

Full Paper

Changes in Synaptic Properties in Cortical-Limbic Communications Induced by Repeated Treatments With Fluvoxamine in Rats

Satoshi Ohashi, Hiroko Togashi, Machiko Matsumoto, Kiyoshi Mori, Ken-ichi Ueno, and Mitsuhiro Yoshioka*

Department of Pharmacology, Hokkaido University Graduate School of Medicine, Sapporo 060-8638, Japan

Received January 9, 2003; Accepted March 5, 2003

Abstract. There is evidence indicating that dysregulation of coordinated interactions of the cortical-limbic circuitry is associated with anxiety and mood disorders. Our previous study has reported that an enhancement of long-term plasticity in the “limbic-cortical” pathway produced by repeated treatments with fluvoxamine may be involved in the clinical effects of a selective serotonin (5-HT) reuptake inhibitor (SSRI). Here we assessed the effects of single and repeated treatments with fluvoxamine on the synaptic transmission and plasticity in the “cortical-limbic” pathway *in vivo*. The evoked potentials in the basolateral amygdaloid complex (BLA) by stimulation of the medial prefrontal cortex (mPFC) in halothane-anesthetized rats were recorded. Single administration of fluvoxamine (10 and 30 mg/kg, *i.p.*) enhanced the efficacy of synaptic transmission at the mPFC-BLA synapses dose-dependently. The enhanced synaptic efficacy induced by 30 mg/kg fluvoxamine was suppressed after long-term administration of fluvoxamine (30 mg/kg per day \times 21 days, orally). Repeated treatments with fluvoxamine affected short-term, but not long-term, synaptic plasticity in the mPFC-BLA pathway. These findings indicate that the 5-HTergic system contributes to modulation of synaptic changes in this pathway. Our results also suggest that different changes in synaptic properties in cortical-limbic communications induced by repeated treatments with fluvoxamine may be associated with therapeutic effects of SSRI.

Keywords: fluvoxamine, serotonin, medial prefrontal cortex-basolateral amygdaloid complex pathway, synaptic transmission, synaptic plasticity

Introduction

Many studies have suggested that abnormality of serotonin (5-HT) function is involved in the pathophysiology of psychiatric disorders. Selective 5-HT reuptake inhibitors (SSRIs) are effective in the treatment of anxiety and mood disorders in clinical studies (1, 2) as well as their animal models in preclinical experiments (3, 4). Although SSRIs inhibit 5-HT transporters on the neural membrane, resulting in a rapid increase in concentration of 5-HT within the synaptic clefts in terminal regions and cell bodies, the clinical effects of SSRIs appear after a few weeks. Converging evidence indicates that chronic SSRI treatment has therapeutic effects on anxiety and mood disorders via adaptive neurochemical

changes including pre- and post-synaptic regulatory desensitization, up- and down-regulation of various receptors, and receptor-mediated second messenger system and neurotrophic effects (5 – 7).

Recent neuroimaging studies have focused on the networks of various regions in the brain, suggesting that psychiatric patients use different patterns of functional connectivity in the cortical-limbic circuitry (8, 9). Reciprocal pathways linking limbic structures, such as the amygdala and hippocampus, with widely distributed brain stem, striatal, and cortical sites are now well defined; and they are clearly associated with specific cognitive, affective, and emotional behaviors in animals (10, 11). Dysregulation of the coordinated interactions of cortical-limbic communications is implicated in psychiatric illness (12). In particular, neuroimaging studies have found increased activation in the amygdala in anxiety and mood disorders (13 – 18). Furthermore,

