

Considering the Embryopathogenesis of VACTERL Association

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Key Words

Embryology · Embryopathogenesis · Environmental influences · Genetics · Malformations · VACTERL association

Abstract

The nonrandom co-occurrence of vertebral, anorectal, cardiac, tracheoesophageal, genitourinary, and limb malformations, recognized as the VACTERL association, has not been satisfactorily explained from either a causation or embryopathogenesis standpoint. Few familial cases have been identified and maternal diabetes is the only environmental influence implicated to date. Mutations in single genes have been found in a number of syndromes with one or more of the VACTERL malformations, but these syndromes usually have other features which distinguish them from the VACTERL association. Animal models have provided clues to molecular pathways that may be involved in the embryogenesis of the VACTERL structures. What is lacking is the systematic study of individual genes and pathways in well-composed cohorts of patients, which is now possible with high throughput molecular technologies.

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Development of a unifying embryopathogenesis that encompasses the major malformations of the VACTERL association (vertebral defects, anorectal malformations, cardiac defects, tracheoesophageal fistula, renal anomalies,

and limb abnormalities) faces significant challenges. The malformations affect 5 different anatomical systems – skeletal, gastrointestinal, respiratory, cardiac and genitourinary. Some are axial, others appendicular. Some of the malformations make their appearance early in the embryological period, 23–30 days post conception, while others occur later in embryogenesis.

These challenges to a unifying embryopathogenesis notwithstanding, a number of potential explanations should be considered in seeking to understand why or how these malformations co-occur in nonrandom fashion. Among potential explanations are: (1) teratogenic influence(s) that are chronic throughout the period of embryogenesis, (2) an inaugural malformation that secondarily disturbs development of other anatomical structures, the so-called malformation sequence or cascade, (3) disturbances of molecular pathways or mutations of single genes that are critical in formation of the multiple anatomic systems affected, and (4) disturbances in a developmental process that is essential to all systems affected.

The Timing and Embryological Processes Involved in the VACTERL Malformations

The malformations in the VACTERL association develop at different stages of embryogenesis (fig. 1) [O’Rahilly and Müller, 2001; Sadler, 2012]. The anatomical structures involved are uniformly absent prior to 23 days post

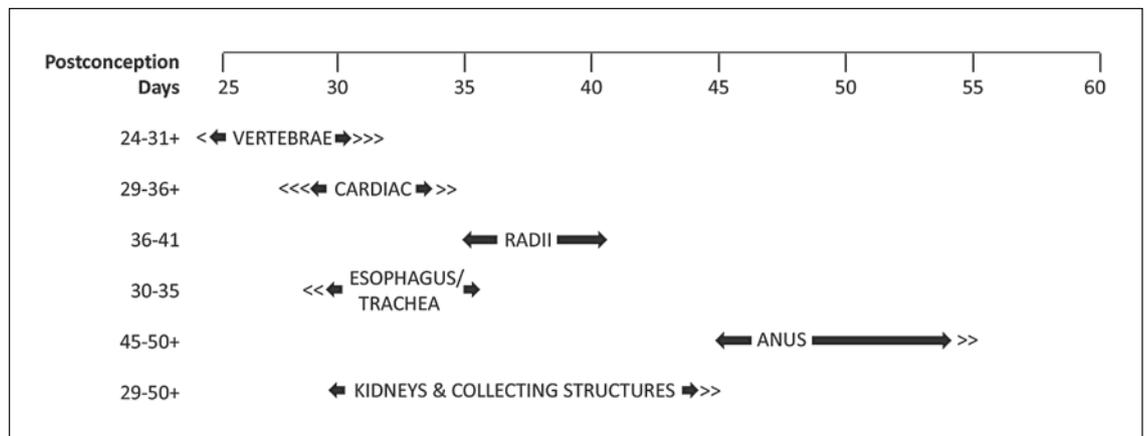


Fig. 1. Approximate days post conception during which anatomic structures in VACTERL association are formed. Genetic or environmental influences that affect the anlage of these structures may occur earlier than the days indicated.

conception and are more or less fully formed by 56 days post conception. The vertebrae, especially the thoracic vertebrae that are most often affected, are formed earliest in this embryological window (23–32 days), anorectal structures latest (45–56 days and beyond), and the heart, tracheoesophageal structures, and forearm bones in between (29–41 days). That disturbances may occur in processes that are active prior to the visible embryogenesis of an organ must also be considered.

