

Effect of Dietary L-arabinose on the Intestinal Microbiota and Metabolism of Dietary Daidzein in Adult Mice

Motoi TAMURA^{1*}, Yukie KURUSU¹ and Sachiko HORI¹

¹National Food Research Institute, National Agriculture and Food Research Organization, 2–1–12 Kannondai, Tsukuba, Ibaraki 305-8642, Japan

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This study examined the effects of L-arabinose on mouse intestinal microbiota and urinary isoflavonoids. Male mice were randomly divided into two groups: those fed a 0.05% daidzein–2.5% L-arabinose diet (AR group) and those fed a 0.05% daidzein control diet (CO group) for 28 days. The amounts of daidzein detected in urine were significantly lower in the AR group than in the CO group. The ratio of equol/daidzein was significantly higher in the AR group ($p < 0.01$) than in the CO group. The composition of caecal flora differed between the AR and CO groups. The occupation ratios of *Prevotella* and *Lactobacillales* were significantly lower in the AR group. This study suggests that dietary L-arabinose has the potential to affect the metabolism of equol from daidzein by altering the metabolic activity of intestinal microbiota.

Key words: L-arabinose; equol; daidzein; mice; intestinal microbiota

Much attention has recently focused on the health benefits of soy-based foods. These health benefits have been largely attributed to isoflavones. Daidzin, genistin, daidzein (the aglycone of daidzin) and genistein (the aglycone of genistin) are the most common isoflavones found in soy products. Isoflavones are a class of phyto-oestrogens. Phyto-oestrogens are defined as compounds that exert estrogenic effects on the central nervous system, induce estrus and stimulate growth of the genital tract in female animals [1]. Human gastrointestinal bacteria seem to play an important role in isoflavone metabolism [2, 3]. Equol is a metabolite of daidzein and is produced by intestinal flora [4]. Equol is more effective than daidzein in competing with ³H-oestradiol to bind to the estrogen receptor, suggesting that it has a higher affinity for this receptor [5]. It has also been suggested that the ability to produce equol or equol itself is closely related to a lower incidence of prostate cancer [6]. Thus, equol is an important bacterial metabolite in the gut. However, interindividual variations in equol production have been identified. Only 30% to 50% of humans are equol producers [7]. It has been reported that the frequency of equol producers among vegetarians is

59%, similar to the reported frequency in Japanese adults consuming soy and much higher than that among non-vegetarian adults (25%), suggesting that dietary components other than soy influence S-equol synthesis by intestinal bacteria [8]. Rowland et al. [9] demonstrated that humans who excrete high concentrations of equol consume less fat and more carbohydrates as a percentage of energy than humans who excrete low concentrations of equol. The bioavailability of isoflavonoids seems to be affected by the composition of the diet. L-Arabinose, a natural, sweet and less absorptive pentose, has been recognized to have a selective inhibitory effect on intestinal sucrase activity in rats [10] and humans [11]. It has been reported that 4% L-arabinose in sucrose beverages reduces postprandial glucose, insulin and C-peptide responses and enhances the glucagon-like peptide-1 response in humans without gastrointestinal adverse effects [11]. Plasma insulin and triacylglycerol concentrations were significantly reduced by dietary arabinose in rats [12]. L-Arabinose ingestion with sucrose appears to effectively increase the production of organic acids in the caecum of the rat [13]. L-Arabinose affects gut function and intestinal microbiota [11, 13].

On the other hand, intestinal microbiota seem to play an important role in isoflavone metabolism [2, 4]. We tested the hypothesis that dietary L-arabinose changes the metabolism of daidzein and intestinal microbiota in