*Corresponding author. FAX: +81-11-706-7872
E-mail: flute@med.hokudai.ac.jp

the prefrontal cortex (PFC) has abnormally decreased blood flow, metabolism, and volume in depressive patients (12, 19–21). From these findings, hyperfunction in the amygdala and hypofunction in the prefrontal cortical areas may reflect dysregulation in emotional and cognitive processing systems associated with psychiatric disorders. However, the precise interactions between the limbic structures and prefrontal cortical regions are unclear in affective and cognitive dysfunctions of anxiety and mood disorders. We previously showed that repeated treatments with fluvoxamine, an SSRI, produce an enhancement of long-term potentiation (LTP) in the “limbic-cortical” pathway, i.e., from the hippocampus to the medial PFC (mPFC) pathway, suggesting that an SSRI-induced enhancement of the limbic-cortical LTP could contribute to a therapeutic effect with cognitive recovery (22). In the present study, we focused on the “cortical-limbic” connection, namely the inputs of the amygdala received from the prefrontal cortical region. The basolateral complex of amygdala (BLA), which is composed of the lateral, basolateral, and basomedial nucleus, receives a projection from the mPFC. Anatomical and electrophysiological studies indicate that inputs from the infralimbic and prelimbic prefrontal areas terminate in the BLA (23–26).

Here we investigated the effects of single and repeated treatments with fluvoxamine on the synaptic transmission and plasticity in the rat mPFC-BLA pathway by using *in vivo* electrophysiological methods.

Materials and Methods

Male Wistar rats (230–290 g) were purchased from Shizuoka Laboratory Animal Center (Hamamatsu) and were housed at an ambient temperature of 22°C under a 12 h–12 h light-dark cycle (lights on, 19:00 h) with free access to food and water. All procedures were performed in accordance with the Guidelines for the Care and Use of Laboratory Animals of the Animal Research Committee of the Hokkaido University School of Medicine.

Single and repeated treatments with fluvoxamine were given according to our previous study (22). Doses of 10 and 30 mg/kg of fluvoxamine dissolved in saline were given intraperitoneally in the single-injection group. In the repeated-treatment group, fluvoxamine (30 mg/kg), dissolved in deionized water when used, was administered orally once a day for 21 days. All administrations were performed between 09:00 and 11:00 h. On the 22nd day, rats subjected to repeat treatments were systemically injected at a dose of 30 mg/kg of fluvoxamine dissolved in saline.

Rats were anesthetized with 1% halothane in 21% O₂

and 79% N₂ mixture, which was administered through a tracheal cannula, and they were fixed in a stereotaxic frame with the bregma and lambda in the same horizontal plane. Halothane used as an anesthesia enabled us to compare this study with our previous study (22). Throughout the experiment, the blood pressure and the heart rate were monitored and body temperature was maintained at 37°C by using a thermoregulated heating pad. A stimulating electrode with a tip separation of 500 μ m was used to stimulate the prelimbic and infralimbic regions of the PFC (coordinates: 2.7 mm anterior to the bregma, 0.7 mm lateral, 3.6–4.3 mm ventral from the cortical surface) according to the atlas of Paxinos and Watson (27). A recording electrode (100 μ m diameter, stainless steel) was placed in the ipsilateral BLA (coordinates: 3.3 mm posterior to the bregma, 5.2 mm lateral, 6.0–7.3 mm ventral from the cortical surface). The field potentials evoked by mPFC stimulation were amplified and were monitored with an oscilloscope (VC-10; Nihon Koden, Tokyo); they could be divided into four primary components: two positive and two negative components, named P1, P2, N1, and N2, respectively (26) (Fig. 2A). We focused on the N1 and P2 components in the present study. These components were used as an index of changes in the mPFC-BLA afferent drive (26). The integrated responses were averaged by using a data analyzing system (MASSCOMP; Concurrent, Tokyo) and the amplitude of the N1 and P2 field potentials was measured.

After inserting the electrode, the input/output characterization of each individual rat was determined for the mPFC-BLA pathway by varying the stimulus intensity from 200 μ A to 1200 μ A stepwise. A test stimulus was then chosen to give an approximately 60% maximal response.

We used a paired-pulse ratio (PPR) protocol to investigate the effects of repeated treatments with fluvoxamine on short-term synaptic plasticity in some saline- and repeated-treatment rats after a test stimulus had been chosen. For the paired-pulse experiments, two stimuli were applied at interstimulus intervals (ISIs) ranging from 50 to 150 ms, in steps of 50 ms. The PPR was expressed as the value of S2/S1, where S2 was the amplitude of the second response and S1 was the amplitude of the first one. Here we assessed the negative component N1 for PPR.

