

Heterosis in Japonica/Indica F₁ Rice Hybrids with Special Reference to Nodal Root Function*

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Abstract : Although root system of japonica/indica F₁ hybrids of rice (*Oryza sativa* L.) must play an important role for expression of heterosis, few studies have been conducted. This study was carried out to evaluate how the function of root system in japonica/indica F₁ rice hybrids is related to heterosis for shoot growth. Two F₁ hybrids grown in culture solution for 45 days after sowing and F₁ hybrid cultured in soil-filled pots till full ripening were compared with parents. Shoot dry weight of F₁ hybrids was greater than that of parents when their stem and nodal root numbers were similar to those of parents. Heterobeltiosis for shoot dry weight reached the maximum 180 when the F₁ hybrid surpassed parents in the stem and nodal root numbers. F₁ hybrids were the same as one or both parents in S/R ratio, but dry weight of a single nodal root of F₁ hybrids exceeded that of parents in most cases. Moreover, nitrogen uptake rate of a single nodal root (N_{no}) as well as leaf area and shoot dry weight of a single nodal root in F₁ hybrids was greater than those in parents during vegetative growth. Nitrogen uptake rate per root system dry weight (N_{dw}), however, was much the same. These results strongly suggest that in japonica/indica F₁ hybrids, more photosynthate is partitioned to each nodal root, so that this enhances its function and consequently, the entire root system.

Key words : Dry matter partitioning, Heterosis, Japonica/indica F₁ hybrid, Nitrogen uptake, Rice, Root system, Single nodal root.

ジャポニカ/インディカ F₁ 雑種水稻の雑種強勢とその節根の機能 : 長谷川浩・金 忠男・河野恭広* (北陸農業試験場・*名古屋大学農学部)

要 旨 : 一般に作物の雑種強勢において根系が重要な役割を果たしていると考えられる。にもかかわらず、ジャポニカ/インディカ F₁ 雑種水稻の根系に関する研究は極めて少ない。そこで、本研究は根系の機能が地上部の雑種強勢にどう関連しているかを明らかにするために行われた。播種から 45 日間水耕栽培した F₁ 雑種 (MS 1003/密陽 23 号, MS 1003/新青矮 1 号) および成熟期までポットで土壌栽培した F₁ 雑種 (MS (N)/密陽 23 号) を両親と比較した。F₁ 雑種では茎数や節根数が両親と同程度であっても地上部乾物重は両親を上回ったが、茎数や節根数が多かった場合には地上部乾物重の Heterobeltiosis は最高 180 に達した。栄養生長期には S/R 比は両親と大差なかったが、節根 1 本当たりの乾物重は生育初期から完熟期までのほとんどの場合両親より大きかった。さらに、栄養生長期には節根 1 本当たりの葉面積、地上部乾物重および窒素吸収速度が高かった。しかしながら、根系乾物重当たりの窒素吸収速度は両親と同様の値であった。以上の結果は、ジャポニカ/インディカ F₁ 雑種水稻では節根 1 本当たりの乾物分配量が多く、このことが節根 1 本当たりの機能を高め、結果として根系全体の機能も高くなることを強く示唆している。

キーワード : 乾物分配, 根系, 雑種強勢, ジャポニカ/インディカ F₁ 雑種, 節根 1 本当たりの機能, 水稻, 窒素吸収速度。

Since Jones¹¹⁾ discovered heterosis in F₁ rice hybrid, a number of physiological studies on heterosis for their shoot growth^{1,2,5,14,15,23~27)} have been conducted.

Root system of F₁ rice hybrid must play an important role for expression of heterosis. Although root development^{15,18,33)}, nutrient uptake^{8~10,16,28)}, root RNA content⁵⁾, root ATP content and enzyme activity³³⁾, root exudate^{4,5,15,16)}, root density³³⁾, root/whole

plant dry weight ratio¹³⁾ and pulling force^{7,20)} of F₁ hybrids have already been investigated, the role of the root system is still unclear. Japonica/indica F₁ rice hybrids being developed at Hokuriku National Agricultural Experiment Station are expected to exhibit larger heterosis and higher yield. However, studies about the japonica/indica F₁ rice hybrids^{18,28)} are few in number.