*Corresponding author. Mailing address: Motoi Tamura, National Food Research Institute, National Agriculture and Food Research Organization, 2–1–12 Kannondai, Tsukuba, Ibaraki 305-8642, Japan. Phone: +81-298-38-8089. Fax: +81-298-38-7996. E-mail: motoita@affrc.go.jp

mice. The aim of the present study was to investigate these effects in mice. Daidzein and equol were purchased from LC Laboratories (Woburn, MA, USA). β -Glucuronidase type H-5 was obtained from Sigma (St. Louis, MO, USA). Male Crj:CD-1 mice (6 weeks old) were purchased from Charles River Laboratories Japan, Inc. (Kanagawa, Japan). All mice were specific pathogen-free and were housed under conventional conditions in our laboratory. The mice were randomly divided into two groups of seven animals each. The animals were housed individually in suspended stainless steel cages with wire mesh bottoms in a room kept at $24 \pm 0.5^\circ\text{C}$ and a relative humidity of 65%, with 12-hr periods of light and dark. They were fed an AIN-93M diet for 7 d. After 7 d, the diet was replaced with a 0.05% daidzein control diet (CO diet) or a 0.05% daidzein–2.5% L-arabinose diet (AR diet), and the mice received the CO or AR diet for 28 d. Twenty-one days after beginning to feed the experimental diet, all animals were moved to metabolic cages (Tecniplast S.p.a., Buguggiate (Va), Italy). The animals were housed individually in these metabolic cages. Urine was collected from all mice. Urinary amounts of isoflavonoids were measured. The purified diet and water were provided *ad libitum*. The CO diet (control diet) contained 0.5 g/kg daidzein, 465.192 g/kg corn starch, 140 g/kg casein, 154.5 g/kg α -cornstarch, 100 g/kg sucrose, 40.0 g/kg soybean oil, 50.0 g/kg cellulose, 35.0 g/kg mineral mix (AIN-93M-Mix), (AIN-93-Mix) 10 g/kg vitamin mix, 1.8 g/kg L-cystine, 2.5 g/kg choline bitartrate, 0.008 g/kg tert-butylhydroquinone. The AR diet contained, 0.5 g/kg daidzein, 465.192 g/kg corn starch, 140 g/kg casein, 154.5 g/kg α -cornstarch, 100 g/kg sucrose, 40.0 g/kg soybean oil, 25.0 g/kg cellulose, 25.0 g/kg L-arabinose, 35.0 g/kg mineral mix (AIN-93M-Mix), 10 g/kg vitamin mix (AIN-93-Mix), 1.8 g/kg L-cystine, 2.5 g/kg choline bitartrate, 0.008 g/kg tert-butylhydroquinone. Body weight and food consumption were measured, and faeces were collected during the experiment. Faeces were dried with a freeze dryer (FD-1000; Tokyo Rikakikai Co., Ltd., Tokyo, Japan) for 24 hr. The trap cooling temperature was -45°C . Weights of freeze-dried faeces were measured during the experiment. After the experimental diet-feeding period, the mice were anaesthetized with diethyl ether, and blood samples were taken from the abdominal aorta and placed in heparinized tubes. All mice were euthanized with diethyl ether. The plasma was separated from whole blood by centrifugation and stored at -80°C until analysis of plasma glucose. The liver, visceral fat and caecal contents were collected. Caecal contents were stored at –

80°C until HPLC analysis for later terminal restriction fragment length polymorphism (T-RFLP) analysis of intestinal microbiota. The liver samples and visceral fat were weighed.

