

## Forum Minireview

## Pharmacology in Health Food: Metabolism of Quercetin In Vivo and Its Protective Effect Against Arteriosclerosis

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**Abstract.** Quercetin, a member of the bioflavonoids family, has been proposed to have anti-atherogenic, anti-inflammatory, and anti-hypertensive properties leading to the beneficial effects against cardiovascular diseases. It was recently demonstrated that quercetin 3-*O*- $\beta$ -D-glucuronide (Q3GA) is one of the major quercetin conjugates in human plasma, in which the aglycone could not be detected. Although most of the in vitro pharmacological studies have been carried out using only the quercetin aglycone form, experiments using Q3GA would be important to discover the preventive mechanisms of cardiovascular diseases by quercetin in vivo. Therefore we examined the effects of the chemically synthesized Q3GA, as an in vivo form, on vascular smooth muscle cell (VSMC) disorders related to the progression of arteriosclerosis. Platelet-derived growth factor–induced cell migration and proliferation were inhibited by Q3GA in VSMCs. Q3GA attenuated angiotensin II–induced VSMC hypertrophy via its inhibitory effect on JNK and the AP-1 signaling pathway. Q3GA scavenged 1,1-diphenyl-2-picrylhydrazyl radical measured by the electron paramagnetic resonance method. In addition, immunohistochemical studies with monoclonal antibody 14A2 targeting the Q3GA demonstrated that the positive staining specifically accumulates in human atherosclerotic lesions, but not in the normal aorta. These findings suggest Q3GA would be an active metabolite of quercetin in plasma and may have preventative effects on arteriosclerosis relevant to VSMC disorders.

**Keywords:** quercetin glucuronide, cardiovascular disease, vascular smooth muscle cell, health food

### 1. Introduction

Bioflavonoids are polyphenolic compounds that are ubiquitously present in foods of plant origin (1). These constituents of the diet are believed to be important in the maintenance of health, especially to maintain the integrity of the cardiovascular system. The “French paradox” refers to the correlation of a high-fat and high-cholesterol diet with a lower incidence of coronary heart disease

found in Mediterranean cultures contrasted with a higher incidence of coronary heart disease among most Western cultures (2). It has been shown that the French paradox may be attributable to regular consumption of red wine and that the unique antiatherogenic effects of red wine reside in the action of polyphenols (3). Moreover, it was demonstrated that the DASH (Dietary Approaches to Stop Hypertension) clinical trial diet that was rich in fruit, vegetables, and low-fat dairy products had a significant hypotensive effect (4). Recently the Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH 2009) provides lifestyle modifications including increased intake of fruits and vegetables

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to prevent cardiovascular disease and risk factors (5). Quercetin (3,3',4',5,7-pentahydroxyflavone), a member of the bioflavonoids family, is one of the most widely distributed dietary polyphenolic compounds in foods including vegetables, fruits, tea, and wine (6). Like other members of the bioflavonoids, quercetin has been shown to have biological properties consistent with its sparing effect on the cardiovascular system. It has been shown that quercetin possesses, anti-atherogenic, anti-inflammatory, anti-coagulative, and anti-hypertensive properties (6–8). In addition, within the bioflavonoid family, quercetin is the most potent scavenger of reactive oxygen species (ROS) (9). The anti-oxidant activity of quercetin has frequently been mentioned in connection with its pharmacological function in the cardiovascular system (10) because oxidative modification of plasma low-density lipoprotein (LDL) was suggested to be involved in the initial event of atherosclerosis, leading to coronary heart disease (11). In this review, we have summarized mainly our data regarding the pharmacological effects of quercetin, especially its metabolites, on cardiovascular diseases.