Gramineous root systems consist of seminal and nodal root axes and lateral roots produced on those axes. It is thus important to elucidate relationship between dry matter partitioning

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and function of each seminal and nodal root. Therefore, the objective of this study is to evaluate how the function of root system in japonica/indica F₁ rice hybrids is related to heterosis for shoot growth with special reference to the function of a single nodal root including seminal root.

Materials and Methods

1. Experiment 1

Two F₁ rice hybrids (MS1003/Milyang 23, MS1003/XinGingAi 1) and parents [Akihikari; the maintainer line (japonica), Milyang 23 and XinGingAi 1; the restorer lines (indica)] were grown in Kasugai rice culture solution¹²⁾ in a greenhouse under natural light and temperature for 45 days. The cytoplasmic male sterile (CMS) line, MS1003, has the nuclear DNA of Akihikari and the cytoplasm of Chinsurah Boro II.

Seeds were selected with salt solution (specific gravity of 1.06) and sterilized with 1000-fold dilution of thiram and benomyl for 24 hours. Pregerminated seeds were sown on a stainless net floating in a container filled with 77 l of 8-fold dilution of the culture solution {5ppm N as (NH₄)₂SO₄; 1.1ppm P as Na₂HPO₄; 3.1ppm K as KCl; 0.36ppm Ca as CaCl₂; 0.45ppm Mg as MgCl₂; 0.43ppm Fe-Citrate; 0.041ppm Mn as MnCl₂} on May 27 of 1989. The culture solution was prepared with tap water. The culture solution was renewed on the 7th day (Day 7) and the 10th day (Day 10) after sowing and then the pH of the culture solution was adjusted to 5.1 with 1N of hydrochloric acid and sodium hydroxide. On Day 14, two hundred seedlings were transplanted at a distance of 5cm × 4.5cm into a bed (132 l; length 110cm, width 60cm, depth 20cm plus tank 168 l) filled with 40-fold dilution of the solution (1ppm N; 0.22ppm P; 0.62ppm K; 0.72ppm Ca; 0.90ppm Mg; 0.86ppm Fe; 0.082ppm Mn). Since the actual nitrogen concentration of the paddy soil solution in heavy clay of Hokuriku district (2ppm or less)³¹⁾ is very much lower than that of the usual rice culture solution (10–30ppm). The culture solution was circulated at the rate of 10l per minute and renewed every week. The pH of the solution was adjusted daily to 5.1. Mutual shading among rice plants was negligible.

According to the plan size, five to eighty

plants for each F₁ hybrid and parents were collected seven times beginning on Day 14 until Day 45. The leaf area and number of nodal roots including seminal root were determined. The plants were dried in an oven at 80° C for two days and the dry weights of the shoots and root systems were taken. The dried shoots were ground and their nitrogen content was determined by the semi-micro Kjeldahl digestion and Indophenol method²²⁾ with Autoanalyzer (TRAACS-800, Technicon Industrial Systems, USA).

Since the linear relations between shoot nitrogen content and nodal root number or dry weight of root system were observed, nitrogen uptake rate of a single nodal root (N_{no}) and per dry weight of root system (N_{dw}) were estimated by a formula analogous to mean net assimilation rate used in growth analysis¹⁹⁾:

$$N_{no} = \frac{N_2 - N_1}{RN_2 - RN_1} \times \frac{\text{Log}_e(RN_2) - \text{Log}_e(RN_1)}{T_2 - T_1}$$

$$N_{dw} = \frac{N_2 - N_1}{RW_2 - RW_1} \times \frac{\text{Log}_e(RW_2) - \text{Log}_e(RW_1)}{T_2 - T_1}$$

where T_1 and T_2 is Day T_1 and Day T_2 , respectively, and N_1 , RN_1 , RW_1 and N_2 , RN_2 , RW_2 is shoot nitrogen content, number of nodal roots and dry weight of root system on Day T_1 and Day T_2 , respectively.

2. Experiment 2

F₁ rice hybrid (MS (N)/Milyang 23) and parents [Nekken 2; the maintainer line (japonica) and Milyang 23] were cultured in pots under outdoor conditions till full ripening in 1990. The CMS line, MS (N), has the nuclear DNA of Nekken 2, in which wide-compatibility genes were incorporated³⁾, and the cytoplasm of Chinsurah Boro II.