The evoked potentials were expressed as percentages of the baseline level determined immediately before the administration of the drug to the rats. Throughout the experiment, test stimuli were delivered every 30 s and nine successive responses were averaged and collected every 5 min. To produce the LTP in the BLA by stimulation of the prelimbic and infralimbic regions of the

prefrontal areas, two series of 10 high-frequency stimulations (250 Hz, 250- μ s duration, 50 trains) at 0.1 Hz were applied 40 min after intraperitoneal injection of the drug or saline.

At the last of the electrophysiological recording sessions, a positive current (70 μ A, 10 s) was passed through a recording electrode from which the evoked potentials were recorded to deposit iron ions. Immediately after the experiments, the rats were perfused through the left ventricle with 300 ml of 4% formaldehyde in 0.1 M phosphate buffer (pH 7.2) containing 5% potassium ferrocyanide to produce a Prussian blue reaction at the iron deposits. The brains were removed and were postfixed in 4% formaldehyde in 0.1 M phosphate buffer (pH 7.2) for at least two days. Sections (50 μ m) were cut through the recording and stimulating sites on a microslicer, and they were then mounted on gelatin-coated slides. After drying, the slides were stained with Neutral Red, and the recording and stimulating sites were evaluated under a microscope.

All results are given as the mean \pm S.E.M. Data analysis used analysis of variance (ANOVA) followed by the two-tailed post-hoc Dunnett's multiple comparison procedure or Student's *t*-test as appropriate. The statistical significance was $P < 0.05$.

Results

Effects of single and repeated treatments with fluvoxamine on synaptic transmission in the mPFC-BLA pathway

As shown in Fig. 1, the recording electrodes were positioned in the lateral and basolateral regions of the BLA. The stimulating electrodes were located in the prelimbic and infralimbic regions of the mPFC.

We determined input/output curves for each group. The increase in evoked potentials depended on the stimulation intensity. The test stimulation intensities of the saline-injected group ($466.7 \pm 120.2 \mu$ A), the single-treatment group ($523.3 \pm 89.6 \mu$ A), and the repeated-treatment group ($540.0 \pm 42.3 \mu$ A) showed no significant difference.

We examined the effect of a single administration of fluvoxamine on the efficacy of synaptic transmission in the mPFC-BLA pathway. The synaptic efficacy was evaluated by monitoring the potentials evoked in the BLA by stimulating the prelimbic and infralimbic regions of the prefrontal cortical areas. After the systemic administration of fluvoxamine (10 and 30 mg/kg, $n = 5$ and 6, respectively), field potentials (both N1 and P2 peaks) evoked by test stimulation of the mPFC were enhanced in a dose-dependent manner (Fig. 2: A, C). As shown in Fig. 2B, the enhancement of the synaptic

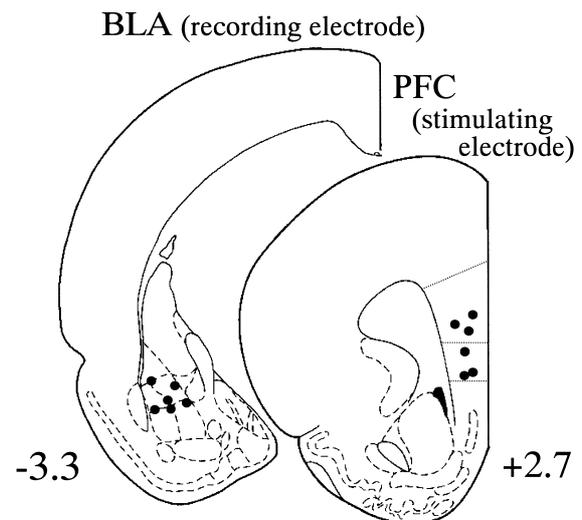


Fig. 1. Histology of the BLA and mPFC after electrophysiological experiments. Black dots show the representative locations of the recording electrode in the BLA (-3.3 mm from the bregma) and the stimulating electrode in the prelimbic and infralimbic areas of the PFC ($+2.7$ mm from the bregma).

efficacy induced by a high dose of fluvoxamine (30 mg/kg) was statistically significant by the area under the curve (AUC) analysis for 80 min after the drug injection when compared with the saline-injected group ($n = 7$, N1, Dunnett's test, $P < 0.001$). AUC analysis for P2 showed a similar significant enhancement of the synaptic efficacy in the mPFC-BLA pathway (Fig. 2D, Dunnett's test, $P < 0.05$ at 10 mg/kg and $P < 0.0005$ at 30 mg/kg).