All procedures involving mice in this study were approved by the Animal Care Committee of the National Food Research Institute in Japan and were performed in accordance with the ‘Guidelines for Animal Care and Experimentation’ of the National Food Research Institute of the National Agriculture and Food Research Organization in Japan. A total of 200 μl of urine was added to 200 μl of β -glucuronidase H-5 solution (35 mg/ml; Sigma, St. Louis, MO, USA) in 0.2 M sodium acetate buffer (pH 5.0). Next, the mixture was incubated at 37°C in a water bath for 3 hr, followed by treatment with 400 μl of ethyl acetate, vortexing for 30 sec, and centrifugation at 5,000 g for 10 min at 4°C . The supernatants were transferred to an eggplant-type flask. The same volume of ethyl acetate as that used in the first extraction was added to the sediment, and the procedure was repeated. The supernatants from both extractions were pooled in the eggplant-type flask and evaporated completely using a rotary evaporator. The sample was then dissolved in 400 μl of 80% methanol and filtrated through a 0.2- μm filter. Filtrates were used for HPLC analysis. For HPLC analysis, we injected 20 μl of each preparation into a 250x4.6-mm Capcell Pak MG C18 5- μm column (Shiseido, Tokyo, Japan). To detect isoflavonoids, a photodiode array detector (MD-1515; JASCO, Corporation, Tokyo, Japan) was used to monitor the spectral data from 200 to 400 nm for each peak. To measure the isoflavonoids, we used daidzein and equol as standard samples. We used the spectral data at 254 nm to quantify daidzein and the spectral data at 280 nm to quantify equol. The mobile phase consisted of methanol/acetic acid/water (35:5:60, V/V/V). The running conditions of HPLC were a column temperature of 40°C and a flow rate of 1 ml/min. DNA extractions from caecal contents were conducted according to Matsuki’s method [14]. Caecal samples (20 mg) were washed three times by suspending them in 1.0 ml of phosphate-buffered saline and centrifuging each preparation at 14,000 g to remove possible PCR inhibitors. Following the third centrifugation, the caecal pellets were resuspended in a solution containing 0.2 ml of phosphate-buffered saline and 250 μl of extraction buffer (200 mM Tris-HCl, 80 mM EDTA; pH 9.0) and 50 μl of 10% sodium dodecyl sulfate. A total of 300 mg of glass beads (diameter, 0.1 mm) and 500 μl of buffer-saturated phenol were added to the suspension, and the mixture was vortexed vigorously for 60 sec using a Mini-

BeadBeater (BioSpec Products Inc., Bartlesville, OK, USA) at 4800 rpm. Following centrifugation at 14,000 g for 5 min, 400 µl of the supernatant was collected. Phenol-chloroform-isoamyl alcohol extractions were then performed, and 250 µl of the supernatant was subjected to isopropanol precipitation. Finally, the DNA was suspended in 1 ml of Tris-EDTA buffer. The DNA preparation was adjusted to a final concentration of 10 µg/ml in TE and checked by 1.5% agarose gel electrophoresis.

The PCR mixture (25 µl) was composed of EX Taq buffer, 2 mM of Mg^{2+} , and each deoxynucleoside triphosphate at a concentration of 200 µM. The amount of caecal DNA was 10 ng. The primers used were 5' FAM-labelled 516f (5'-TGCCAGCAGCCGCGGTA-3') and 1510r (5'-GGTTACCTTGTTACGACTT-3') at a concentration of 0.10 µM, template DNA and 0.625 U of DNA polymerase (TaKaRa EX Taq, Takara Bio Inc., Otsu, Japan). This process was carried out using a Dice PCR system (Takara Bio Inc.). Amplification was performed with one cycle at 95°C for 15 min; thirty cycles at 95°C for 30 sec, 50°C for 30 sec and 72°C for 1 min; and finally one cycle at 72°C for 10 min. The amplification products were subjected to gel electrophoresis in 1.5% agarose followed by ethidium bromide staining. The PCR products were purified using spin columns (QIAquick, Qiagen KK, Tokyo, Japan) according to the manufacturer's instructions. The purified DNA was treated with 2 U of BslI (New England Biolabs) for 3 hr at 55°C [15].

T-RFLP (terminal restriction fragment length polymorphism) analysis is based on PCR amplification of a target gene. The amplification is performed with one primer whose 5' end is labelled with a fluorescent molecule. The mixture of amplicons is then subjected to a restriction reaction using a restriction enzyme. Following the restriction reaction, the mixture of fragments is separated using either capillary or polyacrylamide electrophoresis in a DNA sequencer, and the sizes of the different terminal fragments are determined by the fluorescence detector. We used this T-RFLP analysis in our experiment. The fluorescently labelled terminal restriction fragments (T-RFs) were analyzed by electrophoresis on an automated sequence analyzer (ABI PRISM 310 Genetic Analyzer, Applied Biosystems) in GeneScan mode. The restriction enzyme digestion mixture (2 µl) was mixed with 0.5 µl of size standards (MapMarker 1000, BioVentures, Inc.) and 12 µl of deionized formamide. The mixture was denatured at 96°C for 2 min and immediately chilled on ice. The injection time was 30 sec for analysis of T-RFs from

digestion with BslI. The run time was 40 min. The lengths and peak areas of T-RFs were determined with the GeneMapper software. The predominant operational taxonomic units (OTUs), which correspond to either T-RFs or T-RF clusters, were detected in the T-RFLP profiles and used to identify phylogenetic groups of intestinal flora [15, 16].

