

*Critical Review***The Role of Sigma Receptors in Depression**Jordanna E. Bermack<sup>1</sup> and Guy Debonnel<sup>1,\*</sup><sup>1</sup>Department of Psychiatry, McGill University, 1033, Pine Avenue West, Suite 207, Montréal, Québec, Canada H3A 1A1

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**Abstract.** Behavioral models used to test potential antidepressants have shown that ligands that bind to sigma receptors possess “antidepressant-like” properties. The focus of this review is to discuss the literature concerning sigma receptors and their ligands, with respect to their antidepressant properties. In addition to the behavioral data, we discuss electrophysiological and biochemical models demonstrating sigma receptors’ ability to modulate important factors in the pathophysiology of depression and/or the mechanisms of action of antidepressants such as the serotonergic neurotransmission in the dorsal raphe nucleus (DRN) and the glutamatergic transmission in the hippocampus. We also discuss the significance of these two systems in the mechanism of action of antidepressants. Sigma ligands have potential as antidepressant medications with a fast onset of action as they produce a rapid modulation of the serotonergic system in the DRN and the glutamatergic transmission in the hippocampus. As these effects of sigma ligands may produce antidepressant properties by completely novel mechanisms of action, they may provide an alternative to the antidepressants currently available and may prove to be beneficial for treatment-resistant depressed patients.

**Keywords:** sigma receptor, major depression, antidepressant, serotonin, glutamate, calcium regulation

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**1. Introduction and background**

Since their discovery in 1976 (1), sigma receptors have remained an intriguing entity. As receptors they

present very original aspects but are subjects of controversy regarding their identity(ies), their endogenous ligand(s), their role(s), and so on. Nearly thirty years after their initial description, a consensus has not yet been obtained, but some aspects have started to be more clearly established.

Sigma receptors were first classified as a subtype of opioid receptors (1). Later, the availability of newer and

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more selective ligands established sigma receptors as distinct from opiate receptors or the phencyclidine (PCP) binding site (2). Based on the pharmacological characteristics of several sigma ligands [sometimes not very selective such as the classical antipsychotic haloperidol (3)], sigma receptors were then further divided into two subtypes, denoted sigma<sub>1</sub> and sigma<sub>2</sub> (2). However, this classification remains unsatisfactory and there is debate over the existence of a third subtype (sigma<sub>3</sub>) or an heterogeneity within the sigma<sub>1</sub> subtype. For example, radioligand binding studies with the prototypal sigma ligand [<sup>3</sup>H](+)-pentazocine (4) have demonstrated that its binding sites are heterogenous. Specifically, (+)-pentazocine labeled a high and a low affinity site in several cell lines and the lower affinity site did not correspond to sigma<sub>2</sub> receptors (5, 6). In addition, a 3rd, intermediate site ( $K_d = 30 - 60$  nM) was observed in human cell lines, suggesting a possible 3rd type of (+)-pentazocine binding site (5).

In keeping with the binding data, various electrophysiological data in the model of the modulation of the *N*-methyl-D-aspartate (NMDA) response of dorsal hippocampal pyramidal neurons from our laboratory suggest that the sigma<sub>1</sub> receptor does not represent a single entity. They indicate that (+)-pentazocine acts on a subtype of the sigma receptor different from that of other sigma<sub>1</sub> ligands such as JO-1784 (igmesine) (7), L-687,384 (8), and BD-737 (9). Moreover, Tsao and Su (10) purified a naloxone-sensitive, haloperidol-sensitive (+)-SKF-10,047 binding protein, which actually resembles the sigma opioid site originally proposed by Martin et al. (1), with high affinity for (+)-pentazocine but not for the other sigma ligands DTG (11), (+)-3-PPP (12), and progesterone (13).

### 1.1. Anatomical distribution of sigma receptors in the central nervous system (CNS)

Initial binding investigations, using the nonselective sigma ligand (+)-3-PPP, found the highest densities of binding in the spinal cord, pons-medulla, cerebellum, central gray, red nucleus, and hippocampus with moderate densities detected in the hypothalamus and cerebral cortex and low densities in the basal ganglia and thalamus (14).

Studies comparing sigma<sub>1</sub>- versus sigma<sub>2</sub>-receptor distributions found that sigma<sub>1</sub> sites were most abundant in the dentate gyrus of the hippocampus, facial nucleus, thalamic, and hypothalamic nuclei, with moderate densities found in the striatum, cerebellum, dorsal raphe nucleus (DRN), and locus coeruleus (15 - 17). In agreement, studies of sigma<sub>1</sub>-receptor mRNA (messenger ribonucleic acid) levels found high levels of expression in all layers of the cerebral cortex, striatum, hippo-

campus, and cerebellum (18). Sigma<sub>2</sub> sites were found to be enriched in the substantia nigra, central gray matter, oculomotor nuclei, cerebellum, nucleus accumbens, amygdala, olfactory bulb, hippocampus, and motor cortex (15, 16, 19).

### 1.2. Potential endogenous ligands for sigma receptors

Early studies found that neurosteroids bind to sigma<sub>1</sub> receptors (20 - 22). For example, the neurosteroids progesterone and dihydroepiandrosterone sulfate (DHEAS) dose-dependently inhibit the *in vivo* binding of [<sup>3</sup>H]-SKF-10,047, progesterone being the most potent (20 - 22).

These binding data led to the hypothesis that progesterone might be the endogenous ligand for sigma<sub>1</sub> receptors. This remains however controversial as the affinity of progesterone for sigma receptors does not appear very high for an endogenous ligand. Indeed, Schwarz et al. (23) argued that the concentration of endogenous progesterone, specifically the free serum concentration, was insufficient to occupy the sigma receptors in the brain, even during later pregnancy, when progesterone levels are at their highest. However, there are data suggesting that progesterone binds to sigma receptors under physiological conditions. It has been demonstrated that alterations in endogenous hormonal levels [e.g., adrenalectomy, castration, or ovariectomy (ADX, CX, or OVX, respectively), pregnancy, etc.] affect sigma ligands activity in several models such as the electrophysiological model of the modulation of the NMDA response in the hippocampus (24, 25) and the "antidepressant-like" effects of sigma ligands in behavioral models of depression (26). Moreover, radioligand binding studies show a 30 - 40% decrease in [<sup>3</sup>H]SKF-10,047 binding during pregnancy, while ADX/CX enhanced [<sup>3</sup>H]SKF-10,047 binding, whereas a subsequent treatment with finasteride, which increases progesterone levels, produced decreased [<sup>3</sup>H]SKF-10,047 binding (22, 27).

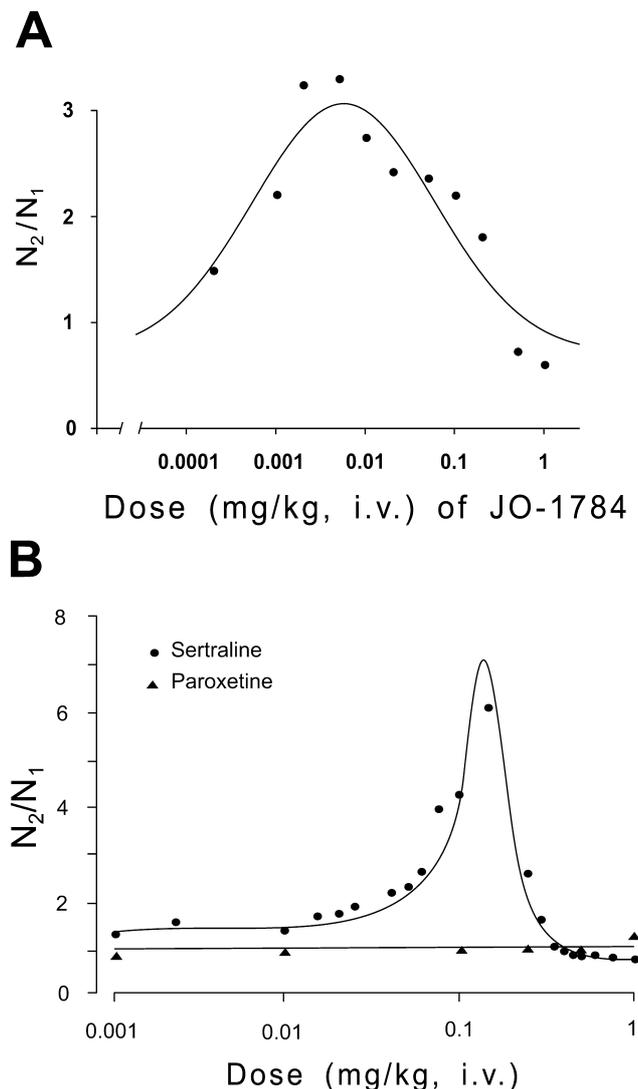
At this point, the literature certainly supports the hypothesis that the endogenous ligand of sigma receptors could be a neurosteroid; however, it remains to be determined if it is really progesterone or perhaps one of its metabolites.

### 1.3. Pharmacology of sigma receptors

An interesting feature of sigma receptors is that in contrast with the classical pharmacology of many compounds that show a more less linear dose-response curve followed by a plateau effect, a biphasic bell-shaped dose response curve has been observed for sigma ligands in various behavioral, biochemical, and electrophysiological paradigms (28 - 31). For example, because of the

bell-shaped dose-response curves, in the electrophysiological paradigm of the modulation of the NMDA response (see below), low doses of sigma agonists induce a potentiation of the NMDA response (32, 33). At higher doses, the effects of sigma agonists such as DTG and JO-1784 progressively decrease and disappear (Fig. 1) and these molecules act as antagonists by preventing the potentiation induced by low doses of other sigma agonists (29). A similar bell-shaped dose-response curve has also been described with sigma

ligands in other models such as in release experiments (30) and in behavioral models (28, 34). The exact reason for such bell-shaped dose-response curves obtained in so many models have not been well established. It has been proposed that they may be due to the fact that low doses of sigma ligands activate one subtype of sigma receptors for which they have high affinity, whereas higher doses may activate another/other subtype(s) of the sigma receptor for which they have a lower affinity, which would counteract the effects observed at lower doses (29, 35, 36). Nonetheless, it is important to note that the different (and sometimes opposite) results obtained with low and high doses of sigma ligands could constitute a very important factor to explain much of the controversy seen in the literature on sigma receptors.



**Fig. 1.** Dose-response curve of the effects of intravenously administered igmesine (A) and of sertraline and paroxetine (B) on the neuronal activation of CA<sub>3</sub> dorsal hippocampus pyramidal neurons induced by microiontophoretic applications of NMDA. The effects of these drugs were assessed by determining the ratio ( $N_2/N_1$ ) of the number of spikes generated/nC of NMDA before ( $N_1$ ) and after ( $N_2$ ) the injection of the drug. One dot represents the effect of one dose of drug administered to one rat while recording from one neuron. Adapted from refs. 29 and 35.

#### 1.4. Cloning of the $\sigma_1$ receptor

A significant breakthrough in sigma receptor research was the cloning of the  $\sigma_1$  receptor. It was cloned from guinea pig and mouse liver, human placental cell line and brain, and mouse and rat brain (37–41). The protein cloned was a 223 amino acid, 1 transmembrane protein with potent (+)-pentazocine, haloperidol, DTG, and (+)-3-PPP binding, but which did not couple with G-proteins (41, 42). The amino acid sequence of the sigma receptor cloned from the rat brain was highly homologous to the sigma receptor cloned from guinea pig liver and human placental cell line, but was not related to other known neurotransmitter receptors (37).

At this point, it is not completely clear whether the cloned  $\sigma_1$  receptor is the ligand binding subunit of a multi-subunit complex or represents one subtype of the  $\sigma_1$  receptor. Regardless, the cloning has led to an important focus on the molecular biology and signal transduction mechanisms of  $\sigma_1$  receptors. However, given the one-transmembrane segment cloned, it is most likely that it does not represent the complete functional receptor, but more experiments using techniques such as the use of selective  $\sigma_1$  receptor gene antisense will elucidate the exact structure of the functional sigma receptor in the future. A recent study investigating putative transmembrane segments based on homology identified two putative transmembrane segments for the  $\sigma_1$  receptor (43). Therefore, the exact structure of the  $\sigma_1$  receptor has yet to be fully elucidated.

The cloning of the  $\sigma_1$  receptor led to the development of  $\sigma_1$ -receptor knockout mice (44). Preliminary studies of these mice determined that they display no significant phenotypic and no behavioral abnormalities under baseline conditions; however, further studies in various paradigms will be required to produce more insight into the functional roles of  $\sigma_1$  receptors.

## 2. Signal transduction by sigma receptors: interaction with G-proteins

The cloning of a one transmembrane domain sigma receptor, which could not correspond to a G-protein coupled receptor, reactivated the long-standing debate over whether or not sigma receptors act through G-protein-dependant signaling cascades. In support of sigma receptors' association with G-proteins, manipulating GTP and 5 guanylylimidodiphosphate [Gpp(NH)p] altered the binding of sigma ligands (45–47). Furthermore, there was a decreased responsiveness to guanine nucleotides following chronic treatments (48). Moreover, some selective sigma agonists were found to stimulate GTPase activity (49). Manipulation of G-proteins also altered sigma-mediated effects on potassium ( $K^+$ ) currents (50), and NMDA-evoked release of [ $^3H$ ]norepinephrine (NE) (51) and suppressed the neuronal response to NMDA (52), but had no effects on  $K^+$  currents in other models or on the NMDA response with other sigma ligands (52–54). Contrasting results were also found for the effects of G-proteins on sigma ligand binding (55–57).

In conclusion, given the presumed heterogeneity of the sigma<sub>1</sub>-receptor subgroup, it is likely that one subtype of the sigma<sub>1</sub> receptor interacts with G-proteins, while another subtype relies on G-protein-independent signal transduction mechanisms.

### 2.1. Modulation of $K^+$ currents

An interaction between sigma receptors and  $K^+$  channels was suggested by the observation that the sigma ligands DTG and (+)-pentazocine inhibited  $K^+$  currents (50, 53, 54). Further investigations of this modulation suggested that a protein-protein interaction is the likely mechanism of signal transduction by sigma receptors as sigma ligands did not interact directly with  $K^+$  channels (43, 54). Furthermore, surprisingly sigma receptors were able to alter channel function in the absence of sigma ligands, suggesting that they are directly associated with  $K^+$  channels, in a manner that can affect current flow, an effect which is enhanced in the presence of sigma ligands. Therefore, it has been suggested that sigma receptors may serve as auxiliary subunits to voltage-gated  $K^+$  channels in the plasma membrane (43), which may involve other proteins such as ankyrin and inositol-1,4,5-trisphosphate ( $IP_3$ ) receptors as suggested by Hayashi and Su (58).