We next evaluated the effect of repeated treatments with fluvoxamine on the synaptic efficacy in this pathway. Fluvoxamine (30 mg/kg per day) dissolved in distilled water were repeatedly administered orally for 21 days. On the following day, repeated-treatment rats were intraperitoneally given the same dose of drug. Repeated treatments with fluvoxamine ($n = 7$) significantly suppressed the enhancement of synaptic efficacy induced by a high dose of single administration (Fig. 3: A, C). The suppression of synaptic efficacy produced by long-term administration of fluvoxamine (30 mg/kg) was statistically significant by AUC analysis for 80 min after the drug injection when compared with the single-treatment group (Fig. 3: B, D, $n = 6$, Student's *t*-test, $P < 0.01$ for N1 and $P < 0.05$ for P2).

Effects of single and repeated treatments with fluvoxamine on mPFC-BLA synaptic plasticity

We tested the repeated fluvoxamine treatment effect on short- and long-term synaptic plasticity in the mPFC-BLA pathway. First, paired-pulse experiments were

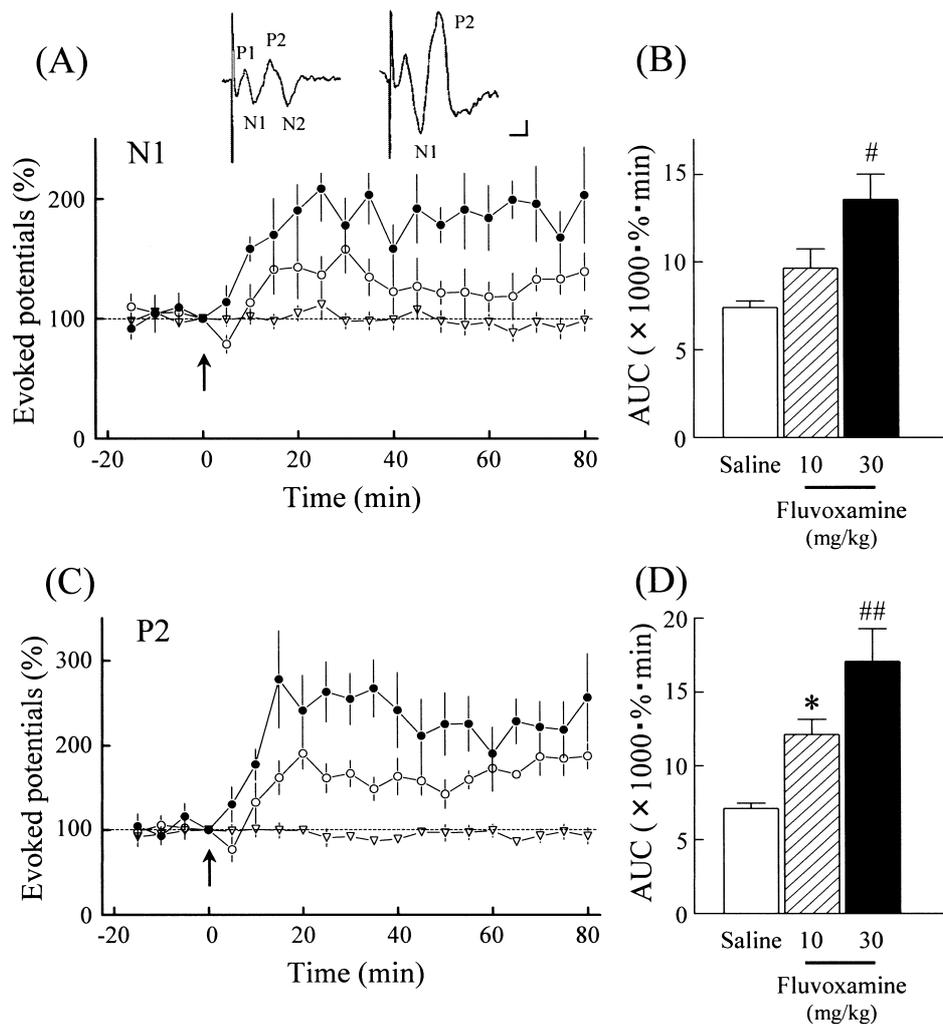


Fig. 2. Effects of a single administration of fluvoxamine on the potentials evoked in the BLA by stimulation of the prelimbic and infralimbic regions of the PFC in anesthetized rats. A, C: Time course responses of evoked potentials after single administration of fluvoxamine (10 and 30 mg/kg, $n = 5$ and 7 , respectively) and saline ($n = 6$). Averaged waveforms at the top of the graph show the evoked potentials immediately before (left) and 80 min after (right) fluvoxamine (30 mg/kg) injection. The evoked potentials are expressed as the percentage of the baseline value