Cylindrical plastic pots (diameter, 25cm, depth; 30cm) were filled with 14.4kg of air-dried heavy clay soil, which was sieved through 5mm mesh. Sixteen g of crushed compound fertilizer (N; 15%, P; 6.5%, K; 12%) was thoroughly mixed with the soil as basal dressing before filling the pots. Five pregerminated seeds of the varieties prepared in the same way as Experiment 1 were sown in a pot under upland conditions in a greenhouse on April 21. After seedling emergence the pots were waterlogged, and the seedlings were thinned into one plant per pot. The pots were

Table 1. Comparison of leaf area, dry weight, shoot nitrogen content and nodal root number of two F₁ hybrids and parents (Experiment 1).

	Plant age in leaf number	Leaf area (cm ² /plant)	Dry weight (mg/plant)		Shoot /root ratio	Shoot nitrogen content (mg/plant)	Number of nodal root
			Shoot	Root system			
Day 14							
A	3.0	0.79 b	9.61 a	4.90 a	1.96 b	0.279 a	8.6 a
M	3.0	0.65 b	8.04 b	3.81 b	2.12 a	0.231 b	6.9 b
FM	2.9	1.08 a (137)	10.15 a (106)	5.27 a (108)	1.93 b (91)	0.284 a (102)	6.1 b (71)
A	3.0	0.79 c	9.61 b	4.90 c	1.96 a	0.279 b	8.6 a
X	3.2	1.21 b	8.86 c	5.46 b	1.62 b	0.258 b	8.7 a
FX	3.1	1.55 a (129)	10.68 a (111)	6.63 a (121)	1.61 b (82)	0.315 a (113)	7.5 a (86)
Day 24							
A	5.0	6.15 b	37.5 b	13.9 b	2.70 a	1.61 b	13.8 a
M	5.1	6.73 b	35.8 b	13.2 b	2.71 a	1.61 b	13.9 a
FM	5.0	9.26 a (138)	44.9 a (120)	16.3 a (117)	2.76 a (102)	2.05 a (127)	13.9 a (100)
A	5.0	6.15 b	37.5 b	13.9 b	2.70 a	1.61 b	13.8 b
X	5.9	8.60 a	42.2 b	17.7 a	2.38 b	2.04 a	17.1 a
FX	5.5	9.52 a (111)	48.8 a (116)	18.4 a (104)	2.65 a (98)	2.25 a (110)	16.2 a (95)
Day 45							
A	9.8	141 b	1180 b	501 b	2.36 a	38.6 b	95.4 a
M	9.2	153 b	945 c	517 b	1.83 b	31.8 c	61.4 c
FM	9.0	207 a (135)	1458 a (124)	740 a (143)	1.99 b (84)	46.5 a (120)	77.0 b (81)
A	9.8	141 b	1180 b	501 b	2.36 a	38.6 ab	95.4 a
X	10.2	144 b	934 c	576 b	1.62 c	34.9 b	72.2 c
FX	9.3	205 a (143)	1422 a (121)	782 a (136)	1.82 b (77)	45.2 a (117)	83.4 b (87)

Day 0 : Sowing day, A : Akihikari, M : Milyang 23, FM : MS1003/Milyang 23, X : XinGingAi 1, FX : MS1003/XinGingAi 1. Values with the same letter in each combination are not significant at 5% level by Tukey's Studentized Test. Values in parentheses indicate heterobeltiosis [= F₁ / (better parent) × 100].

transferred outside the greenhouse on Day 20, and then grown as single plants which were free from mutual shading. On Day 92, 2.4g of ammonium sulfate was applied to each plant as top dressing.

In each sampling the number of stems and nodal roots and shoot dry weights were determined on three to five plants for F₁ hybrid and parents, respectively. Dry weights of root systems were measured by means of the ashing method²⁹⁾ on one to four plants.

3. Data analysis

Data analysis was performed by the ANOVA (Analysis of Variance), the GLM (General Linear Model) and the TTEST (t-test) procedures of PC-SAS (Statistical Analysis Systems for Personal Computers, Verison 6.03, SAS Inc., USA)²¹⁾.