### 2.2. Modulation of $Ca^{2+}$ currents

An interaction between sigma receptors and  $Ca^{2+}$  channels was suggested by the observation that (+)-pentazocine inhibited the rise in  $Ca^{2+}$  levels induced by

depolarization and that sigma ligands decreased basal intracellular  $Ca^{2+}$  concentration ( $[Ca^{2+}]_i$ ), suggesting that sigma receptor activation alone affects  $[Ca^{2+}]_i$  (59, 60). Sigma-induced increases in  $Ca^{2+}$  currents develop progressively following relatively long lasting applications of sigma ligands, suggesting a direct intracellular coupling of sigma receptors to  $Ca^{2+}$  channels, through which sigma ligands can stimulate voltage-activated  $Ca^{2+}$  conductances independent of the  $K^+$  channel pathway (61).

Recently, several lines of evidence added further arguments for the involvement of sigma<sub>1</sub> receptors in  $Ca^{2+}$  signaling. Specifically, the sigma<sub>1</sub> ligands (+)-pentazocine and PRE-084 modulate  $Ca^{2+}$  signaling in NG108 cells via sigma<sub>1</sub> receptors by 2 different modes of action. Firstly, intracellularly, perhaps on the endoplasmic reticulum (ER), sigma ligands potentiate the bradykinin-induced increase in cytosolic free  $Ca^{2+}$  in a bell-shaped manner, which was blocked by sigma<sub>1</sub>-receptor antisense (31). A second mode of action at the plasma membrane was demonstrated as (+)-pentazocine inhibits (in agreement with the effects observed in forebrain synaptosomes) (59, 60), while PRE-084 potentiates the depolarization-induced changes in  $Ca^{2+}$ , effects blocked by sigma<sub>1</sub>-receptor antisense (31).

It has been proposed that the modulation of  $Ca^{2+}$  signaling described thus far involves the formation of a multi-protein complex. Specifically, sigma<sub>1</sub> receptors have recently been found to anchor ankyrin (a cytoskeletal adaptor protein) to the ER membrane and modulate the function of ankyrin and  $IP_3$  receptor-3 on the ER (58). In this model, the presence of the sigma agonist (+)-pentazocine leads to the sigma<sub>1</sub> receptor/ankyrin complex dissociating from the  $IP_3$  receptor-3. This dissociation leads to an increased binding of  $IP_3$ , which in turn increases  $Ca^{2+}$  efflux. On the other hand, in the presence of the sigma<sub>1</sub> antagonist NE-100 (62), the sigma<sub>1</sub> receptor dissociates from ankyrin, which remains coupled to  $IP_3$  receptor-3 on the ER (58). This modulation of  $Ca^{2+}$  currents has important implications towards various cell functions including neurotransmitter release.

The data on the modulation of  $Ca^{2+}$  signaling by sigma receptors, similar to those with  $K^+$  currents, suggests that sigma<sub>1</sub> receptors form multiunit complexes responsible for the modulation of ion channels. This formation of complexes with other proteins enables sigma ligands to exert a wide variety of actions in the CNS, so depending on the different combination of proteins formed, it is likely that different effector systems are activated, which then lead to different results. This hypothesis could also explain some of the discrepancies with respect to different signaling pathways observed to be

involved with the effects of sigma<sub>1</sub> ligands.

### 3. Glutamate and depression

Glutamate is the most widespread excitatory neurotransmitter in the CNS. Glutamate receptors have been pharmacologically classified as ionotropic and metabotropic receptors [reviewed by Bleich et al. (63)]. Ionotropic glutamate receptors include NMDA,  $\alpha$ -amino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) and kainate families of receptors. For the purpose of this review, we will focus on NMDA and AMPA receptors, as they have been linked to depression and/or antidepressant action (see below). NMDA receptors control channels that allow Na<sup>+</sup> and Ca<sup>2+</sup> ion influx and AMPA receptors allow the passage of Na<sup>+</sup> and K<sup>+</sup> ions. Thus, both of these ionotropic glutamate receptors mediate excitatory neurotransmission, and their activation results in cell membrane depolarization [reviewed by Seeburg (64)].

There are several series of data suggesting an altered glutamatergic neurotransmission in depression [reviewed by Sanacora et al. (65)]. Specifically, it has been shown that glutamate metabolism differs significantly between depressed patients and controls when assessing glutamate and/or glutamine levels in plasma or cerebrospinal fluid and when examining platelet intracellular Ca<sup>2+</sup> release in response to glutamate stimulation (66–70). Furthermore, some of these differences have been resolved by chronic antidepressant treatments (71, 72). Magnetic resonance imaging methods have shown reduced glutamate levels in the anterior cingulate cortex (73, 74), which were returned to normal levels following successful antidepressant treatments (74, 75). Thus, an altered glutamatergic neurotransmission appears to be present, but recent data tend to suggest that NMDA and AMPA receptors could be involved in different ways in the pathophysiology of depression.

Preclinical tests for antidepressant efficacy have shown “antidepressant-like” properties of NMDA-receptor antagonists (76, 77), including functional antagonists and ligands for the glutamate, glycine, polyamine, bivalent cation, and ionophore recognition sites of the NMDA receptor complex (76, 78–80). The “antidepressant-like” effects of NMDA antagonists likely involve NMDA receptors located in the hippocampus as the NMDA antagonist 2-amino-7-phosphonoheptanoic acid (AP-7), when injected directly into the hippocampus, decreases immobility time in the forced swimming test (FST) (81).

Conversely, classical antidepressant treatments have been linked to changes in NMDA-receptor properties. For example, antidepressant drugs produce a time- and

dose-dependent change in the radioligand binding properties of the NMDA receptor (82–91). These changes are present with all antidepressant classes (87) and may involve an alteration in the expression of NMDA-receptor subunits (92, 93). Chronic antidepressant treatments can also alter NMDA-receptor function (94–96).

In humans, NMDA-receptor abnormalities have been observed in suicide victims (97). A decrease in the expression of the subunit 1 of the NMDA-receptor mRNA has also been documented in the hippocampus of depressed patients (98). Clinical studies with the NMDA antagonists amantadine, memantine, and ketamine have shown efficacy in depression (99–102). Therefore, these data suggest that a dysfunction of the NMDA receptor could be involved in the pathophysiology of major depression and that drugs that modulate NMDA mediated neurotransmission may possess “antidepressant-like” properties.

Recently, AMPA receptors have also been implicated in antidepressant effects, due to the observation that the AMPA receptor potentiator LY392098 possesses “antidepressant-like” properties in the FST and in the tail suspension test. LY392098 alone dose-dependently decreased immobility similarly to classical antidepressants and at sub-threshold doses, significantly increased the potency of other antidepressants (103). In primary neuronal cultures, LY392098 also increased brain derived neurotrophic factor (BDNF) mRNA (104), which is suggested to play a critical role in the action of antidepressants (105–107). In addition, treatments with classical antidepressants influence AMPA receptors. Specifically, chronic antidepressant treatments increase the expression of AMPA receptors in hippocampal membranes (108) and increase the phosphorylation of AMPA receptor subunits (109), which may indicate an enhanced AMPA-mediated synaptic transmission.

Alternatively, the involvement of glutamate receptors in antidepressant action may originate from a common downstream effect. For example, a downstream effect of the activation of glutamate receptors is an increase in Ca<sup>2+</sup> levels and, in turn, of nitric oxide (NO) synthesis (110). NO may be involved in the relationship between antidepressants and glutamate transmission, as a potential role for NO in affective disorders has been proposed (111, 112). Antidepressants can inhibit the activity of neuronal NO synthase (nNOS) (113, 114). Accordingly, NOS inhibitors are active in acute and chronic preclinical antidepressant screening paradigms (115–118).

### 4. Serotonin and depression

Serotonin (5-HT) receptors are made up of at least 14

different subtypes [reviewed by Barnes and Sharp (119)]. The 5-HT<sub>1A</sub>-receptor subtype is of particular importance in the regulation of 5-HT neurons' activity, as somatodendritic 5-HT<sub>1A</sub> autoreceptors exert auto-regulatory control of the firing activity of 5-HT neurons (120, 121).

In addition to 5-HT<sub>1A</sub> autoreceptors, high densities of 5-HT<sub>1A</sub> receptors located postsynaptically are found in the hypothalamus, amygdala, hippocampus, lateral septum, and medial prefrontal cortex (mPFC) (122–125). Activation of 5-HT<sub>1A</sub> receptors by 5-HT or 5-HT<sub>1A</sub> agonists leads to membrane hyperpolarization and a suppression of the neuronal firing activity (126–131). Other 5-HT receptors have also been implicated in depression, but this review will focus on only the 5-HT<sub>1A</sub> receptors and the effects of antidepressants on this receptor population.

The 5-HT hypothesis of depression originated in the 1960's and proposed that low levels of 5-HT in certain brain receptors led to depression (132). It was then modified to propose that a deficiency in brain 5-HT increased one's vulnerability to depression, as suggested by the observations that interference with the 5-HT system or storage may induce depression in vulnerable individuals and that antidepressants enhance central 5-HT neurotransmission [reviewed by Maes and Meltzer (133)].

Tryptophan depletion studies provided further support for the importance of the 5-HT system in antidepressant activity, but not necessarily in the pathophysiology of depression. In remitted depressed patients receiving selective serotonin reuptake inhibitors (SSRIs), acute L-tryptophan depletion led to a rapid return of depressive symptomatology. However, remitted patients maintained with tricyclics were less prone to relapse following

tryptophan depletion (134, 135).

A more recent hypothesis is that a simple deficiency of 5-HT is not the sole cause of depression and that depression likely involves dysfunction in brain areas that are modulated by monoaminergic systems such as the frontal cortex and the hippocampus [reviewed by Delgado and Moreno (136)].

There are convincing lines of evidence suggesting a role of the 5-HT system in the mechanism of action of antidepressants. Overall, electrophysiological studies have demonstrated that all known antidepressants including tricyclics, SSRIs, monoamine oxidase inhibitors (MAOIs), electroconvulsive treatments, and even more selective noradrenergic treatments such as desipramine, reboxetine or mirtazapine, enhance 5-HT neurotransmission following chronic treatments, via different mechanisms (137) (Table 1).

Following acute and short-term treatments, SSRIs, similarly to MAOI's, decrease the firing activity of DRN 5-HT neurons (138–140), whereas a chronic administration of these medications leads to a desensitization of 5-HT<sub>1A</sub> somatodendritic autoreceptors in the DRN (137, 141, 142). The proportion of 5-HT neurons with desensitized autoreceptors was shown to increase progressively during the course of several weeks of SSRI administration (143), and in microdialysis studies, the acute administration of SSRIs produces a small, transient increase in extracellular 5-HT concentration in the rat frontal cortex, while 14-day continuous infusion produced a 6-fold increase (144).

Moreover, 14-day treatments with fluoxetine were shown to result in a selective uncoupling of 5-HT<sub>1A</sub> receptors from G-proteins in the DRN, but not in the hippocampus (145). However, in the hippocampus, the sensitivity of postsynaptic 5-HT<sub>1A</sub> receptor-mediated

**Table 1.** Effects of long-term antidepressant treatments on 5-HT neurotransmission

	Responsiveness of somatodendritic 5-HT <sub>1A</sub> autoreceptors	Function of terminal 5-HT <sub>1B</sub> autoreceptors	Function of terminal $\alpha_2$ -adrenergic heteroreceptors	Responsiveness of postsynaptic 5-HT <sub>1A</sub> receptors	Net effect on 5-HT neurotransmission
SSRI	↓	↓	○	○	↑
MAOI	↓	○	↓	○ or ↓	↑
5-HT <sub>1A</sub> agonists	↓	○	n.d.	○	↑
TCA	○	○	↓	↑	↑
ECS	○	○	○	↑	↑
Mirtazapine	○	○	↓	○	↑
NRIs	○	○	↓	↑	↑
NK <sub>1</sub> antagonists	↓	n.d.	↓	○	↑
Sigma agonists	n.d.	n.d.	n.d.	n.d.	↑

↓: decrease, ○: no change, n.d.: not determined, ↑: increase. TCA: tricyclic antidepressants, ECS: electroconvulsive treatments, NRIs: noradrenaline reuptake inhibitors. Adapted from ref. 137.

responses is not changed (138, 140, 142, 146). These adaptive changes in 5-HT neurons may explain the delayed enhancement of 5-HT-mediated transmission, consistent with the clinical onset of action of SSRI's (137).

Overall, antidepressants clearly produce adaptive changes in the 5-HT system following chronic treatments. Whether these changes are key to the therapeutic efficacy of antidepressants remains to be determined. It is possible that alterations in 5-HT neurotransmission may just be one step involved. Similarly to what has been proposed for the glutamatergic system's involvement in antidepressant action, a downstream mechanism may also be necessary and therefore a downstream target common to both the glutamatergic and serotonergic systems might be the common target of antidepressant medications. Regardless, changes in 5-HT transmission in electrophysiological and microdialysis models are predictive of antidepressant potential, and therefore, they can be used as indices to assess potential novel antidepressants.

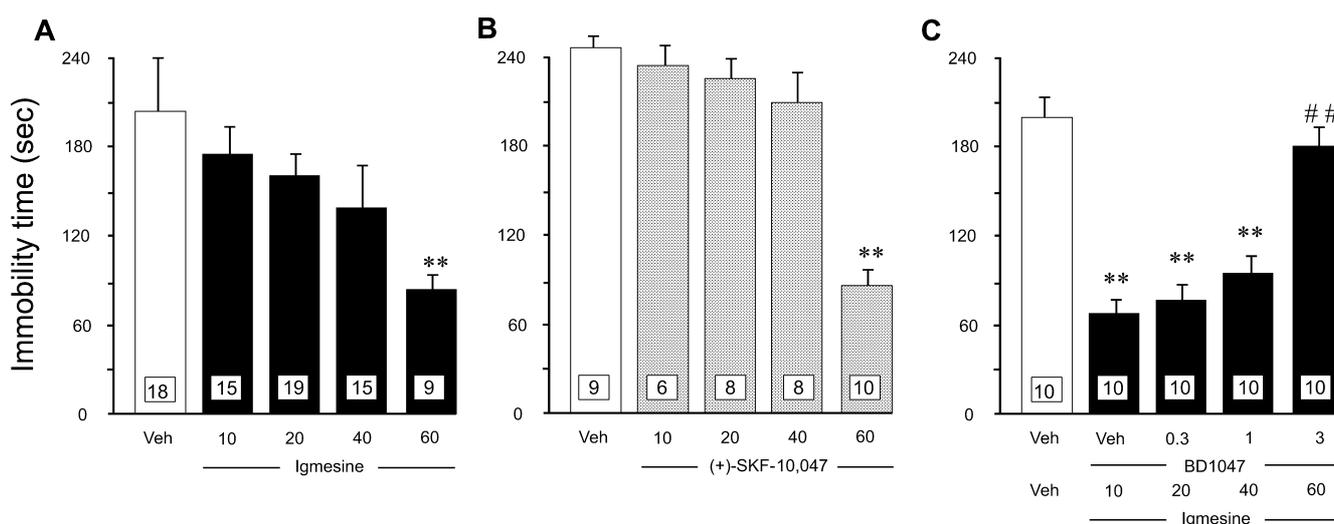
## 5. Sigma receptors and depression

### 5.1. Sigma ligands as antidepressants

The first interest in sigma ligands as antidepressants originated from the observation that the antidepressants fluvoxamine, fluoxetine, citalopram, sertraline, clorgyline, and imipramine all possess moderate to high affinity ( $K_i = 36 - 343$  nM) for sigma<sub>1</sub> sites (147 - 149).