## 2. Metabolism of quercetin in vivo

Quercetin is a prime example of such a flavonoid and is bound to sugars in foods, mainly as  $\beta$ -glycosides. Quercetin is mostly present in the form of glycosides in vegetables and fruits, and dietary glycosides were believed to be converted to the respective aglycones in the large intestine by the glycosidase activity of intestinal bacteria (12). However, it was recently demonstrated that quercetin 3-*O*- $\beta$ -D-glucuronide (Q3GA) and quercetin-3'-sulfate are the major quercetin conjugates in human plasma, in which aglycone could not be detected (13–15). Upon ingestion with the diet, quercetin glycosides are rapidly hydrolyzed during passage across the small intestine or by bacterial activity in the colon to generate quercetin aglycone, which is further metabolized in the so-called phase II reactions into the glucuronidated and/or sulfated derivatives (Fig. 1). Studies using rodents have also shown that orally administered quercetin is converted to its conjugates before accumulation in plasma (16, 17). In addition, it was reported that volunteer study clarified that conjugated metabolites of quercetin accumulate in human plasma in the concentration range of  $10^{-7}$ – $10^{-6}$  M after the periodic ingestion of onions with meals for 1 week (14). Therefore the pharmacological function of dietary quercetin, including anti-oxidant activity, should be exerted exclusively by its conjugated metabolites. Although most of the *in vitro* pharmacological studies have been carried out using only the quercetin aglycone form, experiments using Q3GA

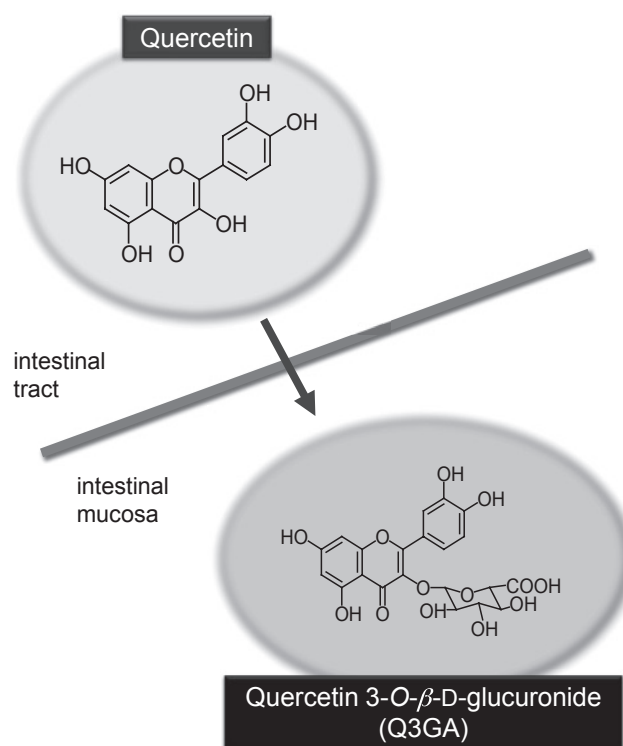


Fig. 1. Metabolism of quercetin in vivo.

would be important to discover the mechanisms through which quercetin exerts its preventative effects on cardiovascular diseases in vivo. Therefore we examined the effects of the chemically synthesized Q3GA, as an *in vivo* form, on vascular smooth muscle cell (VSMC) disorders related to the progression of cardiovascular diseases.

## 3. The effects of Q3GA on migration, proliferation and hypertrophy in VSMCs

The primary cause of many fatal cardiovascular diseases is believed to be atherosclerosis (18). During atherogenesis and the progression of the disease, chronic inflammatory responses induce vascular wall remodeling or the generation of neointima and thickening of the tunica media, which leads to the development of plaque and artery stenosis (19). The neointima and thickened media are primarily composed of abnormally proliferating and migrating VSMCs (20–23). Therefore, we examined the effect of chemically synthesized Q3GA, as an *in vivo* form, on platelet-derived growth factor (PDGF)-induced cell migration in VSMCs. PDGF has been recognized as a major mitogen and one of the most important growth factors, and it also stimulates VSMC migration (24, 25). It has been reported that PDGF-induced VSMC