Results

1. Experiment 1

Table 1 shows leaf area, dry weight, shoot

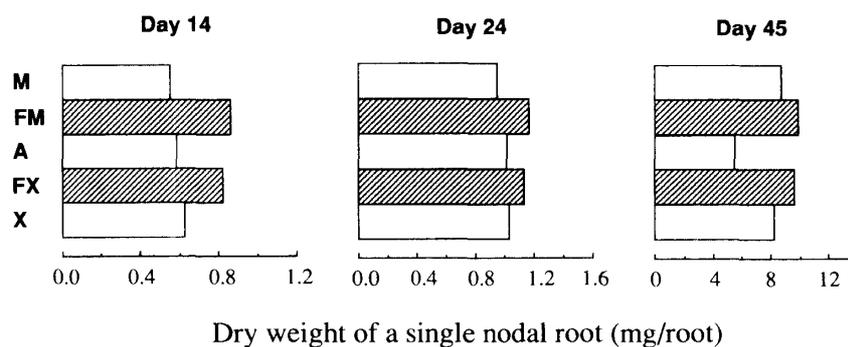


Fig. 1. Comparison of dry weight of a single nodal root of two F₁ hybrids and parents (Experiment 1).
M : Milyang 23, A : Akihikari, X : XinGingAi 1, FM : MS1003/Milyang 23, FX : MS1003/XinGingAi 1.

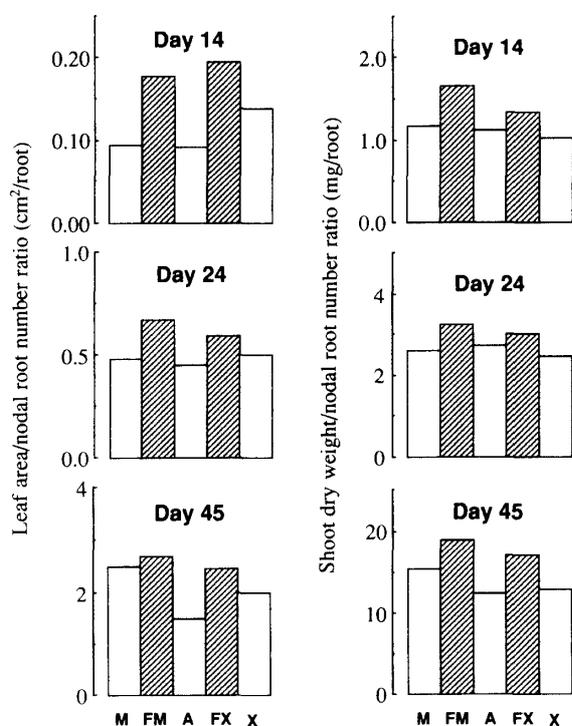


Fig. 2. Comparison of leaf area and shoot dry weight/number of nodal roots ratios of two F₁ hybrids and parents (Experiment 1). Abbreviations M, A, X, FM and FX, see Fig. 1.

nitrogen content, and number of nodal roots on Day 14, Day 24 and Day 45. Shoot dry weight of both F₁ hybrids significantly surpassed that of parents. Leaf area, root system dry weight and shoot nitrogen content of both F₁ hybrids were all greater than those of parents as well, although not always significant.

On the other hand, the number of nodal roots of both F₁ hybrids was about same as that of either parent, or in between that of