Conversely, antidepressant treatments or modifications of the 5-HT system induced changes in sigma receptors binding properties. Repeated treatments with the tricyclic imipramine (14 days) cause a decrease in the total number of sigma receptor binding sites without affecting the affinity of [<sup>3</sup>H]DTG binding to sigma sites in the striatum, hippocampus, and cortex of the rat (150). Similar reductions were observed after chronic administration of fluoxetine. The absence of effect of desipramine and the fact that a depletion of brain 5-HT by *para*(4)-chloroamphetamine (p-CPA) blocks the ability of imipramine to decrease DTG binding, suggest that 5-HT transmission may play a role in the regulation of cerebral sigma binding sites in the rat. It has been proposed that certain differences in the clinical effects of various antidepressants may, in part, be explained by their distinct influence on cerebral sigma receptors (150).

More direct evidence of potential antidepressant properties of sigma ligands were obtained from behavioral investigations. Sigma agonists have been then tested in various behavioral tests classically used to predict an antidepressant activity. SA 4503 (151), (+)-pentazocine, DTG, JO-1784, and SKF-10,047 dose-dependently decrease immobility in the FST. These effects were blocked by the sigma antagonist NE-100 or BD1047 (151 - 153) (Fig. 2). In addition, SA 4503 and (+)-pentazocine also decreased immobility time in the Tail Suspension Test, an effect also antagonized by NE-100 (154). Interestingly the antidepressant-like



**Fig. 2.** Antidepressant-like effect of sigma<sub>1</sub>-receptor ligands in the forced swimming test in Swiss mice: dose-response effect of igmesine (A), (+)-SKF-10,047 (B), and antagonism by BD1047 of the igmesine-induced effect (C). Drugs were injected i.p. 30 min before the session on day 2. BD1047 was administered i.p. 15 min before igmesine, which was given 30 min before the session on day 2. The duration of immobility was recorded for the last 5 min over a 6-min session. Values are expressed as the mean  $\pm$  S.E.M. of the number of animals indicated inside each column. \*\* $P < 0.01$  versus the vehicle-treated group (Veh), ## $P < 0.01$  versus the igmesine-treated group (Dunnett's test). Adapted from ref. 26.

effect of SA 4503 in the FST was potentiated by the non-competitive NMDA antagonist amantadine (155).

Recently, OPC-14523, a combined  $\sigma_1$  and 5-HT<sub>1A</sub> receptor ligand (156), yielded interesting results in the FST (156, 157). Single doses decreased immobility time and the effect of OPC-14523 was enhanced by its daily administration for 7 days. Both the  $\sigma_1$ -receptor antagonist NE-100 and the selective 5-HT<sub>1A</sub> antagonist WAY 100635 (158) antagonized the behavioural effects of a single dose of OPC-14523 in the FST (157). Moreover, a one-week pretreatment with p-CPA, failed to diminish the antidepressant effects of OPC-14523 in this behavioral test (159), suggesting that sigma receptors alone can mediate the antidepressant effects produced by OPC-14523 and that the combination of the sigma- and 5-HT<sub>1A</sub>-receptor activity could induce a more potent or rapid “antidepressant-like” effect. In keeping with this hypothesis, a potentiation of the “antidepressant-like” effects in the rodent FST has been obtained with the combined administration of sigma and 5-HT<sub>1A</sub>-receptor agonists compared with their separate administration (160). In the chronic mild stress behavioral (CMS) model (chronic stress is believed to be involved in the etiology of affective disorders), the sigma ligands SKF-10,047 and dextrometorphan reversed the motor suppression induced by stress (161, 162).

Most of the data on sigma receptors and depression have focussed on the  $\sigma_1$  receptor; however, the  $\sigma_2$  ligand Lu 28-179 also showed “antidepressant-like” activity in the CMS model of depression. Specifically, three-week treatments with antidepressants led to a normalized sucrose intake, which reversed the decreased intake caused by the stress. Lu 28-179 did not affect sucrose intake in non-stressed controls, but produced a significant increase in sucrose intake in rats exposed to CMS (163). However, even if Lu 28-179's has a higher affinity for  $\sigma_2$  receptors, it also has affinity for  $\sigma_1$  receptors (164); therefore, a role of the  $\sigma_1$  receptors in these “antidepressant-like” effects of Lu 28-179 cannot be excluded.

In animal models, neurosteroids with affinity for sigma receptors have also been shown to exert “antidepressant-like” effects that are dependent on the endogenous neurosteroidal systems. For example, in ADX/CX mice, the effect of JO-1784 in the FST was enhanced compared to control animals, whereas another  $\sigma_1$  agonist, PRE-084 (165), demonstrated a significant antidepressant effect only in ADX/CX mice (26). The  $\sigma_1$ -antagonist BD 10047 (166) blocked all these effects (26). Furthermore, treatments with finasteride, which lead to the accumulation of progesterone, blocked the  $\sigma_1$ -mediated effects. Thus, as discussed pre-

viously, circulating steroids appear to exert a tonic modulatory effect on the  $\sigma_1$  receptor and therefore on  $\sigma_1$  receptor-mediated “antidepressant-like” effects (26). It has thus been proposed that the potency of  $\sigma_1$  agonists as antidepressants is highly dependent on the endogenous progesterone levels. Depressed patients such as the elderly with decreased levels of neurosteroids, which would be tonically inhibiting sigma receptors to a lesser degree, might be particularly sensitive to such treatments (26).

Only a few clinical data are available regarding the effect of sigma ligands in depressed patients. In an open study, a small number of depressed patients were treated with JO-1784 which induced a 50% reduction of intensity of the depressive symptoms on the Hamilton depression scale (J.L. Junien and F. Roman, personal communication). Moreover, the only results from a double-blind placebo controlled study, obtained from an interim analysis, showed that a dose of 20 mg/day of JO-1784 was superior to placebo and to 20 mg/day of fluoxetine. However, at 100 mg/day, JO-1784 was not different from the placebo (167), which is in keeping with the bell-shaped dose-response curves mentioned above (167).

Therefore, even if very limited, the clinical data also support the hypothesis that sigma agonists could be effective antidepressant medications. However, the mechanisms of action, through which sigma ligands could exert their antidepressant effects have not been clearly identified.

## 5.2. Possible mechanisms of action for sigma ligands as antidepressants

### 5.2.1. Sigma receptors and glutamatergic neurotransmission

Numerous studies have shown interactions between sigma receptors and NMDA-receptor-mediated responses. For example, sigma ligands, including haloperidol, (+)-pentazocine, 4-IBP (168), (+)-3-PPP, (+)-SKF-10,047, and DTG, antagonize NMDA receptor currents in *Xenopus* oocytes (169). The effect of sigma ligands on NMDA receptors in this study were thought to be indirect; however, high doses ( $\mu$ M) and/or nonselective sigma ligands were used. Furthermore, there was no correlation between the potency of NMDA-receptor inhibition and the affinity or stereoselectivity for sigma sites (169–171). Thus, it is difficult to assess whether these observations were based on sigma-receptor-mediated actions rather than on non-specific effects. More recently, in vitro radioligand binding studies showed that haloperidol, (+)-pentazocine, DTG, (+)-SKF-10,047, and (+)-3-PPP inhibited [<sup>3</sup>H]TCP binding to NMDA receptors in neuronal cells, with a potency

correlated with the affinity for DTG binding sites (172, 173).

In our laboratory's model of modulation of the NMDA response in dorsal hippocampal pyramidal neurons of the CA<sub>3</sub> region, it was found that low doses of the sigma ligands DTG, JO-1784, (+)-pentazocine, and L-687,384 selectively potentiated the response of these neurons to microiontophoretic applications of NMDA (29, 32, 33, 35, 51). Other sigma ligands like BD-737, 4-IBP, and OPC-14523 were less selective as they also modulate quisqualate (QUIS)-induced response (29, 174) (JE. Bermack and G. Debonnel, unpublished observation). Interestingly, it was also found that depending on the initial level of excitatory response to QUIS and NMDA, sigma agonists could increase or decrease the NMDA- and QUIS-induced responses, thus suggesting a real modulatory role of sigma ligands on the glutamatergic response (174) (JE. Bermack and G. Debonnel, unpublished observation). The potentiation induced by these sigma agonists was suppressed by antagonists such as BMY-14802, (+)-3-PPP, and NE-100 (29, 32, 33, 35). The effects of all sigma<sub>1</sub> agonists on the NMDA response produced a bell-shaped dose-response curve, as previously described (29, 175). As stated above, this particular pharmacological profile could explain the discrepancies observed for the effects of sigma ligands with respect to inhibition versus potentiation on NMDA-receptor-mediated responses, as most *in vitro* studies may have used high doses, at which the sigma ligands were acting as antagonists.

Interestingly, in the model of the modulation of the NMDA response in the dorsal hippocampus, the antidepressants sertraline and clorgyline with a high affinity for sigma<sub>1</sub> receptors potentiate the NMDA response. In contrast, the antidepressants paroxetine and tranylcypromine with low affinity for sigma receptors had no effects on the NMDA response despite their similar monoaminergic profiles to sertraline and clorgyline, respectively. Moreover, the effects of sertraline and clorgyline were suppressed by the sigma antagonist haloperidol but not by spiperone, suggesting that they were likely mediated by sigma receptors (35).

The sigma<sub>2</sub> ligands Lu 28-179 (176) and BD-1008 (177) have also been shown to modulate NMDA-mediated responses. Despite their high affinity for sigma<sub>2</sub> receptors, the doses required were, however, 5-10 times higher than sigma<sub>1</sub> ligands (178). The effects of Lu 28-179 were not blocked by the sigma<sub>1</sub> antagonists NE-100, progesterone, or haloperidol, suggesting that these effects were mediated through sigma<sub>2</sub> receptors (178).

*In vitro* models have also suggested a modulatory role for sigma<sub>1</sub> agonists on NMDA-mediated responses.

For example, JO-1784, BD-737, (+)-pentazocine, and (+)-3-PPP potentiated in a concentration-dependent manner NMDA-induced [<sup>3</sup>H]NE release from preloaded rat hippocampal slices (30, 51, 179), whereas DTG and BD-737 acted as inverse agonists, by concentration-dependently inhibiting the overflow of [<sup>3</sup>H]NE evoked by NMDA. Haloperidol and BD-1063 (166) alone did not modify [<sup>3</sup>H]NE release, but completely prevented the effects of JO-1784, BD-737, (+)-pentazocine, DTG, and (+)-3-PPP (30, 51, 179), whereas DuP734 inhibited that of BD-737 (180).

Neurosteroids acting as sigma agonists have also been shown to modulate NMDA-receptor-mediated effects, as DHEA at low doses potentiates the NMDA response in extracellular recordings from the dorsal hippocampus. The effect of DHEA was blocked by NE-100 and haloperidol (25). In this model, neither pregnenolone nor pregnenolone sulfate modified the NMDA response or acted as antagonists, which may be due to their lower affinity for sigma<sub>1</sub> receptors (13, 22). In this model, both progesterone and testosterone act as antagonists and suppress the potentiation of the NMDA response induced by sigma agonists such as DTG, (+)-pentazocine, and JO-1784. DHEA's response is blocked by pertussis toxin, while progesterone's is not, suggesting again that different sigma<sub>1</sub> subtypes may be involved in the modulation of the NMDA response by neurosteroids (25, 181).

Neurosteroids are also active in the *in vitro* model of NMDA-induced [<sup>3</sup>H]NE release from hippocampal slices. Specifically DHEAS potentiates while pregnenolone sulfate inhibits (inverse agonist effect) NE release (179). Both effects are blocked by haloperidol, progesterone, BD-1063, and by a pertussis toxin pretreatment, suggesting G-protein-dependent sigma<sub>1</sub> receptors are responsible (179).

Furthermore, endogenous hormone levels affect the sigma receptor's modulatory effect on NMDA-mediated responses. For example, 2 weeks following OVX, the potentiation of the NMDA response induced by DTG was significantly greater than in control female rats, suggesting that sigma receptors may be tonically inhibited by endogenous progesterone (24, 181). In agreement, 10 times higher doses of (+)-pentazocine and DHEA were required in pregnant females to potentiate the NMDA response. This reduced effect of sigma agonists in late pregnancy may be due to occupation of sigma receptors by high levels of progesterone, which also supports its potential role as an endogenous ligand for sigma<sub>1</sub> receptors (24, 181).

In keeping with the hypothesis of progesterone endogenously binding to sigma receptors, during the post-partum period, the degree of potentiation of the

NMDA response by DTG, (+)-pentazocine, and DHEA was significantly higher than that observed in control females, and at Days 10–15, the potentiation of the NMDA response returned to control values. This apparent supersensitivity of sigma receptors observed during the post-partum period might be due to the rapid drop of progesterone levels after delivery (182, 183). In keeping with these data, in OVX rats treated for 3 weeks with progesterone ( $1000 \mu\text{g kg}^{-1} \text{day}^{-1}$ ), low doses of DTG, JO-1784, DHEA, or (+)-pentazocine did not induce any potentiation of the NMDA response. The potentiation by these ligands returned to normal after a 5-day washout (25). The necessity of such a long washout before the NMDA response returns to normal following progesterone treatments would suggest that long lasting adaptive change in sigma receptors could be involved, following long-term treatments with sigma antagonists.

Overall, many sigma ligands have demonstrated the ability to modulate NMDA-mediated glutamatergic neurotransmission. Since NMDA receptors may be involved in the mechanism of action of antidepressants and as an altered glutamatergic neurotransmission may underlie depressive pathology, the ability of sigma receptors to modulate NMDA-receptor-mediated responses may have important implications towards the potential of sigma ligands as antidepressants.