migration and proliferation are mediated by mitogen-activated protein (MAP) kinases, phosphatidylinositol 3-kinase (PI3-kinase)/Akt, and many other kinases (26–28). MAP kinases are protein kinases and play an important role in cell differentiation, growth, apoptosis, and the regulation of a variety of transcription factors and gene expressions (29, 30). The activation of these MAP kinase pathways has been shown to be involved in the promotion of VSMC proliferation, migration, and hypertrophy that relates to vascular remodeling and altered tone in hypertension. Q3GA pretreatment significantly suppressed PDGF (10 ng/ml, 6 h)-induced VSMC migration in a concentration-dependent manner (10 and 100  $\mu$ M) (31). Also Q3GA significantly attenuated PDGF (10 ng/ml, 48 h)-induced rat aortic smooth muscle cell (RASMC) proliferation (10 and 100  $\mu$ M) (31). It has been reported that PDGF-induced VSMC migration and proliferation mediated by MAP kinases and the PI3-kinase/Akt pathway. PDGF-induced c-Jun N-terminal kinase (JNK) and Akt activations were inhibited by Q3GA in a concentration-dependent manner (10 and 100  $\mu$ M). In contrast, ERK1/2 and p38 MAP kinase activations were not influenced by Q3GA. These results suggested that JNK and Akt, but not extracellular signal-regulated kinase (ERK) 1/2 and p38 MAP kinase, were specifically sensitive to Q3GA in VSMCs. In this study, we used Q3GA at 1, 10, and 100  $\mu$ M. Previously, *in vitro* studies using cultured cells required relatively higher concentrations ( $>>\mu$ M) of the flavonoid metabolites to exert their anti-atherosclerotic effects (32, 33) as compared with the plasma concentrations (up to  $\mu$ M) reported in the human studies (14). These findings suggested that Q3GA specifically inhibited PDGF-induced JNK and Akt activations and resultant cell migration and proliferation in VSMCs.

Angiotensin II (Ang II) is a multifunctional peptide that has numerous actions on VSMCs — it modulates vasomotor tone, regulates cell growth and apoptosis, and influences cell migration and extracellular matrix deposition; it is proinflammatory; and it stimulates production of other growth factors and vasoconstrictors (34). In addition, Ang II has been shown to stimulate protein synthesis and induce cellular hypertrophy in VSMCs by acting through the G protein-coupled AT1 receptor (35). We previously reported that quercetin inhibited Ang II-induced VSMC hypertrophy through the inhibition of Src homology and collagen (Shc)- and PI3-kinase-mediated JNK activation (36). Furthermore, we examined the effect of chemically synthesized Q3GA, as an *in vivo* form, on Ang II-induced MAP kinase activation in VSMCs. Ang II-induced JNK activation was inhibited by Q3GA (10 and 100  $\mu$ M), whereas ERK1/2 and p38 activations were not affected (32). Experiments with

electron paramagnetic resonance revealed that Q3GA showed a scavenging activity for 1, 1-diphenyl-2-picrylhydrazyl radical in VSMCs (32). With respect to quercetin, it has been reported that quercetin inhibited NADPH-dependent oxidation in endothelial cells (37) and hydrogen peroxide generation in platelets (38). The *o*-dihydroxyl structure in the B-ring of quercetin, which Q3GA also possesses, is required to exert maximum free radical-scavenging activity (39, 40). Q3GA inhibited Ang II-induced increases in DNA binding activity of activator protein (AP)-1, a downstream transcription factor of JNK, composed of the c-Jun homo/heterodimer (32). In VSMC, AP-1 activation was suggested to be a critical component in the pathway leading to intimal thickness after balloon injury to the carotid artery of rats (41). Finally, Ang II-induced [ $^3$ H]leucine incorporation into RASMC was abolished by Q3GA (10  $\mu$ M) (32). These findings suggest that the preventing effect of Q3GA on Ang II-induced VSMC hypertrophy is attributable in part to its inhibitory effect on JNK and the AP-1 signaling pathway. Thus, inhibition of JNK by the *in vivo* form of quercetin may imply its usefulness for relief of cardiovascular diseases relevant to VSMC hypertrophy.

#### 4. Localization of Q3GA in the atherosclerotic aorta

The endothelial injury, activation, or dysfunction is an early event during the development of atherosclerosis (42). A previous study showed that the cholesterol accumulation in the aorta of hypercholesterolemic rabbits was decreased by orally administered quercetin glucoside (43). It was also demonstrated that quercetin metabolites were detected in the atherosclerotic aorta using high performance liquid chromatography analysis (43). Moreover, it was reported that Q3GA is capable of inhibiting LDL oxidation *in vitro* (44, 45). To examine the localization of Q3GA associated with the anti-atherosclerotic effects in the aorta, human aortic tissues were immunohistochemically examined using monoclonal antibody mA14A2, a novel monoclonal antibody targeting the Q3GA (33). This investigation was carried out on aortic wall samples obtained during autopsy from patients with generalized arteriosclerosis. Immunohistochemical studies with mA14A2 demonstrated that the positive staining specifically accumulates in human atherosclerotic lesions, but not in the normal aorta (33). Increased permeability of endothelial cells with reduced barrier function has been observed during endothelial injury (46). These observations suggest that Q3GA in circulating blood can permeate through the injured endothelial cells in human atherosclerotic lesions. Thus, our results suggest that the Q3GA is specifically localized in the arteries with ath-

erosclerotic lesions.