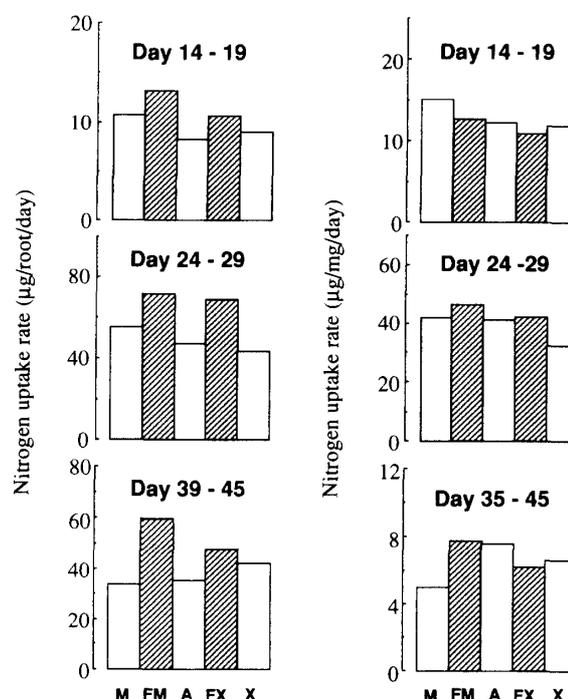


Fig. 3. Comparison of nitrogen uptake rate of a single nodal root (N_{no} , left) and per root system dry weight (N_{dw} , right) in two F₁ hybrids and parents (Experiment 1). Abbreviations M, A, X, FM and FX, see Fig. 1.

parents throughout the experimental period. The stem number of Akihikari, Milyang 23, MS1003/Milyang 23, XinGingAi 1 and MS1003/XinGingAi 1 on Day 45 was 7.2, 7.8, 8.0, 8.4 and 8.8, respectively, but significant differences between mean values was not observed. Shoot/root system dry weight ratio (S/R ratio) showed a similar trend to the number of nodal roots. However, dry weight of a single nodal root of both F₁ hybrids, calculated from dividing the root system dry

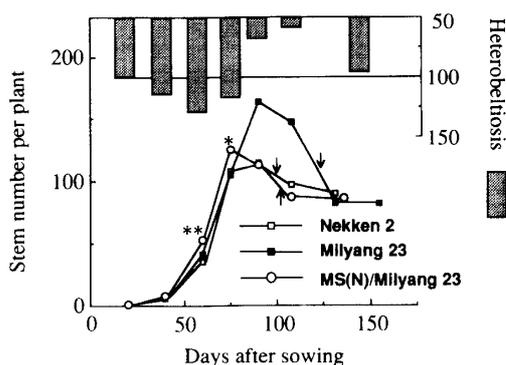


Fig. 4. Comparison of changes in stem number of F_1 hybrid and parents (Experiment 2). \circ with * and ** indicate that F_1 hybrid statistically surpassed parents at 5% and 1% level (t-test), respectively. \circ without * shows not significant. Arrows show the heading days.

weight with the number of nodal roots, was larger than that of parents (Fig. 1).

The ratios of leaf area and shoot dry weight to number of nodal roots are shown in Fig. 2. Both leaf area and shoot dry weight of a single nodal root of F_1 hybrids were always greater than those of parents. Moreover, N_{no} of both F_1 hybrids always exceeded that of parents (Fig. 3). These results can be interpreted that the function of a single nodal root such as water and nutrient uptake is higher in F_1 hybrids than parents. N_{dw} of F_1 hybrids, however, was similar to that of parents (Fig. 3).

2. Experiment 2

Fig. 4 shows time-course changes in stem number of pot-grown F_1 hybrid (MS(N)/Milyang 23) and parents. Stem numbers of F_1 hybrid on Day 60 and Day 75 were greater than those of parents. Then, that of Milyang 23 still increased while that of Nekken 2 and F_1 hybrid decreased gradually, and the number on Day 90 was the highest in Milyang 23 followed by F_1 hybrid and Nekken 2. However, the final panicle number of the three varieties was similar. F_1 hybrid also exceeded parents in the number of nodal roots from Day 30 to Day 90, although not necessarily significant (Fig. 5). Number of nodal roots of the plants reached maximum on Day 108, and the order among the varieties was the same with that of the stem number. Thereafter, number of nodal roots of the plants gradually decreased toward full ripening.

Eventually, shoot dry weight of F_1 hybrid

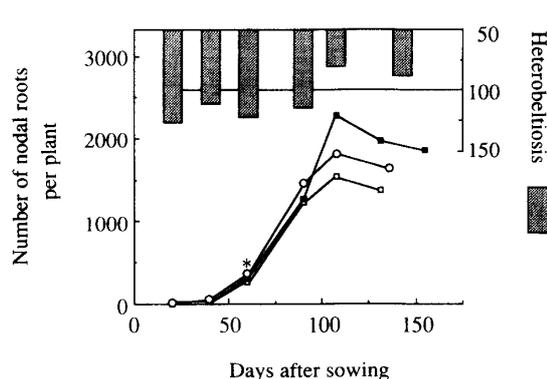


Fig. 5. Comparison of changes in number of nodal roots of F_1 hybrid and parents (Experiment 2). Symbols \square , \blacksquare , \circ , and *, see Fig. 4.

became significantly greater than that of parents from Day 30 to Day 108, and the heterobeltiosis attained 180 on Day 60. In addition, shoot dry weights of Milyang 23 and F_1 hybrid at full ripening were the same, although the growth period was 19 days longer in Milyang 23 (Fig. 6).