Plasma glucose concentrations were measured using a glucose C2-test kit (Wako Pure Chemical Industries, Osaka, Japan) based on the mutarotase glucose oxidase method.

All faeces were collected during the experiment. Weights of faeces were measured. Faeces were then dried with a freeze dryer (FD-1000, Tokyo Rikakikai Co., Ltd., Tokyo, Japan) for 24 hr. The trap cooling temperature was 45°C. After drying, the weights of freeze-dried faeces were measured. Faecal lipid contents (%) were determined from the faeces sampled on the final days of the experiment. Faeces were milled with a food mill (TML17; TESCOM & Co., Ltd., Tokyo, Japan) for 30 sec. Faecal lipids were extracted from the faecal powder by Bligh and Dyer's method [17]. Data are expressed as the mean±standard error (SE). All data were analyzed using Sigma Plot 11 (Systat Software, Inc., San Jose, CA, USA). The data were analyzed with the t-test. Statistical significance was reached with a p-value of <0.05.

RESULTS AND DISCUSSION

No significant differences were observed between the AR and CO groups in final body weight (g) (AR, 33.3±0.6; CO, 32.7±0.6), food consumption (g/d) (AR, 4.15±0.05; CO, 4.23±0.04), visceral fat (g) (AR, 1.29±0.30; CO, 1.34±0.42), faecal contents (AR, 0.20±0.05; CO, 0.18±0.04) or liver weight (g) (AR, 1.35±0.05; CO, 1.40±0.03).

The urinary amounts of daidzein were significantly higher in the CO group than in the AR group ($p<0.05$) (Fig. 1A). Average urinary amounts of equol tended to be higher in the AR group than in the CO group. However, no significant differences were observed between the urinary amounts of equol in the AR and CO groups. The ratio of equol/daidzein was significantly higher in the AR group ($p<0.01$) than in the CO group (Fig. 1B). No significant differences in the amounts of urine (ml) collected for 45 hr were observed between the AR (1.8 ± 0.6) and CO (1.2 ± 0.3) groups. No significant differences in plasma glucose (mg/dl) (AR, 292.8±11.5; CO, 269.6±99.2) were observed between the AR and CO groups. The AR diet significantly affected both the amount of faeces and the faecal lipid content. The

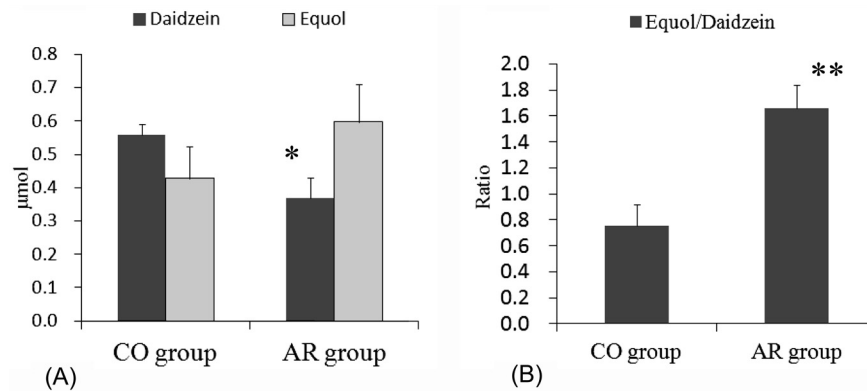


Fig. 1. (A) Amounts of urinary isoflavonoids (aglycones+metabolites) of mice in the AR group and the CO group. Values are means \pm SE (n=7). *Significantly different (p<0.05) from the CO group. (B) The ratio of equol/daidzein from the amounts of urinary isoflavonoids (aglycones+metabolites) of mice in the AR group and the CO group. Values are means \pm SE (n=7). **Significantly different (p<0.01) from the CO group. The data were analyzed using *t*-test analysis. Statistical significance was reached with a p value of less than 0.05.