To test this hypothesis, we investigated the effects of treatments with sigma ligands on NMDA receptor-mediated behaviors at three weeks after an olfactory bulbectomy (OBX), an established animal model of depression (83, 184, 185), which induces a decrease in NMDA receptor ligand binding (186). Treatments with sigma ligands were able to reverse the effects of OBX surgery on NMDA receptor-mediated behaviors. The effects of the sigma ligands tested were dependent on the dose administered and on the duration of the treatments. The effects of the different doses were in keeping with the bell-shaped dose-response curve previously reported for these sigma ligands. Importantly, this behavioral study clearly suggests a potential for sigma ligands as antidepressants in a dose- and duration-dependent manner, as treatments with sigma ligand can reverse OBX-induced NMDA-mediated behaviors.

### 5.2.2. Sigma receptors and serotonergic neurotransmission

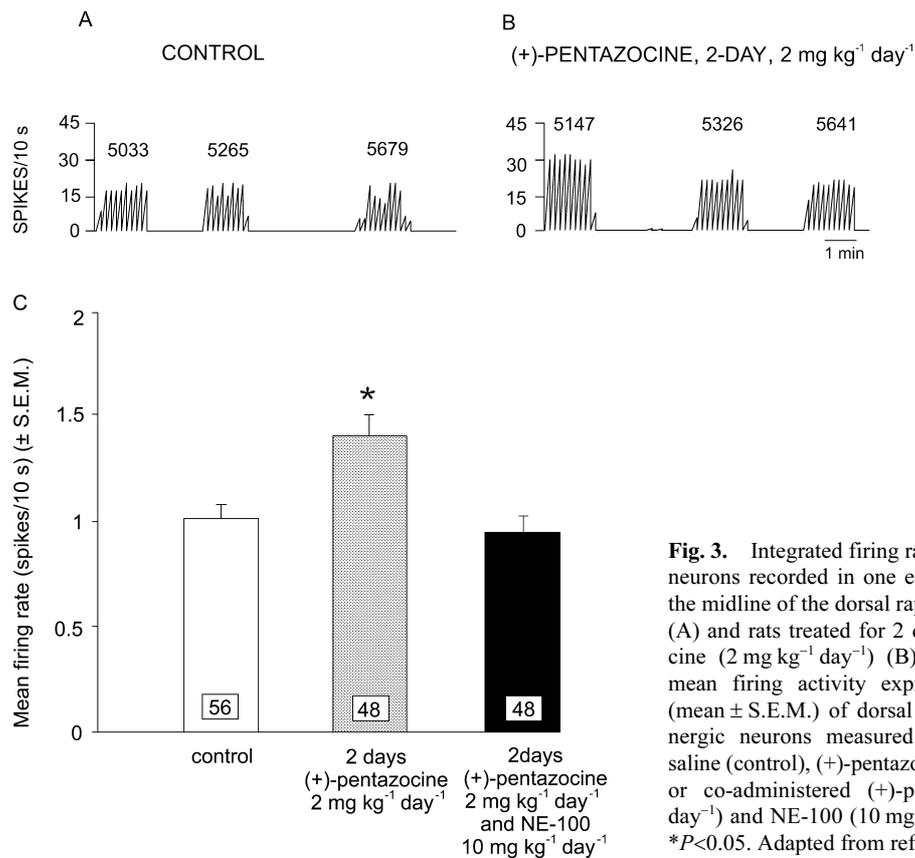
When examining potential novel antidepressants, one cannot exclude the involvement of the 5-HT system. As stated above, 5-HT plays a key role in depression and/or the mechanism of action of antidepressants. Thus, it begs the question, when examining sigma ligands' potential as antidepressants, whether they can modulate

5-HT neurotransmission.

Previous experiments had provided controversial evidence regarding possible interactions between sigma receptors and the 5-HT system. Peripheral 5-HT-sigma interactions have been proposed, as DTG, haloperidol, and BMY-14802 have been found to inhibit the 5-HT-evoked contractions of the guinea pig ileum longitudinal muscle/myenteric plexus preparation in a manner showing high correlation with their potency to compete with DTG binding (187). However, in a behavioral study, the sigma ligand EMD 57445 did not affect several 5-HT related parameters such as 8-OH-DPAT-induced behavioural syndrome, *m*-chlorophenylpiperazine-induced hypothermia or L-5-hydroxytryptophan-induced head twitches (188). In addition, biochemical studies showed that EMD 57445 and the sigma<sub>1</sub> ligand PD144418 did not induce any change in 5-HT or 5-hydroxyindoleacetic acid (5-HIAA) levels in various brain regions, suggesting that these ligands exert no effects on 5-HT-receptor populations or 5-HT metabolism (188, 189). However, EMD 57445 and PD 144415 have been suggested to be a sigma antagonists and therefore may have no effects on their own.

To investigate whether sigma ligands can modulate 5-HT neuronal activity *in vivo* we examined the effects of short- and long-term administration of various sigma ligands on 5-HT basal neuronal activity in the DRN, using extracellular recordings. As stated above, acute treatments with SSRIs and MAOIs induce a decrease in the firing activity of DRN 5-HT neurons (138, 139, 190). However, after long-term treatments, there is a restoration of the firing activity of these neurons (138, 139, 191, 192) due to the desensitization of the 5-HT<sub>1A</sub> auto-receptors (137, 141, 142). This paradigm of recordings from 5-HT neurons provided an ideal model to study sigma ligands' effect in a manner with which the results could be easily compared to those obtained with classical antidepressants.

In contrast with what has been observed in the dorsal hippocampus, the acute intravenous administration of (+)-pentazocine had no effect in the dorsal raphe. Interestingly, however, we found that the sigma ligands 4-IBP, (+)-pentazocine, and DTG after either 2 or 21 days of treatment induced a significant effect on the firing activity of 5-HT neurons of the DRN, which was increased by more than 50% (193) (Fig. 3). These findings suggest a clear modulation of 5-HT neurotransmission by sigma ligands *in vivo*, a novel finding with respect to sigma receptor research, supporting a role for sigma receptors in depression. They also suggested that sigma ligands modulate 5-HT neurotransmission by a mechanism different from those of classical antidepressants, as they induced an increase in



**Fig. 3.** Integrated firing rate histograms of 5-HT neurons recorded in one electrode descent along the midline of the dorsal raphe nucleus in controls (A) and rats treated for 2 days with (+)-pentazocine (2 mg kg<sup>-1</sup> day<sup>-1</sup>) (B). Panel C shows the mean firing activity expressed as spikes/10 s (mean ± S.E.M.) of dorsal raphe nucleus serotonergic neurons measured in rats treated with saline (control), (+)-pentazocine (2 mg kg<sup>-1</sup> day<sup>-1</sup>), or co-administered (+)-pentazocine (2 mg kg<sup>-1</sup> day<sup>-1</sup>) and NE-100 (10 mg kg<sup>-1</sup> day<sup>-1</sup>) for 2 days. \**P*<0.05. Adapted from ref. 193.

the firing activity of 5-HT neurons after only 2 days of treatment. If one accepts the hypothesis that the firing activity of 5-HT neurons represents a direct index of the efficacy of the 5-HT neurotransmission, this suggests that sigma ligands may represent antidepressants with a fast onset of action. Further studies were also conducted with the sigma<sub>1</sub> ligand OPC-14523, which also presents high affinity for the 5-HT<sub>1A</sub> receptor and moderate 5-HT reuptake blocking activity (156, 157). OPC-14523, similarly to other sigma ligands, induced a significant increase in the firing activity of 5-HT neurons of the DRN after 2 days of treatment. This increase in the firing activity was blocked by the coadministration of NE-100, suggesting that it was mediated by sigma<sub>1</sub> receptors. Interestingly, in addition, OPC-14523 after a 2-day treatment induced a decrease in the responsiveness of the 5-HT<sub>1A</sub> autoreceptor (194). This is particularly significant given that classical antidepressant medications require chronic treatment for this decreased autoreceptor response to occur (138, 141, 142). If present with other sigma<sub>1</sub> agonists, this rapid desensitization of the 5-HT<sub>1A</sub> autoreceptor, in addition to the observed rapid increase in the firing activity of 5-HT neurons after only 2 days of treatment, would constitute another argument to suggest that sigma agonists have potential

to produce a fast onset of antidepressant effect.

In keeping with the data obtained with sigma agonists, recent electrophysiological studies in the rat have shown that if progesterone does not have any effect by itself on 5-HT neuronal activity in the DRN, several of its metabolites such as alloprenanolone or other neurosteroids such as DHEA increase the firing activity of DRN 5-HT neurons, similarly to what is observed during pregnancy in correlation with the increase in progesterone level. Interestingly, at least part of these effects of neurosteroids are mediated through an activation of sigma receptors as they are reversed by NE-100 (195).

The precise mechanisms through which sigma ligands increase the firing activity of DRN 5-HT neurons is not established. One possibility is that this effect is mediated locally, in the DRN, as a consequence of the modulation of the glutamatergic neurotransmission as AMPA and NMDA receptors have been shown to mediate the glutamatergic excitatory input in the DRN (196). However, this effect could be also an indirect one; specifically, treatments with sigma ligands may rapidly modulate NMDA receptor-mediated transmission in the hippocampus, and potentially other forebrain regions, which in turn would lead to a modulation of 5-HT neurotransmission in the DRN via feedback loops to

DRN 5-HT neurons. Indeed, an afferent connection has been identified that projects from the hippocampus to the DRN via the lateral habenula (197–201).

Another feedback loop that may be involved is the “long feedback loop” that projects from the DRN to mPFC and back to the DRN (196, 199, 202–206). Therefore, the activity of the DRN neurons is dependent on the balance between the excitatory input from various brain regions (e.g., lateral habenula and mPFC) and inhibitory input from GABAergic interneurons in distal areas (e.g., periaqueductal gray area) and local GABAergic interneurons situated in the DRN (201, 207–209).

Such an indirect effect might explain the delayed onset of action of sigma ligands in the electrophysiological paradigm of DRN 5-HT neurons. It is possible that sustained treatments are leading to a potentiation of NMDA-receptor-mediated responses in the mPFC and/or other brain regions in addition to the hippocampus. This could cause an increase in the excitatory input from the mPFC to the DRN, which at some point may overcome the GABAergic inhibitory influences and shift the balance controlling the activity of the 5-HT neurons of the DRN, leading to an overall net increase in DRN 5-HT neuronal activity. This theory could explain why chronic treatments are needed for any alterations to be observed in the DRN, but not in the hippocampus.

Another factor likely contributing to the requirement of a sustained treatment to observe an effect is based on the density of sigma receptors at the plasma membrane, which is progressively altered by the presence of sigma ligands. This was shown in cell lines in which treatments with sigma agonists induced an increase in the sigma receptor density at the plasma membrane, following a minimum of 2 days of treatment (210). Thus, as treatments with sigma ligands continue and sigma receptors density increased in the plasma membrane, they can exert more significant effects on NMDA-receptor-mediated signalling. Based on this hypothesis, sigma receptors would also be increasing in concentration in DRN neuronal plasma membranes so that a stronger modulation of NMDA-receptor-mediated signalling occurs.

### 5.2.3. *Sigma receptors and Ca<sup>2+</sup> regulation*

Attempts to identify the mechanisms by which sigma ligands exert their “antidepressant-like” properties have also linked these aspects to sigma receptors’ regulation of Ca<sup>2+</sup>. The effects of JO-1784 in the FST were demonstrated to be Ca<sup>2+</sup>-dependent as the extracellular Ca<sup>2+</sup> chelator EGTA prevented the effect of JO-1784 in a dose-dependent manner. In addition, a lower dose of JO-1784 had no effect by itself, but co-administered

with the L-type voltage-dependent Ca<sup>2+</sup> channel (VDCC) positive modulator (–)-Bay K8644, it significantly reduced immobility. In agreement, the L-type VDCC antagonist verapamil and the N-type VDCC antagonist  $\omega$ -conotoxin blocked the effects of JO-1784 (211). Therefore, sigma<sub>1</sub> receptors may be interacting with pre- or postsynaptic VDCC’s to exert “antidepressant-like” effects in the FST (211).

In addition, bradykinin, which increases IP<sub>3</sub> levels, enhanced the effect of JO-1784 (211), whereas the IP<sub>3</sub>-receptor antagonist xestospongine C blocked the effect of JO-1784. Thus the mobilization of intracellular Ca<sup>2+</sup> from IP<sub>3</sub> receptor-sensitive pools appears also to participate initially in the behavioral effects mediated by sigma<sub>1</sub> receptors located on the ER membranes (211). The sigma<sub>1</sub> receptor then putatively moves to the plasma membrane and interacts with VDCC’s (31, 60).

It is likely that sigma ligands’ ability to modulate both glutamatergic and 5-HT transmissions contribute to the “antidepressant-like” effects observed in behavioral models. The molecular mechanism underlying sigma receptors’ ability to modulate 5-HT and glutamatergic transmissions may involve sigma receptors’ ability to modulate Ca<sup>2+</sup> (31, 58, 60, 212) or K<sup>+</sup> signalling (43, 50, 54, 213). This modulation could represent a downstream target involved in the effects of sigma ligands on both the glutamatergic and the 5-HT systems, thus leading to one primary target. The effect of antidepressants on downstream targets has been a recent and important shift in antidepressant research. As reviewed by recent literature, downstream targets are the focus for a mechanism of action for antidepressants and downstream adaptive changes could explain the time lag seen between changes in neurotransmitter systems and therapeutic effects seen in patients (reviewed by refs. 106 and 214–216). However, besides Ca<sup>2+</sup> regulations, one downstream target of particular interest for antidepressant treatments is represented by neurotrophic factors (reviewed by refs. 106, 107, and 214–216).

### 5.2.4. *Sigma receptors and neurogenesis*

Recent evidence has shown an hippocampal atrophy in major depression, which can persist long after the pathology is resolved (217–220). This atrophy could be due to a regression of dendritic processes, an inhibition of neurogeneration or the loss of hippocampal neurons [reviewed by Sapolsky (221)]. It has also been shown that hippocampal atrophy can be reversed by successful antidepressant treatments and that in vitro, classical antidepressants promote neurogenesis (222). A recent study has provided a role for sigma<sub>1</sub> receptors in cell morphological changes, specifically in the initiation of neurite outgrowth and sprouting (210, 223). Sigma<sub>1</sub> receptors

and ankyrins are highly concentrated in the growth cone of NG-108 cells, a region related to neurite sprouting, extension, and guidance (223). The sigma<sub>1</sub> agonist (+)-pentazocine had no effect by itself on neurite sprouting, but potentiated the neurite-sprouting induced by nerve growth factor (NGF) (210). In contrast, neurite sprouting induced by cAMP in PC12 cells was not affected by (+)-pentazocine. The sigma<sub>1</sub> antagonist NE-100, regardless of the presence of NGF, did not affect neurite sprouting, but antagonized the potentiation induced by (+)-pentazocine, thus clearly indicating a mediation via sigma<sub>1</sub> receptors (210).