Fig. 7 indicates the dry weight of a single nodal root. The dry weight of F_1 hybrid was heavier than that of parents from Day 40 to full ripening of F_1 hybrid. However, the dry weight of Milyang 23 showed marked increase during late ripening stage. Since Milyang 23 belongs to late-maturing variety in Hokuriku district, the mean temperature during the stage was comparatively low (22.4°C). Therefore, surplus photosynthetic assimilates which were not translocated to panicles³⁰⁾ may have been accumulated in the root system.

Discussion

F_1 rice hybrids used in Experiment 1 exhibited obvious hybrid vigor for shoot growth during vegetative growth. The mean heterobeltiosis for shoot dry weight was 116 (Table 1), whereas the stem number of F_1 hybrids on Day 45 was almost the same as that of parents. The numbers of nodal roots of F_1 hybrids were also similar to those of parents during the experimental period. On the other hand, F_1 hybrid selected for Experiment 2 was greater in the number of both stems and nodal roots than parents on Day 60 (Figs. 4 and 5), and heterobeltiosis for shoot dry weight reached 180 at that time (Fig. 6). Accordingly, an increase in stem number should result in an increase of nodal root number in F_1 hybrid,

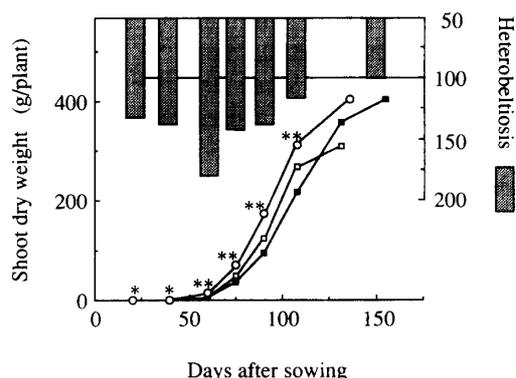


Fig. 6. Comparison of changes in shoot dry weight of F₁ hybrid and parents (Experiment 2). Symbols □, ■, ○, * and **, see Fig. 4.

and the degree of heterosis could become maximum during vegetative growth. The depression of tillering in Experiment 1 must be due to high temperature in the greenhouse¹⁷⁾.

Kawano et al.¹³⁾ reported that 22 out of 33 F₁ rice hybrids, which were grown in culture solution for seven weeks after transplanting, surpassed parents in the partitioning rate of dry matter to roots. In this study, however, S/R ratio of F₁ hybrids was the same as for either parent or in between parents (Table 1). This means the partitioning rate of dry matter to root system of japonica/indica F₁ rice hybrids is much the same as that of parents during vegetative growth.

On the other hand, the dry weight of a single nodal root of F₁ hybrids exceeded that of parents from young seedling to full ripening stage with one exception (Figs. 1 and 7), whether the number of nodal roots of F₁ hybrids and parents was similar or not. According to our preliminary experiment where MS1003/Milyang 23 and parents were grown in 0.2, 2.0 and 20.0ppm NH₄-N of culture solution, the F₁ hybrid was from 1.2 to 1.3 times larger in cross sectional area of nodal root axes, and was from 1.0 to 1.2 times longer in nodal axis length than better parent. This indicates that the F₁ hybrids used partition more dry matter to each nodal root than parents, and form thicker and longer nodal root axes.