It appears that multiple embryological processes are disturbed in the genesis of the VACTERL malformations. Abnormal or asymmetric timing of a molecular oscillator termed the segmentation clock has been shown to result in malsegmentation of the vertebrae [Pourquié and Kusumi, 2001; Oates et al., 2012]. Mesodermal proliferation and migration, epithelial-mesenchymal interactions and programmed cell death are involved in atresias of the esophagus and anus and in separation of the trachea and esophagus [Bergmann et al., 2003; Sadler, 2012]. Failure to lay down, condense or chondrify the anlage of the radius is involved in radial aplasia. Renal agenesis could result from failure, early in the 5th week, of the ureteric bud to develop appropriately or because of primary failure of the metanephric mesenchyme. Lower urinary defects involve abnormal development of the mesonephros/mesonephric duct. The common cardiac anomalies in VACTERL association – ventricular and atrial septal defects and Tetralogy of Fallot – share abnormal development of the cardiac septi, and they show clustering when there is intrafamilial recurrence [Fraser and Hunter, 1975].

Environmental Influences and VACTERL Association

Chronic exposure during the first trimester to environmental agents is a plausible explanation for the induction of malformations that make their appearance at different embryological times. Thalidomide exposure in early gestation affected multiple anatomical systems and offers the most dramatic and perhaps most relevant example. Although limb anomalies were the sentinel malformations of thalidomide exposure, craniofacial, cardiac, and gastrointestinal structures were also affected depending on the time of exposure [Knapp et al., 1962; Quibell, 1981; Lenz, 1988]. Midline vascular malformations of the face were present in most affected infants, malformations of the ears in 20%, cardiac malformations in 10%, and atresias or stenoses of the gastrointestinal system in less than 10% [Taussig, 1962; Quibell, 1981].

The evidence for an environmental influence that could account for a significant percentage of VACTERL association cases is quite meager. However, certain VACTERL malformations, particularly vertebral, cardiac and limb malformations, have been commonly found in infants of mothers with diabetes mellitus [Pedersen et al., 1964; Mills, 1986; Becerra et al., 1990; Janssen et al., 1996]. The mechanism by which maternal diabetes predisposes to malformations appears complex, involving hyperglycemia, oxidative stress, mitochondrial dysfunction, and disturbance of certain key developmental pathways [Reece, 2012]. Although some infants meeting VACTERL association criteria have been born of mothers with maternal diabetes, this is not the case for most affected in-

fants [Ewart-Toland et al., 2000; Stevenson, unpubl. observations].

Among the chemical teratogens, adriamycin, an anti-biotic cancer agent, can produce malformations in rats very similar to those of the VACTERL association [Beasley et al., 2000]. Most rat embryos exposed to adriamycin between gestation days 6 and 9 develop esophageal atresia (EA) with or without tracheoesophageal fistula (TEF), vertebral malsegmentation defects and cardiovascular anomalies, predominantly those involving conotruncal separation [Diez Pardo et al., 1996; Kotsios et al., 1998]. The upper limbs are more commonly malformed than the lower, but in a less severe and less specific way than in VACTERL association [Abu-Hijleh et al., 2000]. Anorectal and genitourinary malformations have not been reported in this model [Beasley et al., 2000]. Clues from the adriamycin rat model suggest that failure of the notochord to regionally organize the axial structures and failure of programmed cell death (apoptosis) to correctly separate and/or connect developing structures may be key processes in the induced embryopathology.

Gestational exposure to certain other drugs and chemicals (anticonvulsants, folic acid antagonists, alcohol) has also been linked to one or more of the VACTERL malformations, but none has been implicated in the full group of major VACTERL malformations [Milunsky et al., 1968; Shaw and Steinbach, 1968; Hanson and Smith, 1975; Hanson et al., 1976].