before saline or fluvoxamine administration. Calibration: $50 \mu\text{V}$, 20 ms. B, D: AUC of the facilitation of synaptic efficacy in the mPFC-BLA pathway induced by fluvoxamine (10 and 30 mg/kg) dose-dependently. The arrow indicates the injection of the drug or saline. Values are the mean \pm S.E.M. Open triangle, saline-injected group; open circle, 10 mg/kg fluvoxamine group; closed circle, 30 mg/kg fluvoxamine group. * $P < 0.05$, # $P < 0.001$, and ## $P < 0.0005$, compared with the saline-injected group.

conducted to examine the effects of repeated treatments with fluvoxamine on short-term plasticity. ISIs ranged from 50 to 150 ms, in steps of 50 ms. The rats treated repeatedly with fluvoxamine showed a significant suppression of the PPRs at each ISI compared with the saline-injected group (Fig. 4). The PPRs were 1.53 ± 0.17 and 1.04 ± 0.09 at 50 ms ($P < 0.05$), 2.25 ± 0.34 , and 1.32 ± 0.11 at 100 ms ($P < 0.05$) and 1.71 ± 0.09 and 1.27 ± 0.06 at 150 ms ($P < 0.005$, Student's t -test, the saline-injected group and the repeated-treatment group, $n = 4$ and 6 , respectively). Second, two series of high-frequency stimulations of the mPFC region 40 min after

the administration of fluvoxamine (30 mg/kg) induced long-lasting increases in evoked potentials; i.e., LTP establishment in the saline-, single-, and repeated-treatment groups. Single and repeated treatments with fluvoxamine had no effect on the LTP, measured by AUC for 60 min after high-frequency stimulation of the mPFC region, in the BLA (data not shown).

Discussion

The results of this study show that single and repeated treatments with an SSRI can change synaptic properties