Ichii and Nakamura⁹⁾ showed that most of 25-day-old F₁ hybrids of japonica/japonica combination were higher than parents in NH₄-N, NO₃-N, P and K uptake rates (whole plant dry weight basis), whereas in whole

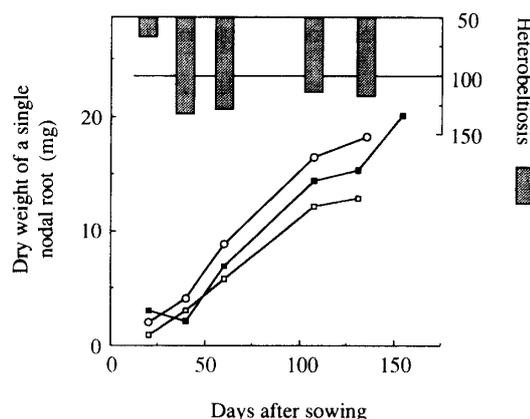


Fig. 7. Comparison of changes in dry weight of a single nodal root of F₁ hybrid and parents (Experiment 2). Symbols □, ■, ○, see Fig. 4.

plant dry weight the F₁ hybrids were much the same as either parent. However, N_{no} as well as leaf area and shoot dry weight of a single nodal root in F₁ hybrids was greater than those of parents in this study (Figs. 2 and 3). These results strongly suggest that in japonica/indica F₁ hybrids more photosynthate is partitioned to each nodal root, so that this enhances its function and consequently, the entire root system.

Suzuki et al.²⁸⁾ reported that nitrogen uptake rate (whole plant dry weight basis) was much the same between F₁ hybrids and parents irrespective of japonica/japonica or japonica/indica combinations. They suggested that higher nitrogen uptake rate per plant of F₁ hybrids may not be due to higher uptake rate per dry weight, but due to their larger weight as a whole plant. In addition, in the present study, japonica/indica F₁ hybrids exhibited similar value to parents in N_{dw} during vegetative growth.

It is well-known that in gramineous crop plants which form fibrous root system lateral roots greatly contribute to the total length and surface area of the root system^{6,32)}, although their dry weight may not be in large proportion. Therefore, it is indispensable to know how F₁ rice hybrids utilize more dry matter partitioned to each nodal root to produce lateral roots. However, no quantitative analysis of the root system in F₁ rice hybrids seems to be conducted. Thus, further study is needed to quantify development of the root system in F₁ rice hybrids.