## 5. Conclusions

In conclusion, we demonstrated that Q3GA, an active metabolite of quercetin, specifically inhibited PDGF-induced JNK and Akt activations and resultant cell migration and proliferation in VSMCs. Also we clarified that the preventing effect of Q3GA on Ang II-induced VSMC hypertrophy is attributable in part to its inhibitory effect on JNK and the AP-1 signaling pathway. It is suggested that the inhibition of these pathways by Q3GA may imply its usefulness for cardiovascular diseases in which VSMC growth may be involved. Although most of the *in vitro* pharmacological studies have been carried out using only the quercetin aglycone form, experiments with the *in vivo* form of quercetin would be important to elucidate the efficacy of orally administered antioxidants including quercetin. Considering the above results, Q3GA may possess preventing effects for cardiovascular diseases relevant to VSMC disorders. These findings of our study may shed light on the pharmacological basis for oral administration of bioflavonoids in cardiovascular diseases.

## References

- Hollman PC, Katan MB. Health effects and bioavailability of dietary flavonols. *Free Radic Res.* 1999;31 Suppl:S75–S80.
- Renaud S, de Lorgeril M. Wine, alcohol, platelets, and the French paradox for coronary heart disease. *Lancet.* 1992;339:1523–1526.
- St Leger AS, Cochrane AL, Moore F. Factors associated with cardiac mortality in developed countries with particular reference to the consumption of wine. *Lancet.* 1979;1:1017–1020.
- Moore TJ, Conlin PR, Ard J, Svetkey LP. DASH (Dietary Approaches to Stop Hypertension) diet is effective treatment for stage 1 isolated systolic hypertension. *Hypertension.* 2001;38:155–158.
- Ogihara T, Kikuchi K, Matsuoka H, Fujita T, Higaki J, Horiuchi M, et al. The Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH 2009). *Hypertens Res.* 2009;32:3–107.
- Formica JV, Regelson W. Review of the biology of quercetin and related bioflavonoids. *Food Chem Toxicol.* 1995;33:1061–1080.
- Perez-Vizcaino F, Bishop-Bailley D, Lodi F, Duarte J, Cogolludo A, Moreno L, et al. The flavonoid quercetin induces apoptosis and inhibits JNK activation in intimal vascular smooth muscle cells. *Biochem Biophys Res Commun.* 2006;346:919–925.
- Bucki R, Pastore JJ, Giraud F, Sulpice JC, Janmey PA. Flavonoid inhibition of platelet procoagulant activity and phosphoinositide synthesis. *J Thromb Haemost.* 2003;1:1820–1828.
- Hanasaki Y, Ogawa S, Fukui S. The correlation between active oxygens scavenging and antioxidative effects of flavonoids. *Free Radic Biol Med.* 1994;16:845–850.
- Hayek T, Fuhrman B, Vaya J, Rosenblat M, Belinky P, Coleman R, et al. Reduced progression of atherosclerosis in apolipoprotein E-deficient mice following consumption of red wine, or its polyphenols quercetin or catechin, is associated with reduced susceptibility of LDL to oxidation and aggregation. *Arterioscler Thromb Vasc Biol.* 1997;17:2744–2752.