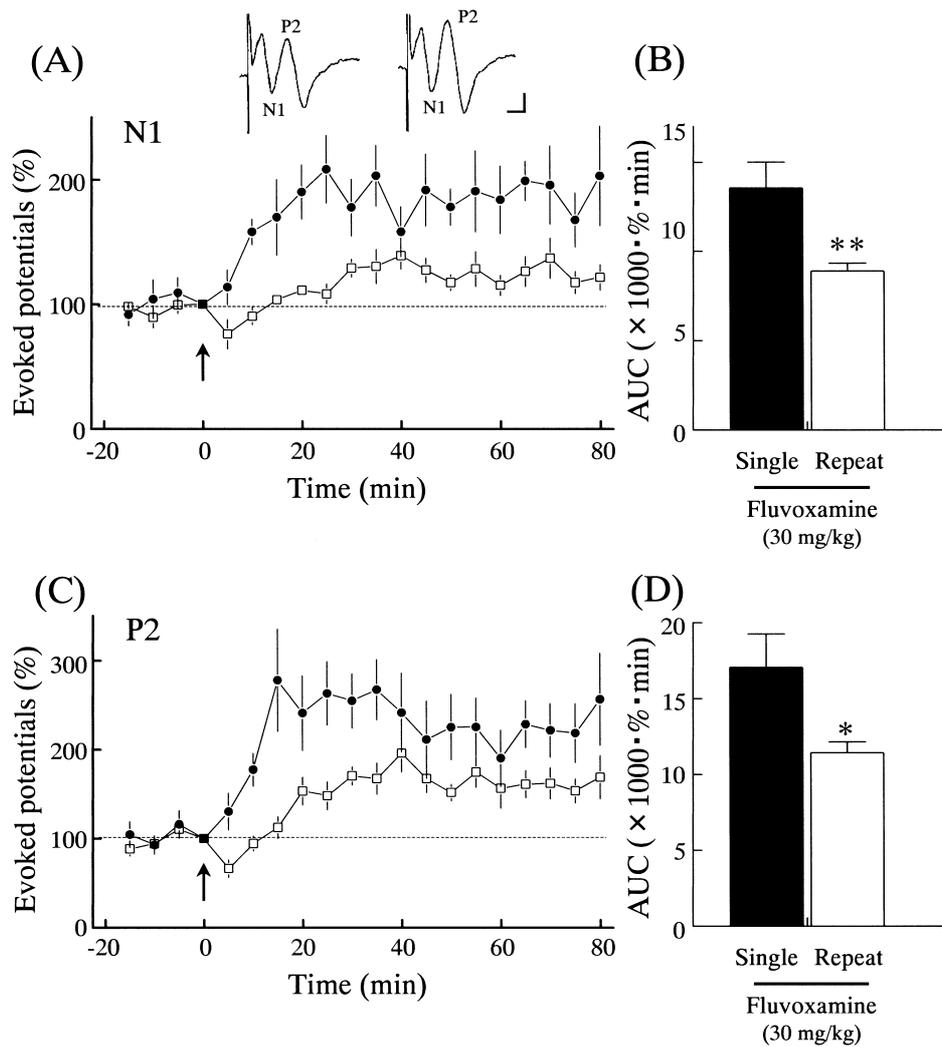


Fig. 3. Effects of single and repeated treatments with fluvoxamine (30 mg/kg) on the efficacy of synaptic transmission in the mPFC-BLA pathway. A, C: Time course responses of the evoked potentials after single treatment (n = 6) and repeated treatments with fluvoxamine (n = 7). Averaged waveforms at the top of the graph show the evoked potentials immediately before (left) and 80 min after (right) of the repeated-treatment rat. The evoked potentials are expressed as the percentage of the baseline value before fluvoxamine administration. Calibration: 50 μV , 20 ms. B, D: AUC of repeated treatments with fluvoxamine depressed the synaptic efficacy in the mPFC-BLA pathway. The arrow indicates the injection of the drug. Values are the mean \pm S.E.M. Closed circle, single fluvoxamine group; open square, repeated fluvoxamine group. * $P < 0.05$ and ** $P < 0.01$, compared with the single-treatment group.

in the cortical-limbic pathway. First, single treatment with fluvoxamine produced an enhancement of the efficacy of synaptic transmission at the mPFC-BLA synapses dose-dependently. Second, an enhancement of synaptic efficacy by a high dose of a single administration was suppressed after long-term treatment with fluvoxamine. Finally, repeated treatments with fluvoxamine modulated the short-term, but not long-term, synaptic plasticity in the mPFC-BLA pathway. These findings suggest that cortical-limbic synaptic changes are induced by long-term administration of fluvoxamine in the therapeutic effects of SSRI.