Interestingly, similar to sigma agonists, the antidepressants imipramine and fluvoxamine potentiated the effects of NGF in this model (210). These effects of imipramine and fluvoxamine were antagonized by NE-100, while no concentration of 5-HT tested affected neurite sprouting induced by NGF (210), therefore suggesting that the effect on NGF-induced neurite outgrowth of both sigma<sub>1</sub> agonists and classical antidepressants are mediated by sigma<sub>1</sub> receptors. Moreover, cell treatments with NGF, even in the absence of sigma<sub>1</sub>-receptor agonists, increased the level of sigma<sub>1</sub> receptors in a dose-dependent manner, and the effects of (+)-pentazocine and NGF were additive (210). Interestingly, treatment with imipramine and fluvoxamine also increased sigma<sub>1</sub> receptors. Furthermore, in MT40 cells expressing higher levels of sigma<sub>1</sub> receptors, NGF was found to be more potent in inducing neurite sprouting, whereas treatments with sigma<sub>1</sub>-receptor antisense significantly reduced the degree of neurite sprouting (210). Together, these data suggest a primary role for sigma<sub>1</sub> ligands in enhancing NGF-induced neurite growth.

One member of the neurotrophin family, BDNF, has been heavily implicated in the action of antidepressants, as chronic treatments with a variety of antidepressant therapies induce an increase in BDNF expression (224–226). Moreover, BDNF administration itself has been shown to produce antidepressant effects in behavioural models of depression (227, 228).

It would be interesting to know the effects of sigma ligands on BDNF. Thus far, this has only been studied with the sigma ligand E-5842, which showed no effects on BDNF or NGF levels following chronic treatments (229). However, again, E-5842 presents a sigma antagonist profile, so its lack of efficacy cannot be considered as an indication of what might be the effects generated by sigma agonists. It is still conceivable that sigma agonist would potentiate the effects of BDNF, similarly to that observed previously with NGF.

Another growth factor is the epidermal growth factor (EGF). EGF is present in the CNS and known to stimu-

late cell proliferation. A recent report from the group of Su indicated that in PC<sub>12</sub> cells, the overexpression of sigma<sub>1</sub> receptors induced a 3-fold increase of neurite sprouting, which was suppressed by the sigma<sub>1</sub> antagonist NE-100 (230). In the context of this review, these data are even more interesting if one considers that EGF has been shown to enhance NMDA-induced modulation of intracellular Ca<sup>2+</sup> (231).

More research will be required to elucidate the exact basis for the observed potentiation of neurotrophin effects by sigma agonists and whether sigma ligands always affect neuronal survival and neurogenesis. From there, it would be interesting to see whether these effects are necessary for the observed modulatory effects of sigma receptors on many neurotransmitter systems. This will provide many new avenues to explore the field of sigma receptors and offer valuable information about the clinical potential of these ligands.

## 6. Conclusions

The glutamatergic and serotonergic systems are heavily implicated in antidepressant action. Sigma<sub>1</sub> ligands have demonstrated “antidepressant-like” effects in animal models screening for antidepressant action. Moreover, sigma<sub>1</sub> ligands have been shown to modulate glutamatergic and serotonergic neuronal activity in *in vivo* electrophysiological paradigms. These actions of sigma<sub>1</sub> ligands suggest that sigma<sub>1</sub> ligands may have potential as novel antidepressant agents. Furthermore, the data also suggests that sigma<sub>1</sub> ligands may present a novel mechanism of antidepressant action with potential for a faster onset of action versus classical antidepressants. Therefore, the data presented in the current Review strongly support the further investigation into sigma<sub>1</sub> ligands as potential antidepressants.

## References

- 1 Martin WR, Eades CG, Thompson JA, Huppler RE, Gilbert PE. The effects of morphine- and nalorphine-like drugs in the nondependent and morphine-dependent chronic spinal dog. *J Pharmacol Exp Ther.* 1976;197:517–532.
- 2 Quirion R, Chichportiche R, Contreras P, Johnson K, Lodge D, Tam S, et al. Classification and nomenclature of phencyclidine and sigma receptor sites. *Trends Neurol Sci.* 1987;10: 444–446.
- 3 Su TP. Evidence for sigma opioid receptor: binding of [<sup>3</sup>H]SKF-10047 to etorphineinaccessible sites in guinea-pig brain. *J Pharmacol Exp Ther.* 1982;223:284–290.
- 4 Steinfels GF, Alberici GP, Tam SW, Cook L. Biochemical, behavioral, and electrophysiologic actions of the selective sigma receptor ligand (+)pentazocine. *Neuropsychopharmacology.* 1988;1:321–327.
- 5 Vilner BJ, John CS, Bowen WD. Sigma-1 and sigma-2 receptors are present in a wide variety of human and rodent cell lines.

- Cancer Research. 1995;88:408–413.
- 6 Basile AS, Paul IA, Mirchevitch A, Kurjpers G, De Costa B. Modulation of (+)-(3H) pentazocine to guinea pig cerebellum by divalent cations. *Mol Pharmacol*. 1992;42:882–889.
  - 7 Roman FJ, Pascaud X, Martin B, Vauche D, Junien JL. JO-1784, a potent and selective ligand for rat and mouse brain sigma sites. *J Pharm Pharmacol*. 1990;42:439–440.
  - 8 Middlemiss DN, et al. L-687,384 is a potent, selective ligand at the central sigma recognition site. *Br J Pharmacol*. 1991;102:153.
  - 9 Mcmillan DE, Hardwick WC, Decosta BR, Rice KC. Effects of drugs that bind to PCP and sigma receptors on punished responding. *J Pharmacol Exp Ther*. 1991;258:1015–1018.
  - 10 Tsao LI, Su TP. Naloxone-sensitive, haloperidol-sensitive [<sup>3</sup>H](+)-SKF-10047-binding protein partially purified from rat liver and rat brain membranes: An opioid/sigma receptor? *Synapse*. 1997;25:117–124.
  - 11 Weber E, Sonders M, Quarum M, McLean S, Pou S, Keana JF. 1,3-Di(2-[5-3H]tolyl)guanidine: a selective ligand that labels sigma-type receptors for psychotomimetic opiates and anti-psychotic drugs. *Proc Natl Acad Sci USA*. 1986;83:8784–8788.
  - 12 Clark D, Engberg G, Pileblad E, Svensson TH, Carlsson A, Freeman AS, et al. An electrophysiological analysis of the action of the 3PPP enantiomers on the nigrostriatal dopamine system. *Naunyn Schmiedebergs Arch Pharmacol*. 1985;329:344–354.
  - 13 Su TP, London ED, Jaffe JH. Steroid binding at sigma receptors suggests a link between endocrine, nervous, and immune systems. *Science*. 1988;240:219–221.
  - 14 Gundlach AL, Largent BL, Snyder SH. Autoradiographic localization of sigma receptor binding sites in guinea pig and rat central nervous system with (+)[<sup>3</sup>H]-3-(3-hydroxyphenyl)-N-(1-propyl)piperidine. *J Neurosci*. 1986;6:1757–1770.
  - 15 Bouchard P, Quirion R. [<sup>3</sup>H]1,3-Di(2-tolyl)guanidine and [<sup>3</sup>H](+)-pentazocine binding sites in the rat brain: autoradiographic visualization of the putative sigma<sub>1</sub> and sigma<sub>2</sub> receptor subtypes. *Neuroscience*. 1997;76:467–477.
  - 16 Inoue A, Sugita S, Shoji H, Ichimoto H, Hide I, Nakata Y. Repeated haloperidol treatment decreases sigma(1) receptor binding but does not affect its mRNA levels in the guinea pig or rat brain. *Eur J Pharmacol*. 2000;401:307–316.
  - 17 Alonso G, Phan V, Guillemain I, Saunier M, Legrand A, Anoaï M, et al. Immunocytochemical localization of the sigma<sub>1</sub> receptor in the adult rat central nervous system. *Neuroscience*. 2000;97:155–170.
  - 18 Mei J, Pan Y-X, Xiu J, King M, Chang P, Pasternak GW, et al. Cloning and functional characterization of the mouse sigma<sub>1</sub> receptor [abstract]. *Soc Neurosci*. 1997;23:2318.
  - 19 Soby KK, Meier F, Mikkelsen JD. Autoradiographic distribution of sigma<sub>2</sub> binding in the rat brain identified with a new receptor ligand [<sup>3</sup>H]Lu 28-179 [abstract]. *Soc Neurosci*. 1997;23:588.20.
  - 20 Yamada M, Nishigami T, Nakasho K, Nishimoto Y, Miyaji H. Relationship between sigma-like site and progesterone-binding site of adult male rat liver microsomes. *Hepatology*. 1994;20:1271–1280.
  - 21 Ramamoorthy JD, Ramamoorthy S, Mahesh VB, Leibach FH, Ganapathy V. Cocainesensitive sigma-receptor and its interaction with steroid hormones in the human placental syncytiotrophoblast and in choriocarcinoma cells. *Endocrinology*. 1995;136:924–932.
  - 22 Maurice T, Roman FJ, Privat A. Modulation by neurosteroids of the in vivo (+)-[<sup>3</sup>H]SKF-10,047 binding to sigma<sub>1</sub> receptors in the mouse forebrain. *J Neurosci Res*. 1996;46:734–743.
  - 23 Schwarz S, Pohl P, Zhou GZ. Steroid binding at sigma-opioid receptors. *Science*. 1989;246:1635–1637.
  - 24 Debonnel G, Bergeron R, de Montigny C. Potentiation by dehydroepiandrosterone of the neuronal response to N-methyl-D-aspartate in the CA<sub>3</sub> region of the rat dorsal hippocampus: an effect mediated via sigma receptors. *J Endocrinol*. 1996;150:S33–S42.
  - 25 Bergeron R, de Montigny C, Debonnel G. Pregnancy reduces brain sigma receptor function. *Br J Pharmacol*. 1999;127:1769–1776.
  - 26 Urani A, Roman FJ, Phan VL, Su TP, Maurice T. The antidepressant-like effect induced by sigma(1)-receptor agonists and neuroactive steroids in mice submitted to the forced swimming test. *J Pharmacol Exp Ther*. 2001;298:1269–1279.
  - 27 Maurice T, Phan V-L, Su T-P. Modulations of steroidal levels by adrenalectomy/castration and inhibition of neurosteroid synthesis enzymes affect the sigma<sub>1</sub> receptor-mediated behaviors in mice [abstract]. *Soc Neurosci*. 1998;24:1353.
  - 28 Maurice T, Hiramatsu M, Itoh J, Kameyama T, Hasegawa T, Nabeshima T. Behavioral evidence for a modulating role of sigma ligands in memory processes. I. Attenuation of dizocilpine (MK-801)-induced amnesia. *Brain Res*. 1994;647:44–56.
  - 29 Bergeron R, de Montigny C, Debonnel G. Biphasic effects of sigma ligands on the neuronal response to N-methyl-D-aspartate. *Naunyn Schmiedebergs Arch Pharmacol*. 1995;351:252–260.
  - 30 Monnet FP, De Costa BR, Bowen WD. Differentiation of sigma ligand-activated receptor subtypes that modulate NMDA-evoked [<sup>3</sup>H]-noradrenaline release in rat hippocampal slices. *Br J Pharmacol*. 1996;119:65–72.
  - 31 Hayashi T, Maurice T, Su TP. Ca(2+) signaling via sigma(1)-receptors: novel regulatory mechanism affecting intracellular Ca(2+) concentration. *J Pharmacol Exp Ther*. 2000;293:788–798.
  - 32 Monnet FP, Debonnel G, Junien JL, de Montigny C. N-methyl-D-aspartate-induced neuronal activation is selectively modulated by sigma receptors. *Eur J Pharmacol*. 1990;179:441–445.
  - 33 Monnet FP, Debonnel G, de Montigny C. In vivo electrophysiological evidence for a selective modulation of N-methyl-D-aspartate-induced neuronal activation in rat CA<sub>3</sub> dorsal hippocampus by sigma ligands. *J Pharmacol Exp Ther*. 1992;261:123–130.
  - 34 Maurice T, Hiramatsu M, Kameyama T, Hasegawa T, Nabeshima T. Behavioral evidence for a modulating role of sigma ligands in memory processes. II. Reversion of carbon monoxide-induced amnesia. *Brain Res*. 1994;647:57–64.
  - 35 Bergeron R, Debonnel G, de Montigny C. Modification of the N-methyl-D-aspartate response by antidepressant sigma receptor ligands. *Eur J Pharmacol*. 1993;240:319–323.
  - 36 Bergeron R, Debonnel G. Effects of low and high doses of selective sigma ligands: Further evidence suggesting the existence of different subtypes of sigma receptors. *Psychopharmacology*. 1997;129:215–224.
  - 37 Hanner M, Moebius FF, Flandorfer A, Knaus HG, Striessing J, Kempner E, et al. Purification, molecular cloning, and expression of the mammalian sigma<sub>1</sub>-binding site. *Proc Natl Acad Sci USA*. 1996;93:8072–8077.
  - 38 Prasad PD, Li HW, Fei YJ, Ganapathy ME, Fujita T, Plumley LH, et al. Exon-intron structure, analysis of promoter region, and