- Steinberg D, Parthasarathy S, Carew TE, Khoo JC, Witztum JL. Beyond cholesterol. Modifications of low-density lipoprotein that increase its atherogenicity. *N Engl J Med.* 1989;320:915–924.
- Tamura G, Gold C, Ferro-Luzzi A, Ames BN. Fecalase: a model for activation of dietary glycosides to mutagens by intestinal flora. *Proc Natl Acad Sci U S A.* 1980;77:4961–4965.
- Manach C, Morand C, Crespy V, Demigné C, Texier O, Régéat F, et al. Quercetin is recovered in human plasma as conjugated derivatives which retain antioxidant properties. *FEBS Lett.* 1998;426:331–336.
- Moon JH, Nakata R, Oshima S, Inakuma T, Terao J. Accumulation of quercetin conjugates in blood plasma after the short-term ingestion of onion by women. *Am J Physiol Regul Integr Comp Physiol.* 2000;279:R461–R467.
- Moon J, Tsushida T, Nakahara K, Terao J. Identification of quercetin 3-O- $\beta$ -D-glucuronide as an antioxidative metabolite in rat plasma after oral administration of quercetin. *Free Radic Biol Med.* 2001;30:1274–1285.
- Manach C, Texier O, Régéat F, Agullo G, Demigné C, Rémésy C. Dietary quercetin is recovered in rat plasma as conjugated derivatives of isorhamnetin and quercetin. *Nutr Biochem.* 1996;7:375–380.
- da Silva EL, Piskula MK, Yamamoto N, Moon JH, Terao J. Quercetin metabolites inhibit copper ion-induced lipid peroxidation in rat plasma. *FEBS Lett.* 1998;430:405–408.
- Ross R. Atherosclerosis – an inflammatory disease. *N Engl J Med.* 1999;340:115–126.
- Cunningham KS, Gotlieb AI. The role of shear stress in the pathogenesis of atherosclerosis. *Lab Invest.* 2005;85:9–23.
- Touyz RM. Intracellular mechanisms involved in vascular remodelling of resistance arteries in hypertension: role of angiotensin II. *Exp Physiol.* 2005;90:449–455.
- Mason DP, Kenagy RD, Hasenstab D, Bowen-Pope DF, Seifert RA, Coats S, et al. Matrix metalloproteinase-9 overexpression enhances vascular smooth muscle cell migration and alters remodeling in the injured rat carotid artery. *Circ Res.* 1999;85:1179–1185.
- Kishi K, Muramatsu M, Jin D, Furubayashi K, Takai S, Tamai H, et al. The effects of chymase on matrix metalloproteinase-2 activation in neointimal hyperplasia after balloon injury in dogs. *Hypertens Res.* 2007;30:77–83.
- Yamamoto Y, Ogino K, Igawa G, Matsuura T, Kaetsu Y, Sugihara S, et al. Allopurinol reduces neointimal hyperplasia in the carotid artery ligation model in spontaneously hypertensive rats. *Hypertens Res.* 2006;29:915–921.
- Grotendorst GR, Seppa HE, Kleinman HK, Martin GR. Attachment of smooth muscle cells to collagen and their migration toward platelet-derived growth factor. *Proc Natl Acad Sci U S A.* 1981;78:3669–3672.
- Nakagawa M, Ohno T, Maruyama R, Okubo M, Nagatsu A, Inoue M, et al. Sesquiterpene lactone suppresses vascular smooth muscle cell proliferation and migration via inhibition of cell cycle progression. *Biol Pharm Bull.* 2007;30:1754–1757.
- Izawa Y, Yoshizumi M, Ishizawa K, Fujita Y, Kondo S, Kagami