Malformation Sequences and VACTERL Association

David Smith, the founder of dysmorphology, proposed the concept of *malformation sequence* to encompass an inaugural malformation and the secondary consequences of that malformation [Smith, 1982]. ‘Potter Sequence’ was used to describe the secondary signs including oligohydramnios, pulmonary immaturity, and deformation of facial structures and limbs that result from renal agenesis. ‘Pierre Robin Sequence’ was used to describe micrognathia/retrognathia and the associated secondary glossoptosis and cleft palate. Although no such sequence has been proposed for VACTERL association, one might speculate that such a sequence could be initiated by vertebral malsegmentation with secondary malformative consequences on adjacent (tracheoesophageal, cardiac, and renal) structures and remote (limb and anal) structures. The secondary consequences might involve vascular disturbance(s) as an intermediary phenomenon (see below). The hint of a scenario such as this could be taken

from the adriamycin-induced rat model of VACTERL association in which maldevelopment of the notochord might be the initiating embryological disturbance [Beasley et al., 2000].

Single Gene Mutations among Conditions with Component Malformations Present in VACTERL Association

Few instances of first degree relatives with 3 or more VACTERL malformations have been identified [Nezerati and McLeod, 1999; Solomon et al., 2010b; Hilger et al., 2012]. In none of these families was a specific causative factor – genetic or environmental – found. This is in contrast with VACTERL-hydrocephalus in which numerous familial cases, with apparently both X-linked and autosomal patterns of inheritance, are reported [Hunter and MacMurray, 1987; Evans et al., 1989; Genuardi et al., 1993; Froster et al., 1996; Lomas et al., 1998]. *FANCB* mutations have been found in a minority of the X-linked cases [Holden et al., 2006; McCauley et al., 2011].

Each of the major malformations in VACTERL association occurs as an isolated anomaly and as a component of a number of malformation syndromes (fig. 2). Mutations in single genes have been identified in many of these syndromes, but it is the exception for all of the major VACTERL malformations to occur in any of these syndromes that generally also include malformations that are not seen in VACTERL association. One exception is the report of a missense mutation in *ZIC3*, an X-linked gene associated with heterotaxy, in a patient who had all of the VACTERL malformations and whose cousin and a nephew had cardiovascular anomalies [Ware et al., 2004]. *ZIC3* deletion or mutation has been reported in 2 other males who met criteria for VACTERL association [Wessels et al., 2010; Chung et al., 2011].

Limb anomalies, most often radial ray defects, occur in about half of patients diagnosed with VACTERL association [Weaver et al., 1986; Botto et al., 1997; Solomon et al., 2010a]. Syndromes with radial ray defects include those caused by mutations in *TBX5* (Holt-Oram syndrome), *RECQL4* (Baller-Gerold syndrome), *SALL1* (Townes-Brocks syndrome), and several *FANCF* genes (Fanconi anemia). Each of these syndromes includes one or more additional VACTERL malformation(s).

Tracheoesophageal anomalies are sentinel features of VACTERL association, occurring in over half of patients so diagnosed. The current knowledge of the genes involved in tracheoesophageal development was recently