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References

1. Akita, S. 1988. Physiological bases of heterosis in rice. In Hybrid Rice. International Rice Research Institute, Manila, Philippines. 67—77.
2. Akita, S., L. Blanco and K. Katayama 1990. Physiological mechanism of heterosis in seeding growth of indica F₁ rice hybrids. Jpn. J. Crop Sci. 59 : 548—556.
3. Araki, H., K. Toya and H. Ikehashi 1988. Role of wide-compatibility genes in hybrid rice breeding. In Hybrid Rice. International Rice Research Institute, Manila, Philippines. 79—83.
4. Cheng, Q., N. Hu, N. Jing and F. Huang 1988. Physiological and biochemical characters of hybrid rice in Dongting Lake region, China. In Hybrid Rice. International Rice Research Institute, Manila, Philippines. 279.
5. Deng, H. 1988. Biochemical basis of heterosis in rice. In Hybrid Rice. International Rice Research Institute, Manila, Philippines. 55—66.
6. Dittmer, H. J. 1937. A quantitative study of the roots and root hairs of a winter rye plant (*Secale cereale*). Am. J. Bot. 24 : 417—420.
7. Ekanayake, I. J., D. P. Garrity and S. S. Virmani 1986. Heterosis for root pulling resistance in F₁ rice hybrids. Int. Rice Res. News. 11 : 6.
8. Ichii, M. and M. Nakamura 1987. Macronutrient uptake in F₁ rice seedlings. Japan. J. Breed. 37 (Supp. 1) : 224—225*.
9. Ichii, M. and M. Nakamura 1990. Heterosis for nutrient uptake in F₁ rice hybrid-seedlings. Jpn. J. Crop Sci. 59 : 140—145***.
10. Ichii, M. and M. Sako 1987. Heterosis of macronutrient uptake in F₁ hybrid rice plant. Jpn. J. Breed. 37 (Supp. 2) : 276—277*.
11. Jones, J.W. 1926 Hybrid vigor in rice. J. Am. Soc. Agron. 18 : 423—428.
12. Kasugai, S. 1939. A study on the method of water culture. J. Soil Sci. Manure, Jpn. 13 : 669—822****.
13. Kawano, K., K. Kurosawa and M. Takahashi 1969. Heterosis in vegetative growth of the rice plant. Jpn. J. Breed. 19 : 335—342.
14. Kim, C.H. and J.N. Rutger 1988. Heterosis in rice. In Hybrid Rice. International Rice Research Institute, Manila, Philippines. 39—54.
15. Lin, S.C. and L.P. Yuan 1980. Hybrid rice breeding in China. In Innovative Approaches to Rice Breeding. International Rice Research Institute, Manila, Philippines. 35—51.
16. Lu, D. 1988. Relations between physiological heterosis of root and shoot systems of hybrid rice. In Hybrid Rice. International Rice Research Institute, Manila, Philippines. 281—282.
17. Matsuba, K., A. Kusuda and H. Hasegawa 1990. Heterosis for tillering ability in F₁ rice seedlings. Jpn. J. Crop Sci. 59 (Extra issue 2) : 9—10**.
18. Nakayama, M. 1986. Root function of F₁ rice hybrids during ripening stage. Jpn. J. Crop Sci. 55 (Extra issue 1) : 16—17**.
19. Radford, P. J. 1967. Growth analysis formulae—Their use and abuse. Crop Sci. 7 : 171—175.
20. Sahai, V.N. and R.C. Chaudhary 1986. Root system in hybrid rice. Int. Rice Res. News. 11 : 13—14.
21. SAS Institute 1988. SAS/STAT User's Guide (Release 6.03 Edition). SAS Institute Inc. 125—154, 549—640, 941—947.
22. Scheiner, D. 1976. Determination of ammonia and Kjeldahl nitrogen by indophenol method. Water Res. 10 : 31—36.
23. Sinha, S.K. and R. Khanna 1975. Physiological, biochemical, and genetic basis of heterosis. Adv. Agron. 27 : 123—174.
24. Song, X., W. Agata and Y. Kawamitsu 1990. Studies on dry matter and grain production of F₁ hybrid rice in China. I. Characteristics of dry matter production. Jpn. J. Crop Sci. 59 : 19—28***.
25. Song, X., W. Agata and Y. Kawamitsu 1990. Studies on dry matter and grain production of F₁ hybrid rice in China. II. Characteristics of grain production. Jpn. J. Crop Sci. 59 : 29—33***.
26. Song, X., W. Agata and Y. Kawamitsu 1990. Studies on dry matter and grain production of F₁ hybrid rice in China. III. Grain production character from the view point of time changes in non-structural carbohydrate and nitrogen contents during the yield production. Jpn. J. Crop Sci. 59 : 107—112***.
27. Suzuki, Y. 1988. Physiological and ecological characteristics of F₁ rice hybrids, and problems to practical use. Japan. J. Soil Sci. Plant Nutr. 59 : 511—520*.
28. Suzuki, Y., H. Yoshida and M. Morooka 1988. Heterosis for rate of nitrogen uptake in F₁ rice hybrids. Soil Sci. Plant Nutr. 34 : 87—95.
29. Tatsumi, J., A. Yamauchi, T. Nonoyama and Y. Kono 1988. Evaluation of soil contamination on the washed root samples of cereal crops under pot experiment. Jpn. J. Crop Sci. 57. 65—70***.

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30. Tajima, K. and T. Washio 1982. Productivity of indica high-yielding varieties in relation to climatic factors. *Jpn. J. Crop Sci.* 51 (Extra issue 2) : 17—18**.
31. Toriyama, K and H. Ishida 1987. Method of estimating time of NH₄-N disappearance in paddy field by soil solution analysis. *Jpn. J Soil Sci. Plant Nutr.* 58 : 747—749*.
32. Yamauchi, A., Y. Kono and J. Tatsumi 1987. Quantitative analysis on root system structures of upland rice and maize. *Jpn. J. Crop Sci.* 56 : 608—617.
33. Yang, H. and X. Sun 1988. Physiological characteristics of hybrid rice roots. In *Hybrid Rice*. International Rice Research Institute, Manila, Philippines. 281.
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- * In Japanese with English title.
** In Japanese and translated into English by the present authors.
*** In Japanese with English abstract.
**** In Japanese with German abstract.
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