- chromosomal localization of the human type 1  $\sigma$  receptor gene. *J Neurochem.* 1998;70:443–451.
- 39 Pan Y-X, Mei J, Xu J, Wan BL, Zuckerman A, Pasternak GW. Cloning and characterization of a mouse  $\sigma_1$  receptor. *J Neurochem.* 1998;70:2279–2285.
- 40 Seth P, Leibach F, Ganapathy V. Cloning and structural analysis of the cDNA and the gene encoding the murine type 1 sigma receptor. *Biochem Biophys Res Commun.* 1997;241:535–540.
- 41 Seth P, Fei YJ, Li HW, Huang W, Leibach FH, Ganapathy V. Cloning and functional characterization of a  $\sigma$  receptor from rat brain. *J Neurochem.* 1998;70:922–931.
- 42 Kekuda R, Prasad PD, Fei YJ, Leibach FH, Ganapathy V. Cloning and functional expression of the human type 1 sigma receptor (hSigmaR1). *Biochem Biophys Res Commun.* 1996;229:553–558.
- 43 Aydar E, Palmer CP, Klyachko VA, Jackson MB. The sigma receptor as a ligand-regulated auxiliary potassium channel subunit. *Neuron.* 2002;34:399–410.
- 44 Langa F, Codony X, Tovar V, Lavado A, Gimenez E, Cozar P, et al. Generation and phenotypic analysis of sigma receptor type I (sigma 1) knockout mice. *Eur J Neurosci.* 2003;18:2188–2196.
- 45 Beart PM, Oshea RD, Manallack DT. Regulation of sigma-receptors – high-affinity and low-affinity agonist states, GTP shifts, and up-regulation by rimcazole and 1,3-di(2-tolyl)guanidine. *J Neurochem.* 1989;53:779–788.
- 46 Connick JH, Hanlon G, Roberts J, France L, Fox PK, Nicholson CD. Multiple  $\sigma$  binding sites in guinea-pig and rat brain membranes: G-protein interactions. *Br J Pharmacol.* 1992;107:726–731.
- 47 Itzhak Y. Different modulation of the binding to 2 phencyclidine (PCP) receptor subtypes – Effects of N-methyl-D-aspartate agonists and antagonists. *Neurosci Lett.* 1989;104:314–319.
- 48 Itzhak Y, Stein I. Regulation of  $\sigma$  receptors and responsiveness to guanine nucleotides following repeated exposure of rats to haloperidol: Further evidence for multiple  $\sigma$  binding sites. *Brain Res.* 1991;566:166–172.
- 49 Tokuyama S, Hirata K, Ide A, Ueda H. Sigma ligands stimulate GTPase activity in mouse prefrontal membranes: evidence for the existence of metabotropic sigma receptor. *Neurosci Lett.* 1997;233:141–144.
- 50 Soriani O, Vaudry H, Mei Y, Roman F, Cazin L. Sigma ligands stimulate the electrical activity of frog pituitary melanotrope cells through a G-protein-dependent inhibition of potassium conductances. *J Pharmacol Exp Ther.* 1998;286:163–171.
- 51 Monnet FP, Blier P, Debonnel G, de Montigny C. Modulation by sigma ligands of N-methyl-D-aspartate-induced [ $^3$ H]noradrenaline release in the rat hippocampus: G-protein dependency. *Naunyn Schmiedeberg Arch Pharmacol.* 1992;346:32–39.
- 52 Monnet FP, Debonnel G, Bergeron R, Gronier B, de Montigny C. The effects of sigma ligands and of neuropeptide Y on N-methyl-D-aspartate-induced neuronal activation of CA3 dorsal hippocampus neurones are differentially affected by pertussis toxin. *Br J Pharmacol.* 1994;112:709–715.
- 53 Wilke RA, Mehta RP, Lupardus PJ, Chen Y, Ruoho AE, Jackson MB, et al. Sigma receptor photolabeling and sigma receptor-mediated modulation of potassium channels in tumor cells. *J Biol Chem.* 1999;274:18387–18392.
- 54 Lupardus PJ, Wilke RA, Aydar E, Palmer CP, Chen Y, Ruoho AE, et al. Membrane-delimited coupling between sigma receptors and  $K^+$  channels in rat neurohypophysial terminals requires neither G-protein nor ATP. *J Physiol (Lond).* 2000;526 Pt 3:527–539.
- 55 McCann DJ, Weissman AD, Su TP. Sigma-1 and sigma-2 sites in rat brain: comparison of regional, ontogenetic, and subcellular patterns. *Synapse.* 1994;17:182–189.
- 56 DeHaven-Hudkins D, Fleissner L, Ford-Rice F. Characterization of the binding of [ $^3$ H](+)-pentazocine to  $\sigma$  recognition sites in guinea pig brain. *Eur J Pharmacol.* 1992;227:371–378.
- 57 Hong W, Werling LL. Evidence that the sigma(1) receptor is not directly coupled to G proteins. *Eur J Pharmacol.* 2000;408:117–125.
- 58 Hayashi T, Su TP. Regulating ankyrin dynamics: roles of sigma-1 receptors. *Proc Natl Acad Sci USA.* 2001;98:491–496.
- 59 Brent PJ, Saunders H, Dunkley PR. Intrasyaptosomal free calcium levels in rat forebrain synaptosomes : modulation by sigma ( $\sigma$ ) receptor ligands. *Neurosci Lett.* 1996;211:1–5.
- 60 Brent PJ, Herd L, Saunders H, Sim ATR, Dunkley PR. Protein phosphorylation and calcium uptake into rat forebrain synaptosomes: modulation by the  $\sigma$  ligand, 1,3-ditolylguanidine. *J Neurochem.* 1997;68:2201–2211.
- 61 Soriani O, Le Foll F, Galas L, Roman F, Vaudry H, Cazin L. The sigma-ligand (+)-pentazocine depresses M current and enhances calcium conductances in frog melanotrophs. *Am J Physiol.* 1999;277:E73–E80.
- 62 Okuyama S, Imagawa Y, Ogawa S, Araki H, Aijima A, Tanaka M, et al. NE-100, a novel sigma receptor ligand: In vivo tests. *Life Sci.* 1993;53:PL285–PL290.
- 63 Bleich S, Romer K, Wiltfang J, Kornhuber J. Glutamate and the glutamate receptor system: a target for drug action. *Int J Geriatr Psychiatry.* 2003;18:S33–S40.
- 64 Seeburg PH. The TINS/TiPS lecture. The molecular biology of mammalian glutamate receptor channels. *Trends Neurosci.* 1993;16:359–365.
- 65 Sanacora G, Rothman DL, Mason G, Krystal JH. Clinical studies implementing glutamate neurotransmission in mood disorders. *Ann NY Acad Sci.* 2003;1003:292–308.
- 66 Kim JS, Schmid-Burgk W, Claus D, Kornhuber HH. Increased serum glutamate in depressed patients. *Arch Psychiatr Nervenkr.* 1982;232:299–304.
- 67 Altamura CA, Mauri MC, Ferrara A, Moro AR, D’Andrea G, Zamberlan F. Plasma and platelet excitatory amino acids in psychiatric disorders. *Am J Psychiatry.* 1993;150:1731–1733.
- 68 Altamura C, Maes M, Dai J, Meltzer HY. Plasma concentrations of excitatory amino acids, serine, glycine, taurine and histidine in major depression. *Eur Neuropsychopharmacol.* 1995;5 Suppl: 71–75.
- 69 Levine J, Panchalingam K, Rapoport A, Gershon S, McClure RJ, Pettegrew JW. Increased cerebrospinal fluid glutamine levels in depressed patients. *Biol Psychiatry.* 2000;47:586–593.
- 70 Berk M, Plein H, Ferreira D. Platelet glutamate receptor supersensitivity in major depressive disorder. *Clin Neuropharmacol.* 2001;24:129–132.
- 71 Mauri MC, Ferrara A, Boscati L, Bravin S, Zamberlan F, Alecci M, et al. Plasma and platelet amino acid concentrations in patients affected by major depression and under fluvoxamine treatment. *Neuropsychobiology.* 1998;37:124–129.
- 72 Maes M, Verkerk R, Vandoolaeghe E, Lin A, Scharpe S. Serum levels of excitatory amino acids, serine, glycine, histidine, threonine, taurine, alanine and arginine in treatment-resistant depression: modulation by treatment with antidepressants and

- prediction of clinical responsivity. *Acta Psychiatr Scand.* 1998;97:302–308.
- 73 Auer DP, Putz B, Kraft E, Lipinski B, Schill J, Holsboer F. Reduced glutamate in the anterior cingulate cortex in depression: an in vivo proton magnetic resonance spectroscopy study. *Biol Psychiatry.* 2000;47:305–313.
  - 74 Michael N, Erfruth A, Ohrmann P, Arolt V, Heindel W, Pfeleiderer B. Metabolic changes within the left dorsolateral prefrontal cortex occurring with electroconvulsive therapy in patients with treatment resistant unipolar depression. *Psychol Med.* 2003;33:1277–1284.
  - 75 Pfeleiderer B, Michael N, Erfruth A, Ohrmann P, Hohmann P, Wolgast M, et al. Effective electroconvulsive therapy reverses glutamate/glutamine deficit in the left anterior cingulum of unipolar depressed patients. *Psychiatry Res.* 2003;122:185–192.
  - 76 Trullas R, Skolnick P. Functional antagonists at the NMDA receptor complex exhibit antidepressant actions. *Eur J Pharmacol.* 1990;185:1–10.
  - 77 Petrie R, Reid I, Stewart C. The N-methyl-D-aspartate receptor, synaptic plasticity, and depressive disorder A critical review. *Pharmacol Ther.* 2000;87:11–25.
  - 78 Trullas R, Jackson B, Skolnick P. Anxiolytic properties of 1-aminocyclopropanecarboxylic acid, a ligand at strychnine-insensitive glycine receptors. *Pharmacol Biochem Behav.* 1989;34:313–316.
  - 79 Layer RT, Popik P, Olds T, Skolnick P. Antidepressant-like actions of the polyamine site NMDA antagonist, eliprodil (SL-82.0715). *Pharmacol Biochem Behav.* 1995;52:621–627.
  - 80 Yilmaz A, Schulz D, Aksoy A, Canbeyli R. Prolonged effect of an anesthetic dose of ketamine on behavioral despair. *Pharmacol Biochem Behav.* 2002;71:341–344.
  - 81 Padovan CM, Guimaraes FS. Antidepressant-like effects of NMDA-receptor antagonist injected into the dorsal hippocampus of rats. *Pharmacol Biochem Behav.* 2004;77:15–19.
  - 82 Mjelle N, Lund A, Hole K. Reduction of NMDA-induced behaviour after acute and chronic administration of desipramine in mice. *Neuropharmacology.* 1993;32:591–595.
  - 83 Kelly JP, Wrynn AS, Leonard BE. The olfactory bulbectomized rat as a model of depression: an update. *Pharmacol Ther.* 1997;74:299–316.
  - 84 Maj J, Rogoz Z, Skuza G. Antidepressant drugs increase the locomotor hyperactivity induced by MK-801 in rats. *J Neural Transm.* 1991;85:169–179.
  - 85 Maj J, Rogoz Z, Skuza G, Sowinska H. The effect of antidepressant drugs on the locomotor hyperactivity induced by MK-801, a non-competitive NMDA receptor antagonist. *Neuropharmacology.* 1992;31:685–691.
  - 86 Paul AP, Layer RT, Skolnick P, Nowak G. Adaptation of the NMDA receptor in rat cortex following chronic electroconvulsive shock or imipramine. *Eur J Pharmacol.* 1993;247:305–311.
  - 87 Paul IA, Nowak G, Layer RT, Popik P, Skolnick P. Adaptation of the N-methyl-D-Aspartate receptor complex following chronic antidepressant treatments. *J Pharmacol Exp Ther.* 1994;269:95–102.
  - 88 Nowak G, Paul I, Popik P, Young A, Skolnick P. Ca<sup>2+</sup> antagonists effect an antidepressantlike adaptation of the NMDA receptor complex. *Eur J Pharmacol.* 1993;247:101–102.
  - 89 Nowak G, Legutko B, Skolnick P, Popik P. Adaptation of cortical NMDA receptors by chronic treatment with specific serotonin reuptake inhibitors. *Eur J Pharmacol.* 1998;342:367–370.
  - 90 Pilc A, Legutko B. The influence of prolonged antidepressant treatment on the changes in cyclic AMP accumulation induced by excitatory amino acids in rat cerebral cortical slices. *NeuroReport.* 1995;7:85–88.
  - 91 Skolnick P, Layer RT, Popik P, Nowak G, Paul IA, Trullas R. Adaptation of N-methyl-D-aspartate (NMDA) receptors following antidepressant treatment: implications for the pharmacotherapy of depression. *Pharmacopsychiatry.* 1996;29:23–26.
  - 92 Boyer PA, Skolnick P, Fossum LH. Chronic administration of imipramine and citalopram alters the expression of NMDA receptor subunit mRNAs in mouse brain. A quantitative in situ hybridization study. *J Mol Neurosci.* 1998;10:219–233.
  - 93 Watkins CJ, Pei Q, Newberry NR. Differential effects of electroconvulsive shock on the glutamate receptor mRNAs for NR2A, NR2B and mGluR5b. *Brain Res Mol Brain Res.* 1998;61:108–113.
  - 94 Pallotta M, Segieth J, Sadideen F, Whitton PS. Repeated but not acute clomipramine decreases the effect of N-methyl-D-aspartate receptor activation on serotonergic transmission between the raphe nuclei and frontal cortex. *Neuropharmacology.* 2001;41:294–300.
  - 95 De La Garza R, Jentsch JD, Verrico CD, Roth RH. Adaptation of monoaminergic responses to phencyclidine in nucleus accumbens and prefrontal cortex following repeated treatment with fluoxetine or imipramine. *Brain Res.* 2002;958:20–27.
  - 96 Bobula B, Tokarski K, Hess G. Repeated administration of antidepressants decreases field potentials in rat frontal cortex. *Neuroscience.* 2003;120:765–769.
  - 97 Nowak G, Ordway G, Paul I. Alterations in the N-methyl-D-aspartate (NMDA) receptor complex in the frontal cortex of suicide victims. *Brain Res.* 1995;675:157–164.
  - 98 Law AJ, Deakin JF. Asymmetrical reductions of hippocampal NMDAR1 glutamate receptor mRNA in the psychoses. *Neuro Report.* 2001;12:2971–2974.
  - 99 Moryl E, Danysz W, Quack G. Potential antidepressive properties of amantadine, memantine and bifemelane. *Pharmacol Toxicol.* 1993;72:394–397.
  - 100 Huber MT, Gotthardt U, Schreiber W, Krieg JC. Efficacy and safety of the sigma receptor ligand EMD 57445 (panamesine) in patients with schizophrenia: an open clinical trial. *Pharmacopsychiatry.* 1999;32:68–72.
  - 101 Kudoh A, Takahira Y, Katagai H, Takazawa T. Small-dose ketamine improves the postoperative state of depressed patients. *Anesth Analg.* 2002;95:114–118.
  - 102 Stryker R, Strous RD, Shaked G, Bar F, Feldman B, Kotler M, et al. Amantadine as augmentation therapy in the management of treatment-resistant depression. *Int Clin Psychopharmacol.* 2003;18:93–96.
  - 103 Li X, Witkin JM, Need AB, Skolnick P. Enhancement of antidepressant potency by a potentiator of AMPA receptors. *Cell Mol Neurobiol.* 2003;23:419–430.
  - 104 Lauterborn JC, Lynch G, Vanderklish P, Arai A, Gall CM. Positive modulation of AMPA receptors increases neurotrophin expression by hippocampal and cortical neurons. *J Neurosci.* 2000;20:8–21.
  - 105 Altar CA. Neurotrophins and depression. *Trends Pharmacol Sci.* 1999;20:59–61.
  - 106 Duman R, Heninger G, Nestler E. A molecular and cellular theory of depression. *Arch Gen Psychiatry.* 1997;54:597–606.