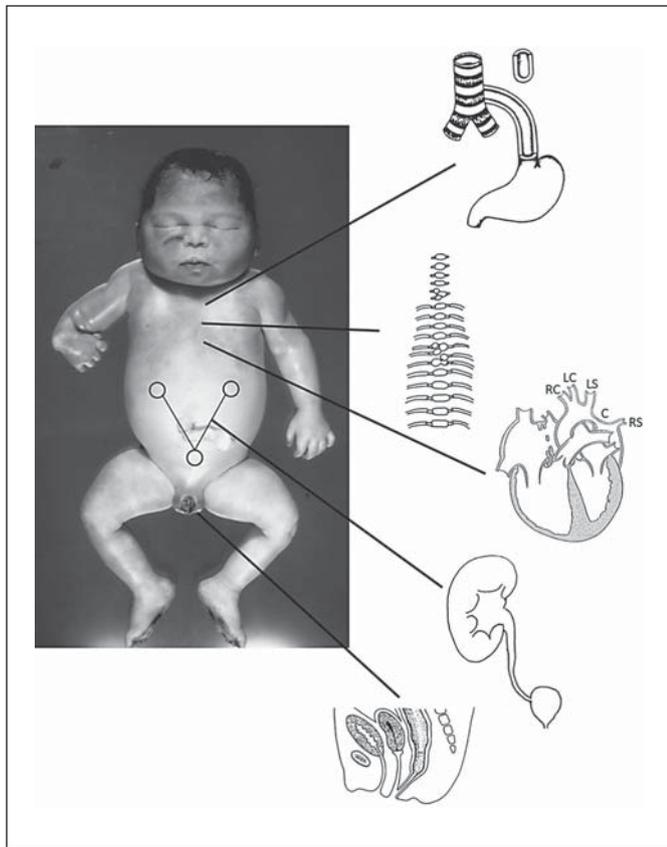


Fig. 2. Thirty-four week female fetus with VACTERL association: 14 thoracic vertebrae and ribs with multiple vertebral segmentation defects at C6–7, T5–6, and T6–7; ectopic hypoplastic anus at base of vagina; cardiomegaly with 2 atrial septal defects, a preductal coarctation (C) and aberrant major branches of aorta in order of right carotid (RC), left carotid (LC), left subclavian (LS), and right subclavian (RS) which coursed behind the esophagus and trachea; tracheoesophageal fistula and esophageal atresia; left renal agenesis and right hydronephrosis; and right radial aplasia with hypoplastic thumb.

reviewed by Faure and De Santa Barbara [2011]. Segmentation of the gut into different regions is controlled by the *Hox* genes with *Hoxa3* and *Hoxb4* being expressed in the foregut endoderm. *Shh* is expressed throughout the gut endoderm but has a specific expression pattern during tracheoesophageal development with earlier expression and patterning in the ventral foregut and a temporary inhibition of its expression in tracheal endoderm. *Sox2* is also expressed in the foregut, and the transcription factor *Nkx2* is specifically expressed in the endoderm of the developing trachea but not in the esophagus. Lack of *Nkx2* or *Shh* expression leads to failure of separation of the trachea from the esophagus. Endodermal cells secrete SHH

that binds to and activates Patched in the associated mesenchymal cells. Activated PTCH1 in turn causes activation of GLI1/2, which has effects on various mesenchyme-specific genes including *Ptch1*, *Gli1* and *Bmp4*, the latter of which is only expressed in the ventral foregut. Condensation of mesenchyme at the site of tracheoesophageal separation is noted and may be vital to that process. Syndromes in which TEF and/or EA occur, at least in a minority of cases, include Hypertelorism-hypospadias syndrome (*MID1* mutations), CHARGE syndrome (*CHD7* mutations) and VACTERL-hydrocephalus syndrome (*FANCB* mutations). TEF and the other major VACTERL anomalies were reported in a patient with a mutation in *ZIC3* by Ware et al. [2004]. Wessels et al. [2010] described a patient with anal atresia, laryngoesophageal and tracheoesophageal atresia, cardiac dextroposition with persistent left superior vena cava, and unilateral multicystic kidney who also had a *ZIC3* mutation. Chung et al. [2011] described a family in which an Xq26.3 deletion including the *ZIC3* gene segregated. One male met VACTERL-H criteria with imperforate anus, duplication of the descending colon, multiple cardiovascular anomalies, dysraphic spine with L3–4 block vertebra, fused kidneys, and hydrocephalus due to aqueductal stenosis. Two maternal male relatives with the deletion had imperforate anus and one maternal nephew (fetus) had heterotaxy, cardiovascular malformations and imperforate anus. Reardon et al. [2001] found a heterozygous missense mutation in *PTEN* in a male with TEF, macrocephaly, mild radial ray deficiency, and ‘fullness of the ventricles’, features they considered to resemble the VACTERL association. Arrington et al. [2010] proposed *LPP* as a candidate gene for EA/TEF/VACTERL. They reported a patient with EA/TEF that met VATER criteria and was deleted for only *LPP*. However, a screen of 195 patients with EA/TEF/VACTERL (70 with VACTERL) found no copy number variations or loss of heterozygosity [Hernández-García et al., 2012].

Multiple vertebral anomalies, usually in the form of hemivertebrae, fusions, extra or missing vertebrae and rib defects occur in most patients diagnosed with VACTERL association. Among other malformation syndromes with high rates of vertebral malformations are Alagille, spondylocostal dysplasia and Okihiro. Mutations in genes in the Notch signaling pathway (*JAG1* and *DLL3*) have been identified in Alagille syndrome and spondylocostal dysplasia and mutations in *SALL4* in Okihiro syndrome. NOTCH has a pivotal role in the coordination of gene expression in contiguous cells and hence the development of the somite segmentation clock [Lew-

is et al., 2009]. A signaling cell secretes a ligand, such as Jagged, that binds to the NOTCH receptor of the contiguous receiving cell, which results in NOTCH cleavage and release of its intracellular component which targets, among other genes, the *Hes* family. Interaction with the *Hes* pathway has been a suggested site of interaction with the *FANCB* gene in VACTERL-H (see below).