amount of faeces was significantly greater in the CO group (0.34 ± 0.003) than in the AR group (0.25 ± 0.002) (p<0.01). The faecal lipid contents (% DW) of the faeces sampled on the final days of the experiment were significantly greater in the AR group (6.4 ± 0.3) than in the CO group (4.1 ± 0.4) (p<0.01). It has been confirmed that human intestinal microbiota predominantly consist of members of approximately ten phylogenetic bacterial groups and that these bacterial groups can be distinguished by the T-RFLP system developed by Nagashima et al. [15, 16]. Figure 2 depicts the compositions of the caecal flora, which differed between the two dietary groups. The occupation ratios of *Bacteroides* (OTUs 469, 853) were significantly higher in the AR group (p<0.01). The occupation ratios of *Bifidobacterium* (OTU 124) tended to be high in the AR group (p=0.068). On the other hand, the occupation ratios of *Lactobacillales* (OTUs 332, 520, 657) and *Prevotella* (OTUs 137, 317) were significantly higher in the CO group (p<0.01). Daidzein, a major component of the isoflavones, is metabolized to equol by the intestinal bacterial flora [4]. Conversion of daidzein to equol in the gut is important because of its strong estrogenicity compared with daidzein. However, there is large interindividual variation in the metabolism of isoflavones [18]. Equol was reportedly detected in urine from only 20% of subjects in a soya group across all ages and in only 5% of subjects in the control group following a soya challenge [19]. On the other hand, the metabolic activity of daidzein seems to be affected by diet [9].

In our results, the urinary amounts of daidzein were

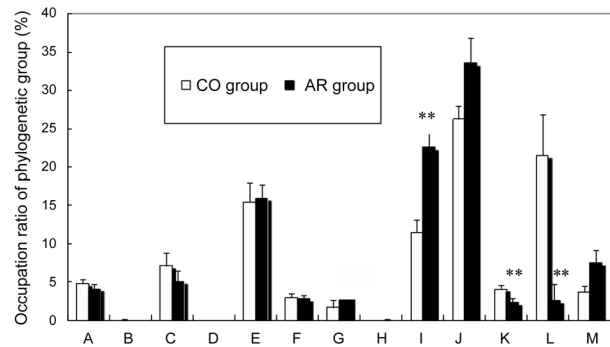


Fig. 2. Composition of caecal intestinal microflora of mice in the AR and CO groups. OTUs (operational taxonomic units), which correspond to either T-RFs (terminal restriction fragments) or T-RF clusters, were detected by T-RFLP analysis. Values are means \pm SE (n = 7). **Significantly different (p<0.01) from the CO group. The data were analyzed using *t*-test analysis. The letters correspond to the following phylogenetic bacterial groups: (A) *Bacteroides*, *Clostridium* cluster IV (OTU 370); (B) *Clostridium* cluster IV (OTUs 168, 749); (C) *Clostridium* cluster IX, *Megamonas* (OTU 110); (D) *Clostridium* cluster XI (OTU 338); (E) *Clostridium* subcluster XIVa (OTUs 106, 494, 505, 517, 754, 955, 990); (F) *Clostridium* cluster XI, *Clostridium* subcluster XIVa (OTU 919); (G) *Clostridium* subcluster XIVa, *Enterobacteriales* (OTU 940); (H) *Clostridium* cluster XVIII (OTUs 423, 650); (I) *Bacteroides* (OTUs 469, 853); (J) *Bifidobacterium* (OTU124); (K) *Lactobacillales* (OTUs 332, 520, 657); (L) *Prevotella* (OTUs 137, 317); (M) Others.

significantly higher in the CO group than in the AR group. The ratio of equol/daidzein was significantly higher in the AR group (p<0.01) than in the CO group.