- 107 Coyle JT, Duman RS. Finding the intracellular signaling pathways affected by mood disorder treatments. *Neuron*. 2003; 38:157–160.
- 108 Martinez-Turrillas R, Frechilla D, Del Rio J. Chronic antidepressant treatment increases the membrane expression of AMPA receptors in rat hippocampus. *Neuropharmacology*. 2002;43: 1230–1237.
- 109 Svenningsson P, Tzavara ET, Witkin JM, Fienberg AA, Nomikos GG, Greengard P. Involvement of striatal and extra-striatal DARPP-32 in biochemical and behavioral effects of fluoxetine (Prozac). *Proc Natl Acad Sci USA*. 2002;99:3182–3187.
- 110 Southam E, Garthwaite J. The nitric oxide-cyclic GMP signaling pathway in rat brain. *Neuropharmacology*. 1993;32:1267–1277.
- 111 Harvey BH. Affective disorders and nitric oxide: a role in pathways to relapse and refractoriness? *Human Psychopharmacol*. 1996;11:309–319.
- 112 van Amsterdam JG, Opperhuizen A. Nitric oxide and bipterin in depression and stress. *Psychiatry Res*. 1999;85:33–38.
- 113 Finkel MS, Laghrissi-Thode F, Pollock BG, Rong J. Paroxetine is a novel nitric oxide synthase inhibitor. *Psychopharmacol Bull*. 1996;32:653–658.
- 114 Wegener G, Volke V, Harvey B, Rosenberg R. Local, but not systematic, administration of serotonergic antidepressants decreases hippocampal nitric oxide synthase activity. *Brain Res*. 2003;959:128–134.
- 115 Jefferys D, Funder J. Nitric oxide modulates retention of immobility in the forced swimming test in rats. *Eur J Pharmacol*. 1996;295:131–135.
- 116 Harkin AJ, Bruce KH, Craft B, Paul IA. Nitric oxide synthase inhibitors have antidepressant-like properties in mice. 1. Acute treatments are active in the forced swim test. *Eur J Pharmacol*. 1999;372:207–213.
- 117 Karolewicz B, Bruce KH, Lee B, Paul IA. Nitric oxide synthase inhibitors have antidepressant-like properties in mice. 2. Chronic treatment results in downregulation of cortical beta-adrenoceptors. *Eur J Pharmacol*. 1999;372:215–220.
- 118 Yildiz F, Erden BF, Ulak G, Utkan T, Gacar N. Antidepressant-like effect of 7-nitroindazole in the forced swimming test in rats. *Psychopharmacology (Berl)*. 2000;149:41–44.
- 119 Barnes NM, Sharp T. A review of central 5-HT receptors and their function. *Neuropharmacology*. 1999;38:1083–1152.
- 120 Aghajanian GK, Wang RY, Baraban J. Serotonergic and non-serotonergic neurons of the dorsal raphe: reciprocal changes in firing induced by peripheral nerve stimulation. *Brain Res*. 1978;153:169–175.
- 121 Vandermaelen C, Aghajanian G. Electrophysiological and pharmacological characterization of serotonergic dorsal raphe neurons recorded extracellularly and intracellularly in rat brain slices. *Brain Res*. 1983;289:109–119.
- 122 Marcinkiewicz M, Verge D, Gozlan H, Pichat L, Hamon M. Autoradiographic evidence for the heterogeneity of 5-HT<sub>1</sub> sites in the rat brain. *Brain Res*. 1984;291:159–163.
- 123 Kohler C. The distribution of serotonin binding sites in the hippocampal region of the rat brain. An autoradiographic study. *Neuroscience*. 1984;13:667–680.
- 124 Pazos A, Palacios JM. Quantitative autoradiographic mapping of serotonin receptors in the rat brain. I. Serotonin-1 receptors. *Brain Res*. 1985;346:205–230.
- 125 Kia H, Miquel MC, Brisorgueil MJ, Daval G, Riad M, El Mestikawy S, et al. Immunocytochemical localization of serotonin<sub>1A</sub> receptors in the rat central nervous system. *J Comp Neurol*. 1996;365:289–305.
- 126 Andrade R, Malenka RC, Nicoll RA. A G protein couples serotonin and GABA<sub>B</sub> receptors to the same channels in hippocampus. *Science*. 1986;234:1261–1265.
- 127 Andrade R, Nicoll RA. Pharmacologically distinct actions of serotonin on single pyramidal neurones of the rat hippocampus recorded in vitro. *J Physiol (Lond)*. 1987;394:99–124.
- 128 Colino A, Halliwell JV. Differential modulation of three separate K-conductances in hippocampal CA1 neurons by serotonin. *Nature*. 1987;328:73–77.
- 129 Chaput Y, de Montigny C. Effects of the 5-hydroxytryptamine<sub>1</sub> receptor antagonist, BMY 7378, on 5-hydroxytryptamine neurotransmission: electrophysiological studies in the rat central nervous system. *J Pharmacol Exp Ther*. 1988;246:359–369.
- 130 Beck SG, Halloran PM. Imipramine alters beta-adrenergic, but not serotonergic, mediated responses in rat hippocampal pyramidal cells. *Brain Res*. 1989;504:72–81.
- 131 Tada K, Kasamo K, Ueda N, Suzuki T, Kojima T, Ishikawa K. Anxiolytic 5-hydroxytryptamine<sub>1A</sub> agonists suppress firing activity of dorsal hippocampus CA1 pyramidal neurons through a postsynaptic mechanism: Single-unit study in unanesthetized, unrestrained rats. *J Pharmacol Exp Ther*. 1999;288:843–848.
- 132 Lapin IP, Oxenkrug GF. Intensification of the central serotonergic processes as a possible determinant of the thymoleptic effect. *Lancet*. 1969;1:132–136.
- 133 Maes M, Meltzer HY. The serotonin hypothesis of depression. In: Bloom FE, Kupfer DJ, editors. *Psychopharmacology: the fourth generation of progress*. New York; Raven Press: 1995. p. 933–944.
- 134 Delgado PL, Charney DS, Price LH, Aghajanian GK, Landis H, Henninger GR. Serotonin function and the mechanism of antidepressant action. Reversal of antidepressant-induced remission by rapid depletion of plasma tryptophan. *Arch Gen Psychiatry*. 1990;47:411–418.
- 135 Heninger GR, Delgado PL, Charney DS, Price LH, Aghajanian GK. Tryptophan-deficient diet and amino acid drink deplete plasma tryptophan and induce a relapse of depression in susceptible patients. *J Chem Neuroanat*. 1992;5:347–348.
- 136 Delgado PL, Moreno FA. Role of norepinephrine in depression. *J Clin Psychiatry*. 2000;61 Suppl 1:5–12.
- 137 Blier P, de Montigny C. Current advances and trends in the treatment of depression. *Trends Pharmacol Sci*. 1994;15:220–226.
- 138 Chaput Y, de Montigny C, Blier P. Effects of a selective 5-HT reuptake blocker, citalopram, on the sensitivity of 5-HT auto-receptors: Electrophysiological studies in the rat. *Naunyn Schmiedeberg Arch Pharmacol*. 1986;333:342–348.
- 139 Blier P, de Montigny C, Tardif D. Effects of the two antidepressant drugs mianserin and indalpine on the serotonergic system: single cell studies in the rat. *Psychopharmacology*. 1984;84: 242–249.
- 140 Blier P, de Montigny C. Electrophysiological investigations on the effect of repeated zimelidine administration on serotonergic neurotransmission in the rat. *J Neurosci*. 1983;3:1270–1278.
- 141 Kreiss D, Lucki I. Effects of acute and repeated administration of antidepressant drugs on extracellular levels of 5-hydroxytryptamine measured in vivo. *J Pharmacol Exp Ther*. 1995;

- 274:866–876.
- 142 Le Poul E, Boni C, Hanoun N, Laporte AM, Laaris N, Chauveau J, et al. Differential adaptation of brain 5-HT<sub>1A</sub> and 5-HT<sub>1B</sub> receptors and 5-HT transporter in rats treated chronically with fluoxetine. *Neuropharmacology*. 2000;39:110–122.
  - 143 Le Poul E, Laaris N, Doucet E, Laporte AM, Hamon M, Lanfumey L. Early desensitization of somato-dendritic 5-HT<sub>1A</sub> autoreceptors in rats treated with fluoxetine or paroxetine. *Naunyn Schmiedeberg Arch Pharmacol*. 1995;352:141–148.
  - 144 Bel N, Artigas F. Chronic treatment with fluvoxamine increases extracellular serotonin in frontal cortex but not in raphe nuclei. *Synapse*. 1993;15:243–245.
  - 145 Pejchal T, Foley M, Kosofsky B, Waeber C. Chronic fluoxetine treatment selectively uncouples raphe 5-HT<sub>1A</sub> receptors as measured by [<sup>35</sup>S]-GTP $\gamma$ S autoradiography. *Br J Pharmacol*. 2002;135:1115–1122.
  - 146 Varrault A, Levieil V, Bockaert J. 5-HT<sub>1A</sub>-sensitive adenylyl cyclase of rodent hippocampal neurons: effects of antidepressant treatments and chronic stimulation with agonists. *J Pharmacol Exp Ther*. 1991;257:433–438.
  - 147 Narita N, Hashimoto K, Tomitaka S-I, Minabe Y. Interaction of selective serotonin reuptake inhibitors with subtypes of  $\sigma$  receptors in the rat brain. *Eur J Pharmacol*. 1996;307:117–119.
  - 148 Schmidt A, Lebel L, Koe BK, Seeger T, Heym J. Sertraline potently displaces (+)-[<sup>3</sup>H]-PPP binding to sigma-sites in rat brain. *Eur J Pharmacol*. 1989;165:335–336.
  - 149 Itzhak Y, Kassim CO. Clorgyline displays high affinity for sigma-binding sites in C57BL/6 mouse brain. *Eur J Pharmacol*. 1990;176:107–108.
  - 150 Shirayama Y, Nishikawa T, Umino A, Takahashi K. p-Chlorophenylalanine-reversible reduction of  $\sigma$  binding sites by chronic imipramine treatment in rat brain. *Eur J Pharmacol*. 1993;237:117–126.
  - 151 Matsuno K, Nakazawa M, Okamoto K, Kawashima Y, Mita S. Binding properties of SA4503, a novel and selective sigma 1 receptor agonist. *Eur J Pharmacol*. 1996;306:271–279.
  - 152 Kinsora JJ Jr, Corbin A, Snider B, Wiley J, Meltzer L, Heffner T, et al. Effects of igmesine in preclinical antidepressant tests [abstract]. *Soc Neurosci*. 1998;24:744.
  - 153 Urani A, Romieu P, Roman FJ, Yamada K, Noda Y, Kamei H, et al. Enhanced antidepressant efficacy of sigma(1) receptor agonists in rats after chronic intracerebroventricular infusion of beta-amyloid-(1-40) protein. *Eur J Pharmacol*. 2004;486:151–161.
  - 154 Ukai M, Maeda H, Nanya Y, Kameyama T, Matsuno K. Beneficial effects of acute and repeated administrations of sigma receptor agonists on behavioral despair in mice exposed to tail suspension. *Pharmacol Biochem Behav*. 1998;61:247–252.
  - 155 Skuza G, Rogoz Z. A potential antidepressant activity of SA4503, a selective sigma1 receptor agonist. *Behav Pharmacol*. 2002;13:537–543.
  - 156 Oshiro Y, Sakurai Y, Sato S, Kurahashi N, Tanaka T, Kikuchi T, et al. 3,4-dihydro-2(1H)-quinolinone as a novel antidepressant drug: synthesis and pharmacology of 1-[3-[4-(3-chlorophenyl)-1-piperazinyl]propyl]-3,4-dihydro-5-methoxy-2(1H)-quinolinone and its derivatives. *J Med Chem*. 2000;43:177–189.
  - 157 Tottori K, Miwa T, Uwahodo Y, Yamada S, Nakai M, Oshiro Y, et al. Antidepressant-like responses to the combined sigma and 5-HT<sub>1A</sub> receptor agonist OPC-14523. *Neuropharmacology*. 2001;41:976–988.
  - 158 Craven R, Grahame-Smith D, Newberry N. WAY-100635 and GR127935: effects on 5-hydroxytryptamine-containing neurones. *Eur J Pharmacol*. 1994;271:R1–R3.
  - 159 Watanabe Y, Yamada S, Nakai M, Uwahodo Y, Miwa T, Tottori K, et al. The rapid antidepressant-like effect of OPC-14523 in the forced swimming test is independent of action on 5-HT nerve terminals [abstract]. *Soc Neurosci*. 2000;26.
  - 160 Yamada S, Uwahodo Y, Tottori K, Kikuchi T, Altar C. Role of sigma and 5-HT<sub>1A</sub> receptors in the forced swimming test: supporting the mechanism of action of OPC-14523 [abstract]. *Soc Neurosci*. 2000;26:2326.
  - 161 Kamei H, Noda Y, Nabeshima T. The psychological stress model using motor suppression. *Folia Pharmacol Jpn (Nippon Yakurigaku Zasshi)*. 1999;113:113–120. (text in Japanese with English abstract)
  - 162 Noda Y, Kamei H, Nabeshima T. Sigma-receptor ligands and anti-stress actions. *Folia Pharmacol Jpn (Nippon Yakurigaku Zasshi)*. 1999;114:43–49. (text in Japanese with English abstract)
  - 163 Sanchez C, Papp M. The selective sigma2 ligand Lu 28-179 has an antidepressant-like profile in the rat chronic mild stress model of depression. *Behav Pharmacol*. 2000;11:117–124.
  - 164 Sanchez C, Arnt J, Costall B, Kelly ME, Meier E, Naylor RJ, et al. The selective  $\sigma_2$ -ligand Lu 28-179 has potent anxiolytic-like effects in rodents. *J Pharmacol Exp Ther*. 1997;283:1323–1332.
  - 165 Su TP, Wu XZ, Cone EJ, Shukla K, Gund TM, Dodge AL, et al. Sigma compounds derived from phencyclidine: identification of PRE-084, a new, selective sigma ligand. *J Pharmacol Exp Ther*. 1991;259:543–550.
  - 166 Matsumoto RR, Bowen WD, Tom MA, Vo VN, Truong DD, De Costa BR. Characterization of two novel  $\sigma$  receptor ligands: Antidystonic effects in rats suggest  $\sigma$  receptor antagonism. *Eur J Pharmacol*. 1995;280:301–310.
  - 167 Pande AC, Geneve J, Scherrer B. Igmesine, a novel sigma ligand, has antidepressant properties. *Int J Neuropsychopharmacol*. 1998;1:S8–S9.
  - 168 John CS, Vilner BJ, Bowen WD. Synthesis and characterization of [<sup>125</sup>I]-N-(N-benzylpiperidin-4-yl)-4-iodobenzamide, a new  $\sigma$  receptor radiopharmaceutical: high affinity binding to MCF-7 breast tumor cells. *J Med Chem*. 1994;37:1737–1739.
  - 169 Whittemore ER, Ilyin VI, Woodward RM. Antagonism of N-methyl-D-aspartate receptors by  $\sigma$  site ligands: potency, subtype-selectivity and mechanisms of inhibition. *J Pharmacol Exp Ther*. 1997;282:326–338.
  - 170 Fletcher EJ, Church J, Abdel-Hamid K, Macdonald JF. Blockade by sigma site ligands of N-methyl-D-aspartate-evoked responses in rat and mouse cultured hippocampal pyramidal neurones. *Br J Pharmacol*. 1995;116:2791–2800.
  - 171 Pontecorvo MJ, Karbon EW, Goode S, Clissold DB, Borosky SA, Patch RJ, et al. Possible cerebroprotective and in vivo NMDA antagonist activities of sigma agents. *Brain Res Bull*. 1991;26:461–465.
  - 172 Yamamoto H, Yamamoto T, Sagi N, Klenerova V, Goji K, Kawai N, et al. Sigma ligands indirectly modulate the NMDA receptor-ion channel complex on intact neuronal cells via sigma 1 site. *J Neurosci*. 1995;15:731–736.
  - 173 Hayashi T, Kagaya A, Takebayashi M, Shimizu M, Uchitomi Y, Motohashi N, et al. Modulation by sigma ligands of intracellular free Ca<sup>++</sup> mobilization by N-methyl-D-aspartate in primary