Cardiac malformations occur in over three-fourths of patients with VACTERL association [Solomon et al., 2010a]. Malformations with functional significance probably account for only half of this number. Numerous malformation syndromes that include cardiac defects are due to single gene mutations: Alagille, CHARGE, Feingold, Holt-Oram syndrome, Hypertelorism-hypospadias, Townes-Brocks, and VACTERL-hydrocephalus syndromes. In each, at least one other VACTERL malformation is usually present.

Imperforate anus occurs in about half of patients with VACTERL association. A spectrum of anorectal anomalies have been reported including fistulas into the genitourinary system and other genitourinary anomalies. Anal anomalies occur in several syndromes known to have single gene causation: Townes-Brocks (*SALL1*), Pallister-Hall (*GLI3*), Hypertelorism-hypospadias (*MIDI1*) and Currarino syndromes (*HLXB9* mutations).

Genitourinary anomalies in the form of horseshoe kidney, renal cysts, renal dysplasia, and renal agenesis with or without associated anomalies of the collecting structures or genitals occur in over half of patients. These malformations often exist in a subclinical state and rarely lead to organ failure. Genitourinary anomalies are commonly seen in a number of conditions which have other malformations of VACTERL association (CHARGE, Alagille, Feingold, Fryns, Pallister-Hall, Townes-Brocks, Smith-Lemli-Opitz, Okihiro, and other syndromes).

VACTERL-hydrocephalus is the only single gene disorder that encompasses all the VACTERL acronym malformations. This serious condition almost always results in early lethality. Hydrocephalus, the feature that distinguishes the condition from the sporadic VACTERL association, is due to aqueductal stenosis. Mutations in *FANCB* have been found in a minority of cases [Holden et al., 2006; McCauley et al., 2011].

Szumaska et al. [2008] noted that a mutation in *Pcsk5* in the mouse produced VACTERL-caudal regression-Currarino syndrome-like malformations. They screened 36 patients with VACTERL and 13 with caudal regression and found non-synonymous variants that were not observed in ethnically matched controls in 4 of the former

and 1 of the latter patients. The mouse mutant was shown to cause abnormal expression of several *Hox* genes and *Mnx1* (*Hlxb9*). The latter gene is a principal gene responsible for familial Currarino syndrome [Kim et al., 2007].

Molecular Pathways Involved in Formation of Anatomical Structures Affected in VACTERL Association

A number of signaling pathways are involved in the formation of the diverse anatomical structures affected in VACTERL association. Prominent among these are the Sonic hedgehog (SHH) pathway, known to be involved in brain, limb, and spine formation; the Notch (NOTCH) pathway, known to be involved in cardiovascular and spine formation; and the fibroblast growth factor (FGF) pathway, known to be involved in limb, vasculature, and spine formation.

Although argument has been made for disturbance of the SHH pathway as a cause for VACTERL association, no specific mutations in *SHH* or the closely related *GLI* genes have been found in patients with VACTERL association [Garcia-Barceló et al., 2008a; Aguinaga et al., 2010]. Mutations in *Shh*, *Gli2*, and *Gli3* have produced VACTERL malformations in mice [Kim et al., 2001]. Mutations in *HOXD13* and *FOXF1*, two genes linked to the SHH pathway, have been described in patients with VACTERL or VACTERL-like presentations [Garcia-Barceló et al., 2008b; Agochukwu et al., 2011]. However, the SHH pathway is complex with additional participants that may have potential to be involved in VACTERL association. These might include the mammalian equivalents of the activator Smoothed (Smo), the G-protein coupled receptor, and the cytoplasmic signaling components Cos2, Fused (Fu) and Suppressor of Fused (Sufu) [Varjosalo and Taipale, 2008].

