For example, in a simulation study performed by Glud & Blackburn (2002) for a deep-sea cohesive sediment dominated by relict burrow structures (30 to 120 burrows m⁻²), 4 replicate 0.1 m² benthic chambers were able to determine the benthic O₂ uptake rate to within ~95% accuracy. This was reduced to ~80% when the same analysis was applied at a more heterogeneous coastal cohesive site dominated by sparsely distributed large epifauna (Glud & Blackburn 2002). Logistical constraints frequently limit deployment of multiple benthic chambers, and here the EC method holds a clear advantage in its ability to integrate larger spatial scales and to better resolve temporal dynamics in the O₂ flux. In this respect, the EC method represents a powerful new approach for detailed studies on benthic O₂ dynamics in complex coastal environments such as maerl beds and other permeable settings.

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