- culture of rat frontal cortical neurons. *J Pharmacol Exp Ther.* 1995;275:207–214.
- 174 Bermack JE, Debonnel G. Distinct modulatory roles of sigma receptor subtypes on glutamatergic responses in the dorsal hippocampus. *Synapse.* 2005;55:37–44.
- 175 Debonnel G, Bergeron R, de Montigny C. Electrophysiological evidence for the existence of sub-types of sigma receptors in the rat dorsal hippocampus. I. Antagonistic effects of high doses of sertraline, clorgyline and L-687-384 [abstract]. *Soc Neurosci.* 1992;18:16.10.
- 176 Perregaard J, Moltzen EK, Meier E, Sánchez C.  $\sigma$  ligands with subnanomolar affinity and preference for the  $\sigma_2$  binding site. 1. 3-(omega-aminoalkyl)-1H-indoles. *J Med Chem.* 1995;38:1998–2008.
- 177 De Costa BR, He XS, Dominguez C, Cutts J, Williams W, Bowen WD. A new approach to the design of sigma-2-selective ligands: synthesis and evaluation of N-[2-(3,4-dichlorophenyl)ethyl]-N-methyl-2-(1-pyrrolidinyl)ethylamine-related polyamines at sigma-1 and sigma-2 receptor subtypes. *J Med Chem.* 1994;37:314–321.
- 178 Couture S, Debonnel G. Modulation of the neuronal response to N-methyl-D-aspartate by selective sigma<sub>2</sub> ligands. *Synapse.* 1998;29:62–71.
- 179 Monnet FP, Mahe V, Robel P, Baulieu EE. Neurosteroids, via  $\sigma$  receptors, modulate the [<sup>3</sup>H]norepinephrine release evoked by N-methyl-D-aspartate in the rat hippocampus. *Proc Natl Acad Sci USA.* 1995;92:3774–3778.
- 180 Gonzalez-Alvarez GM, Werling LL. Sigma receptor regulation of norepinephrine release from rat hippocampal slices. *Brain Res.* 1995;673:61–69.
- 181 Bergeron R, de Montigny C, Debonnel G. Potentiation of neuronal NMDA response induced by dehydroepiandrosterone and its suppression by progesterone: effects mediated via sigma receptors. *J Neurosci.* 1996;16:1193–1202.
- 182 Pepe GJ, Rothchild I. A comparative study of serum progesterone levels in pregnancy and in various types of pseudopregnancy in the rat. *Endocrinology.* 1974;95:275–279.
- 183 Klink R, Robichaud M, Debonnel G. Gender and gonadal status modulation of dorsal raphe nucleus serotonergic neurons: I. Effects of gender and pregnancy. *Neuropharmacology.* 2002;43:1119–1128.
- 184 Jesberger JA, Richardson JS. Brain output dysregulation induced by olfactory bulbectomy: an approximation in the rat of major depressive disorder in humans? *Intern J Neurosci.* 1988;38:241–265.
- 185 Lumia AR, Teicher MH, Salchli F, Ayers E, Possidente B. Olfactory bulbectomy as a model for agitated hyposerotonergic depression. *Brain Res.* 1992;587:181–185.
- 186 Robichaud M, Beauchemin V, Lavoie N, Dennis T, Debonnel G. Effects of bilateral olfactory bulbectomy on N-methyl-D-aspartate receptor function: autoradiographic and behavioral studies in the rat. *Synapse.* 2001;42:95–103.
- 187 Campbell BG, Scherz MW, Keana JF, Weber E. Sigma receptors regulate contractions of the guinea pig ileum longitudinal muscle/myenteric plexus preparation elicited by both electrical stimulation and exogenous serotonin. *J Neurosci.* 1989;9:3380–3391.
- 188 Skuza G, Rogoz Z, Golembiowska K. EMD 57445, the selective sigma receptor ligand, has no effect on the 5-hydroxytryptamine system. *Pol J Pharmacol.* 1997;49:489–493.
- 189 Akunne HC, Whetzel SZ, Wiley JN, Corbin AE, Ninteman FW, Tecle H, et al. The pharmacology of the novel and selective sigma ligand, PD 144418. *Neuropharmacology.* 1997;36:51–62.
- 190 Blier P, de Montigny C. Serotonergic but not noradrenergic neurons in rat central nervous system adapt to long-term treatment with monoamine oxidase inhibitors. *Neuroscience.* 1985;16:949–955.
- 191 Blier P, Chaput Y, de Montigny C. Long-term 5-HT reuptake blockade, but not monoamine oxidase inhibition, decreases the function of terminal 5-HT autoreceptors: an electrophysiological study in the rat brain. *Naunyn Schmiedebergs Arch Pharmacol.* 1988;337:246–254.
- 192 Chaput Y, de Montigny C, Blier P. Presynaptic and postsynaptic modifications of the serotonin system by long-term administration of antidepressant treatments. *Neuropsychopharmacology.* 1991;5:219–229.
- 193 Bermack J, Debonnel G. Modulation of serotonergic neurotransmission by short- and long-term treatments with sigma ligands. *Br J Pharmacol.* 2001;134:691–699.
- 194 Bermack J, Haddjeri N, Debonnel G. The potential antidepressant OPC-14523 a combined sigma and 5-HT<sub>1A</sub> ligand: modulation of dorsal raphe nucleus firing activity. *J Pharmacol Exp Ther.* 2004;310:578–583.
- 195 Robichaud M, Klink R, Debonnel G. Gender and gonadal status modulation of dorsal raphe nucleus serotonergic neurons. Part II. Regulatory mechanisms. *Neuropharmacology.* 2002;43:1129–1138.
- 196 Celada P, Puig MV, Casanovas JM, Guillazo G, Artigas F. Control of dorsal raphe serotonergic neurons by the medial prefrontal cortex: involvement of serotonin-1A, GABA(A), and glutamate receptors. *J Neurosci.* 2001;21:9917–9929.
- 197 Aghajanian GK, Wang RY. Habenular and other midbrain raphe afferents demonstrated by a modified retrograde tracing technique. *Brain Res.* 1977;122:229–242.
- 198 Kalen P, Karlson M, Wiklund L. Possible excitatory amino acid afferents to nucleus raphe dorsalis of the rat investigated with retrograde wheat germ agglutinin and D-[<sup>3</sup>H]aspartate tracing. *Brain Res.* 1985;360:285–297.
- 199 Peyron C, Petit JM, Rampon C, Jouvet M, Luppi PH. Forebrain afferents to the rat dorsal raphe nucleus demonstrated by retrograde and anterograde tracing methods. *Neuroscience.* 1998;82:443–468.
- 200 Ferraro G, Montalbano ME, Sardo P, La Grutta V. Lateral habenular influence on dorsal raphe neurons. *Brain Res Bull.* 1996;41:47–52.
- 201 Varga V, Kocsis B, Sharp T. Electrophysiological evidence for convergence of inputs from the medial prefrontal cortex and lateral habenula on single neurons in the dorsal raphe nucleus. *Eur J Neurosci.* 2003;17:280–286.
- 202 O'Hearn E, Molliver M. Organization of raphe-cortical projections in rat: a quantitative retrograde study. *Brain Res Bull.* 1984;13:709–726.
- 203 Ceci A, Baschiroto A, Borsini F. The inhibitory effect of 8-OH-DPAT on the firing activity of dorsal raphe serotonergic neurons in rats is attenuated by lesion of the frontal cortex. *Neuropharmacology.* 1994;33:709–713.
- 204 Hajos M, Richards CD, Szekeley AD, Sharp T. An electrophysiological and neuroanatomical study of the medial prefrontal cortical projection to the midbrain raphe nuclei in the rat. *Neuroscience.* 1998;87:95–108.

- 205 Varga V, Szekely AD, Csillag A, Sharp T, Hajos M. Evidence for a role of GABA interneurons in the cortical modulation of midbrain 5-hydroxytryptamine neurones. *Neuroscience*. 2001;106:783–792.
- 206 Hajos M, Gartside SE, Varga V, Sharp T. In vivo inhibition of neuronal activity in the rat ventromedial prefrontal cortex by midbrain-raphe nuclei: role of 5-HT1A receptors. *Neuropharmacology*. 2003;45:72–81.
- 207 Pan ZZ, Williams JT. Differential actions of cocaine and amphetamine on dorsal raphe neurons in vitro. *J Pharmacol Exp Ther*. 1989;251:56–62.
- 208 Wang QP, Ochiai H, Nakai Y. GABAergic innervation of serotonergic neurons in the dorsal raphe nucleus of the rat studied by electron microscopy double immunostaining. *Brain Res Bull*. 1992;29:943–948.
- 209 Abellan MT, Adell A, Honrubia MA, Mengod G, Artigas F. GABAB-RI receptors in serotonergic neurons: effects of baclofen on 5-HT output in rat brain. *NeuroReport*. 2000;11:941–945.
- 210 Takebayashi M, Hayashi T, Su TP. Nerve growth factor-induced neurite sprouting in PC12 cells involves sigma-1 receptors: implications for antidepressants. *J Pharmacol Exp Ther*. 2002;303:1227–1237.
- 211 Urani A, Romieu P, Portales-Casamar E, Roman FJ, Maurice T. The antidepressant-like effect induced by the sigma(1) (sigma(1)) receptor agonist igmesine involves modulation of intracellular calcium mobilization. *Psychopharmacology (Berl)*. 2002;163:26–35.
- 212 Morin-Surun MP, Collin T, Denavit-Saubie M, Baulieu EE, Monnet FP. Intracellular sigma1 receptor modulates phospholipase C and protein kinase C activities in the brainstem. *Proc Natl Acad Sci USA*. 1999;96:8196–8199.
- 213 Wilke R, Brettel M, Prince HM, Wolf M, Mckenna R, Serpell E, et al. K<sup>+</sup> channel modulation in rodent neurohypophysial nerve terminals by sigma receptors and not by dopamine receptors. *J Physiol (Lond)*. 1999;512:391–406.
- 214 Young LT, Bakish D, Beaulieu S. The neurobiology of treatment response to antidepressants and mood stabilizing medications. *J Psychiatry Neurosci*. 2002;27:260–265.
- 215 Chao MV. Neurotrophins and their receptors: a convergence point for many signalling pathways. *Nat Rev Neurosci*. 2003;4:299–309.
- 216 Manji HK, Quiroz JA, Sporn J, Payne JL, Denicoff K, A Gray N, et al. Enhancing neuronal plasticity and cellular resilience to develop novel, improved therapeutics for difficult-to-treat depression. *Biol Psychiatry*. 2003;53:707–742.
- 217 Bremner JD. Does stress damage the brain? *Biol Psychiatry*. 1999;45:797–805.
- 218 Sheline YI, Wang PW, Gado MH, Csernansky JG. Hippocampal atrophy in recurrent major depression. *Proc Natl Acad Sci USA*. 1996;93:3908–3913.
- 219 Sheline YI, Sanghavi M, Mintun MA, Gado MH. Depression duration but not age predicts hippocampal volume loss in medically healthy women with recurrent major depression. *J Neurosci*. 1999;19:5034–5043.
- 220 Sheline Y, Gado M, Kraemer H. Untreated depression and hippocampal volume loss. *Am J Psychiatry*. 2003;160:1516–1518.
- 221 Sapolsky R. The possibility of neurotoxicity in the hippocampus in major depression: A primer on neuron death. *Biol Psychiatry*. 2000;48:755–765.
- 222 Malberg J, Eisch A, Nestler E, Duman R. Chronic antidepressant treatment increases neurogenesis in adult rat hippocampus. *J Neurosci*. 2000;20:9104–9110.
- 223 Hayashi T, Su T-P. Intracellular dynamics of sigma-1 receptor in NG-108 cells [abstract]. *Soc Neurosci*. 2001;31.
- 224 Nibuya M, Morinobu S, Duman RS. Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. *J Neurosci*. 1995;15:7539–7547.
- 225 Zetterstrom TS, Pei Q, Grahame-Smith DG. Repeated electroconvulsive shock extends the duration of enhanced gene expression for BDNF in rat brain compared with a single administration. *Brain Res Mol Brain Res*. 1998;57:106–110.
- 226 Altar CA, Whitehead RE, Chen R, Wortwein G, Madsen TM. Effects of electroconvulsive seizures and antidepressant drugs on brain-derived neurotrophic factor protein in rat brain. *Biol Psychiatry*. 2003;54:703–709.
- 227 Siuciak JA, Lewis DR, Wiegand SJ, Lindsay RM. Antidepressant-like effect of brain derived neurotrophic factor (BDNF). *Pharmacol Biochem Behav*. 1997;56:131–137.
- 228 Shirayama Y, Chen AC, Nakagawa S, Russell DS, Duman RS. Brain-derived neurotrophic factor produces antidepressant effects in behavioral models of depression. *J Neurosci*. 2002;22:3251–3261.
- 229 Ovalle S, Andreu F, Perez MP, Zamanillo D, Guitart X. Effect of the novel sigma1 receptor ligand and putative atypical antipsychotic E-5842 on BDNF mRNA expression in the rat brain. *NeuroReport*. 2002;13:2345–2348.
- 230 Takebayashi M, Hayashi T, Su TP. Sigma-1 receptors potentiate epidermal growth factor signaling towards neuriteogenesis in PC12 cells: potential relation to lipid raft reconstitution. *Synapse*. 2004;53:90–103.
- 231 Abe K, Saito H. Epidermal growth factor selectively enhances NMDA receptor-mediated increase of intracellular Ca<sup>2+</sup> concentration in rat hippocampal neurons. *Brain Res*. 1992;587:102–108.