The NOTCH pathway is an intercellular signaling complex involved in embryonic development of heart, vasculature, hematopoietic system, vertebrae and ribs, and other organs. The NOTCH receptors (NOTCH1–4) and other components of this pathway, including the membrane bound ligands JAG1, JAG2, DLL1, DLL2, and DLL3, influence various cellular processes including proliferation and apoptosis. Gametic mutations are not known for *NOTCH1*, 2 and 4, whereas those in *NOTCH3* cause CADASIL. Certain cardiac and vascular malformations and malsegmentation of the vertebrae are among the consequences of faulty NOTCH signaling; Alagille in the case of *JAG1* and spondylocostal dysplasia in the case of *DLL3* [High and Epstein, 2008]. The signaling pathway

includes the processing and subsequent cleavage and heterodimerization of extracellular, transmembrane and intracellular NOTCH components by the endoplasmic reticulum and Golgi [Kopan and Ilagan, 2009]. The heterodimer is moved to the cell membrane where it binds the JAG and DLL ligands resulting in cleavage from the NOTCH extracellular domain by TACE (ADAM metalloprotease TNF- α converting enzyme). The transmembrane and intracellular domains are split by γ -secretase and the NOTCH intracellular domain is moved to the nucleus where it becomes associated with the CSL family transcription complex with resulting activation of target genes including *Myc*, *p21* and the *Hes* family. Thus, the NOTCH signaling pathway includes a number of potential areas for exploration with respect to the causation of VACTERL association.

The FGF signaling pathway involves the various fibroblast growth factors, their receptors and other components that during embryogenesis stimulate angiogenesis, and influence neurogenesis, skeletal development, tracheal branching, and many other developmental processes. Mutations in the *FGFs* and *FGFRs* have not been implicated in specific VACTERL malformations, but mutations in the *FGFRs* have caused generalized skeletal dysplasias and craniosynostosis. Early in bone development, *FGFs* stimulate cell proliferation, while in more mature cells they promote apoptosis [Mansukhani et al., 2000]. Although disturbances in these cellular processes might play a role in VACTERL association, this pathway does not appear as a prime candidate to explain the association.

Disturbance in a Developmental Process and VACTERL Association

A number of developmental processes are involved in formation of all anatomical systems affected in the VACTERL association. Mesoderm production is an early and essential component of all anatomical structures. Adequate mesoderm production must occur via migration and transformation of precursor ectoderm through the primitive streak, proliferation, and distribution to all sites of embryogenesis. Disturbance of this fundamental embryological process plausibly might disrupt development globally, leading to embryonic death or locally producing faulty formation of individual anatomical structures. In their initial description of the VATER association, Quan and Smith [1973] suggested that a defect in mesoderm prior to day 35 post conception could result in

all of the component malformations. Others have also supported the concept that failure of normal mesoderm migration during the first 4 weeks of embryogenesis could be the initiating pathogenic event [Martinez-Frias and Frias, 1999; Bergmann et al., 2003]. A closely related developmental process is signaling between ectoderm and mesoderm, the so-called epithelial-mesenchymal interactions, which appears to be essential to the correct formation of most anatomical structures.

A later, but equally ubiquitous, developmental process essential to all anatomical structures is vascular supply. Adequate oxygen and nutrient supplies may be interrupted locally by faulty development of blood vessels and by hemorrhage or thrombosis. A vascular embryopathogenesis has been posited to account for most of the individual VACTERL malformations. Absence of the inferior mesenteric artery has been noted in association with anal atresia [Stevenson et al., 1986], disturbed formation of the intersegmental arteries has been observed with malsegmentation of the vertebrae [Müller et al., 1986; Stevenson et al., 1987] and absence of the radial artery recorded in radial aplasia [Duncan and Shapiro, 1993]. The presence of a single umbilical artery in a minority of infants with VACTERL association further invites the question of whether vascular pathology contributes to the co-occurrence of the VACTERL malformations [Quan and Smith, 1973; Auchterlonie and White, 1982; Weaver et al., 1986; Duncan and Shapiro, 1993; Shaw-Smith, 2010; Stevenson, unpubl. observations]. However, the major and unresolved criticism of the vascular hypothesis is whether the vascular changes observed cause malformations or are effects of malformations [Sadler and Rasmussen, 2010]. There may be some reason to suggest that primary thrombosis/hemorrhage is an unlikely explanation for VACTERL. For one there is a strong propensity for upper limb involvement and there is no obvious vascular basis for that observation. In addition, the limb defects in VACTERL tend to differ from those more often ascribed (without proof) to such thrombo-hemorrhagic events where there are generally terminal deficiencies with or without hypoplastic digits and nails.