- S, et al. Big mitogen-activated protein kinase 1 (BMK1)/extracellular signal regulated kinase 5 (ERK5) is involved in platelet-derived growth factor (PDGF)-induced vascular smooth muscle cell migration. *Hypertens Res.* 2007;30:1107–1117.
- 27 Bilato C, Pauly RR, Melillo G, Monticone R, Gorelick-Feldman D, Gluzband YA, et al. Intracellular signaling pathways required for rat vascular smooth muscle cell migration. Interactions between basic fibroblast growth factor and platelet-derived growth factor. *J Clin Invest.* 1995;96:1905–1915.
- 28 Zhan Y, Kim S, Izumi Y, Izumiya Y, Nakao T, Miyazaki H, et al. Role of JNK, p38, and ERK in platelet-derived growth factor-induced vascular proliferation, migration, and gene expression. *Arterioscler Thromb Vasc Biol.* 2003;23:795–801.
- 29 Ishizawa K, Izawa Y, Ito H, Miki C, Miyata K, Fujita Y, et al. Aldosterone stimulates vascular smooth muscle cell proliferation via big mitogen-activated protein kinase 1 activation. *Hypertension.* 2005;46:1046–1052.
- 30 Motobayashi Y, Izawa-Ishizawa Y, Ishizawa K, Orino S, Yamaguchi K, Kawazoe K, et al. Adiponectin inhibits insulin-like growth factor-1-induced cell migration by the suppression of extracellular signal-regulated kinase 1/2 activation, but not Akt in vascular smooth muscle cells. *Hypertens Res.* 2009;32:188–193.
- 31 Ishizawa K, Izawa-Ishizawa Y, Ohnishi S, Motobayashi Y, Kawazoe K, Hamano S, et al. Quercetin glucuronide inhibits cell migration and proliferation by platelet-derived growth factor in vascular smooth muscle cells. *J Pharmacol Sci.* 2009;109:257–264.
- 32 Yoshizumi M, Tsuchiya K, Suzaki Y, Kirima K, Kyaw M, Moon JH, et al. Quercetin glucuronide prevents VSMC hypertrophy by angiotensin II via the inhibition of JNK and AP-1 signaling pathway. *Biochem Biophys Res Commun.* 2002;293:1458–1465.
- 33 Kawai Y, Nishikawa T, Shiba Y, Saito S, Murota K, Shibata N, et al. Macrophage as a target of quercetin glucuronides in human atherosclerotic arteries: implication in the anti-atherosclerotic mechanism of dietary flavonoids. *J Biol Chem.* 2008;283:9424–9434.
- 34 Touyz RM, Schiffrin EL. Signal transduction mechanisms mediating the physiological and pathophysiological actions of angiotensin II in vascular smooth muscle cells. *Pharmacol Rev.* 2000;52:639–672.
- 35 Griendling KK, Minieri CA, Ollerenshaw JD, Alexander RW. Angiotensin II stimulates NADH and NADPH oxidase activity in cultured vascular smooth muscle cells. *Circ Res.* 1994;74:1141–1148.
- 36 Yoshizumi M, Tsuchiya K, Kirima K, Kyaw M, Suzaki Y, Tamaki T. Quercetin inhibits Shc- and phosphatidylinositol 3-kinase-mediated c-Jun N-terminal kinase activation by angiotensin II in cultured rat aortic smooth muscle cells. *Mol Pharmacol.* 2001;60:656–665.
- 37 Holland JA, O'Donnell RW, Chang MM, Johnson DK, Ziegler LM. Endothelial cell oxidant production: effect of NADPH oxidase inhibitors. *Endothelium.* 2000;7:109–119.
- 38 Pignatelli P, Pulcinelli FM, Celestini A, Lenti L, Ghiselli A, Gazzaniga PP, et al. The flavonoids quercetin and catechin synergistically inhibit platelet function by antagonizing the intracellular production of hydrogen peroxide. *Am J Clin Nutr.* 2000;72:1150–1155.
- 39 Cao G, Sofic E, Prior RL. Antioxidant and prooxidant behavior of flavonoids: structure-activity relationships. *Free Radic Biol Med.* 1997;22:749–760.
- 40 van Acker SA, van den Berg DJ, Tromp MN, Griffioen DH, van Bennekom WP, van der Vijgh WJ, et al. Structural aspects of antioxidant activity of flavonoids. *Free Radic Biol Med.* 1996;20:331–342.
- 41 Kim S, Izumi Y, Yano M, Hamaguchi A, Miura K, Yamanaka S, et al. Angiotensin blockade inhibits activation of mitogen-activated protein kinases in rat balloon-injured artery. *Circulation.* 1998;97:1731–1737.
- 42 Ross R. The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature.* 1993;362:801–809.
- 43 Kamada C, da Silva EL, Ohnishi-Kameyama M, Moon JH, Terao J. Attenuation of lipid peroxidation and hyperlipidemia by quercetin glucoside in the aorta of high cholesterol-fed rabbit. *Free Radic Res.* 2005;39:185–194.
- 44 Moon JH, Tsushida T, Nakahara K, Terao J. Identification of quercetin 3-O-beta-D-glucuronide as an antioxidative metabolite in rat plasma after oral administration of quercetin. *Free Radic Biol Med.* 2001;30:1274–1285.
- 45 Terao J, Yamaguchi S, Shirai M, Miyoshi M, Moon JH, Oshima S, et al. Protection by quercetin and quercetin 3-O-beta-D-glucuronide of peroxynitrite-induced antioxidant consumption in human plasma low-density lipoprotein. *Free Radic Res.* 2001;35:925–931.
- 46 Yan S, Chai H, Wang H, Yang H, Nan B, Yao Q, et al. Effects of lysophosphatidylcholine on monolayer cell permeability of human coronary artery endothelial cells. *Surgery.* 2005;138:464–473.