Diet composition affects the gut microbial

composition to a larger extent than previously thought [20], and intestinal microbiota strongly affect isoflavone metabolism *in vivo* [21]. It has also been reported that subjects with higher PUFA and higher alcohol intake are more likely to be strong equol producers [22]. However, it has also been reported that increased probiotic yogurt or resistant starch intake does not affect isoflavone bioavailability in subjects consuming a high-soy diet [23]. Diet composition may affect equol production from daidzein in the gut by modifying the metabolic activity and/or composition of intestinal microflora. In our results, there were significant differences in the composition of microbiota between the AR and CO groups. It has been reported that butyric acid increases the conversion ratio of daidzein to equol in equol-producing bacteria [24]. Short-chain fatty acids (SCFAs) seem to affect the metabolism of daidzein. It has been reported that *Bifidobacterium* [25, 26] and *Bacteroides* [27, 28] produce SCFAs. In our experiment, the occupation ratios of *Bacteroides* and *Bifidobacterium* were higher in the AR group. Changes in productivity of SCFAs in the gut might be caused by *Bacteroides* and *Bifidobacterium*, and these changes might have affected the daidzein-metabolizing bacteria in the faecal flora of the AR group, resulting in the different metabolism between the two dietary groups.

L-Arabinose has been recognized to have a selective inhibitory effect on intestinal sucrase activity and reduces the digestion, absorption and net energy of sucrose when both are simultaneously ingested [10]. Apparent ileal digestibility of L-arabinose was found to be approximately 70%. The presence of L-arabinose in the diet increased ileal flow of volatile fatty acids and lactic acid, suggesting the occurrence of microbial degradation of L-arabinose in the gut of the pig [29]. L-Arabinose can be used as an energy source for pigs only after microbial fermentation. L-Arabinose feeding increased the production of SCFAs in the caecum of the rat [13]. From these reports, we assume that dietary L-arabinose itself may affect the composition and/or metabolic activity of intestinal microbiota by providing sucrose and L-arabinose to the intestinal microbiota in the lower gut. In our results, there were significant differences between the AR and CO groups in the composition of microbiota. Some intestinal bacteria can utilize L-arabinose. It has been reported that the growth of *Bifidobacterium longum* ATCC 15703 on L-arabinose-containing nutrient-poor medium was strong; however, *Bifidobacterium breve* ATCC 15700 showed no growth on L-arabinose-containing nutrient-poor

medium [30]. The different compositions of intestinal microbiota in the two dietary groups might be caused by the difference in availability of L-arabinose of intestinal bacteria. When uniformly ^{14}C -labelled sucrose was orally administered at a dose of 2.5 g/kg with or without L-arabinose and residual radio activities in the gastrointestinal tract were measured for 6 hr, a large quantity of undigested ^{14}C -labelled sucrose and its metabolites were observed in the caecum in L-arabinose-treated rats [10]. The different amounts of undigested sucrose in the caecum also contributed to the different compositions of intestinal microbiota in the two dietary groups.

In our results, no significant differences in plasma glucose or visceral fat were observed between the two groups. It has been reported that dietary L-arabinose reduces the blood glucose in rats [13]. The diet in this experiment by Fujii et al. contained 20% sucrose. On the other hand, our experimental diet contained only 14% sucrose. These different sucrose contents might have resulted in the different plasma glucose levels.

The AR diet significantly affected both the amount of faeces and faecal lipid content. The amount of faeces was significantly lower in the AR group than in the CO group ($p < 0.01$). Daily faeces production was reportedly greater in rats fed a diet containing a higher cellulose concentration than in rats fed a diet containing a lower cellulose concentration [31]. Significant increases in faecal weights of rats fed cellulose diets have also been reported [32]. In our experiment, the cellulose content of the diet was higher in the CO group than in the AR group. The higher cellulose content in the CO group may have caused the higher amounts of faeces in the CO group. The faecal lipid content (% DW) of the faeces sampled on the final days of the experiment were significantly greater in the AR group than in the CO group ($p < 0.01$). These results suggest that the lipid concentration in the lower gut may have been higher in the AR group. It has been reported that dietary fat significantly affects the intestinal microbiota [33, 34]. Differences in lipid concentrations in the gut might also affect the composition and/or metabolic activities of intestinal microbiota.

The limitation of our study was that we could not identify the bacteria that affect equol production from daidzein. This is the first report concerning the effects of dietary L-arabinose on the metabolism of daidzein in mice. Human studies are needed to clarify the relationship between equol production and metabolism of isoflavones.

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