However, there is some evidence in support of a more primary vascular pathogenesis for some VACTERL anomalies. Retinoic acid exposure of Syrian hamsters resulted, within 24 hours, in caudal vessel changes and dissection of the aortic contents into the adjacent unsegmented mesoderm [Wiley, 1983]. This was associated with neural tube and notochordal anomalies, and loss of intercellular relationships in the paraxial mesoderm. By 36 hours there were abnormalities in the appearance of

the caudal somites, and in later studied embryos there were defects in the sclerotomes and the vertebrae. There are experimental data implicating hypoxia as a teratogen [Chernoff and Rogers, 2010]. An early example is in rabbits exposed to hypoxia at human equivalent 23–30 days gestation that resulted in sagittal cleft vertebrae [Degenhardt and Knoche, 1959]. The notochord appeared to be the primary site of damage, resulting in faulty induction of the adjacent paraxial mesoderm. Embryos allowed to continue showed vertebral anomalies.

Discussion

In the 4 decades since the initial description and revisions of the VACTERL association, a vast number of diverse studies have been conducted on this malformation complex. Yet, this large body of clinical and epidemiological observations, animal experimentation, and molecular probing has not yielded a clear and unifying embryopathogenesis. A confounding feature of the VACTERL association that must be taken into account is that the embryologic timing differs between the component malformations.

A key characteristic of teratogenic influences that may bring together seemingly unrelated malformations is that chronic or repetitive exposure of the developing anatomy to an environmental or genetic influence may permit multiple organ systems to be affected even though they are on different developmental schedules. Environmental influences that might be active throughout the embryological period include exposure to teratogenic drugs or a maternal metabolic disturbance. In similar fashion, the effects of a single gene mutation might extend throughout the period of embryogenesis. In the decades of experience with the VACTERL association, no potential teratogen, pregnancy characteristic, or genetic alteration common to the majority or even a substantial minority of cases, has been identified.

Anatomically disparate anomalies may also follow a single early embryonic progenitor event that predestines disturbances in derivative cell populations. Since the VACTERL association was first recognized, there have been suggestions that such an early event that disturbs mesodermal migration, proliferation, differentiation and/or apoptosis could account for the component malformations. No specific observations in humans support this concept. However, the abnormal shape and positioning of the notochord and the abnormal timing and location of apoptosis in the foregut and hindgut in the adria-

mycin-induced rat model hint at an earlier progenitor defect [Beasley et al., 2000]. Clarification of this potential pathogenetic basis for the VACTERL malformation will depend on the further study of early embryogenesis possible only in animal models.

Mutations in certain individual genes have produced multiple malformation syndromes with some phenotypic overlap with VACTERL association [Shaw-Smith, 2010]. Mutations in *NMYC*, *GLI3*, *CHD7*, and *SALL1* cause Feingold, Pallister-Hall, CHARGE, and Townes-Brocks syndromes, respectively, each of which may exhibit 2 or more of the VACTERL malformations, but which generally have other distinctive findings that allow clinical differentiation. This is also the case for autosomal recessive Fanconi anemia, which may be caused by mutations in one of several *FANC* genes, with about 5% of patients meeting VACTERL association criteria.

Among patients with the full composite of major VACTERL malformations, a plausible specific causative factor is rarely identified. The exceptions include some cases associated with maternal diabetes, a few cases with *ZIC3* mutation or deletion and the cases of VACTERL-hydrocephalus with *FANCB* mutations [Ewart-Toland et al., 2000; Ware et al., 2004; Holden et al., 2006; McCauley et al., 2011]. Still, the case for causative mutations in a single gene to explain most cases of VACTERL association cannot be considered excluded. Application of current and future molecular technology in the systematic study of well-composed cohorts of patients is necessary to clarify the role of genetic alterations in the causation of this malformation complex.

While causal heterogeneity for VACTERL association has been widely considered as likely, this is by no means certain [Khoury et al., 1983; Botto et al., 1997; Shaw-Smith, 2006, 2010; Solomon, 2011]. The experience with CHARGE association/syndrome is instructive in this regard. Once considered to be an association, the divergent phenotype of CHARGE syndrome became understandable once the single pleiotropic gene, *CHD7*, was identified.

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