Single administration of fluvoxamine enhanced the efficacy of synaptic transmission in the mPFC-BLA pathway dose-dependently, suggesting that endogenous 5-HT can modulate the synaptic transmission via certain 5-HT receptors on the postsynaptic membrane. Administration of fluvoxamine, an SSRI, increases 5-HT levels within the synaptic clefts by inhibiting 5-HT transporters on the neuronal membrane. Our experiment showed that a single systemic administration of fluvoxamine increased the extracellular levels of 5-HT in the amygdala by using a microdialysis technique (28). Thus, the present findings suggest that increases in endogenous

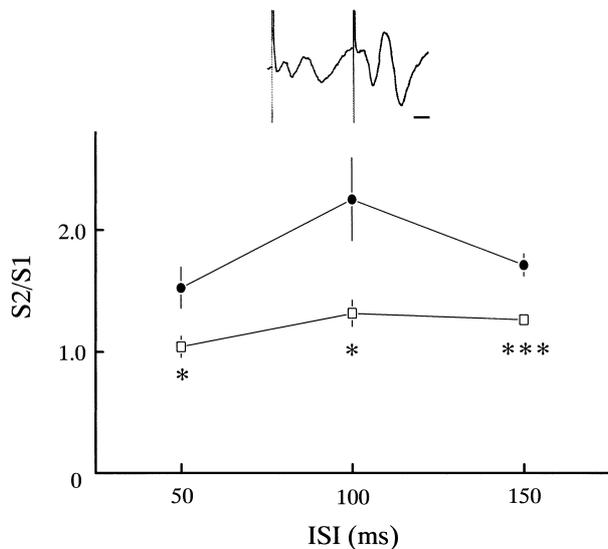


Fig. 4. Paired-pulse facilitation in saline- and repeated-treatment rats as a function of the ISI ($n = 4$ and 6 , respectively). Suppression of paired-pulse facilitation in the BLA was induced by long-term administration of fluvoxamine. Averaged waveforms at the top of the graph show the evoked potentials at 100 ms ISI of the saline-injected rat. Calibration: 20 ms. The second response (S2) is expressed as a ratio the first response (S1). Values are the mean \pm S.E.M. Closed circle, saline-injected group; open square, repeated fluvoxamine group. * $P < 0.05$ and *** $P < 0.005$ represent significant differences.

5-HT concentration produced by fluvoxamine may participate in the enhancement of synaptic efficacy in the mPFC-BLA pathway. Systemic administration of 30 mg/kg of fluvoxamine enhanced the synaptic efficacy in this “cortical-limbic” pathway to 212% and 303% of basal synaptic transmission (N1 and P2, respectively). These enhancements are greater than in the “limbic-cortical” pathway. Our previous experiments in the hippocampo-mPFC pathway showed that the same dose of 30 mg/kg of fluvoxamine enhanced the synaptic efficacy to 158% of basal transmission (22), which may reflect that increases in 5-HT levels in the frontal cortical area induced by SSRIs are smaller than in other brain regions (29). The data shown here provide evidence that the efficacy of synaptic transmission in the mPFC-BLA pathway can be modulated by the 5-HTergic system as well as the dopaminergic system (26). Although the critical role of the mPFC-BLA pathway in emotional and cognitive processes remains elusive, we obtained intriguing results that synaptic transmission in the mPFC-BLA pathway decreased during contextual fear conditioning, as an index of anxiety, in freely moving rats. Our experiments also showed that the anxiolytic agent, tandospirone (a 5-HT_{1A}-receptor partial agonist) produced an increase in synaptic transmission in this pathway (unpublished data). These findings suggest that

changes in the synaptic efficacy in the mPFC-BLA pathway have an important role in anxiety-related behavior. It has, furthermore, suggested that the plastic change within the amygdala contributes to the formation of fear-related memory (30–32).

The enhancement of synaptic transmission in the mPFC-BLA pathway induced by a single treatment with fluvoxamine (30 mg/kg) was significantly suppressed by repeated treatments for 3 weeks. Indeed, clinical studies have shown that fully developed therapeutic efficacy of SSRIs requires 2 to 4 weeks. We also observed that repeated administrations of fluvoxamine suppressed the short-term plasticity (i.e., paired-pulse facilitation) in the mPFC-BLA pathway. This may reflect changes in transmitter release probability at the mPFC-BLA synapses, since paired-pulse facilitation has primarily been attributed to short-term glutamate release and has been used as a tool to implicate presynaptic participation (33). These changes in short-term plasticity also suggest that decreases in glutamate release at the presynapse in the mPFC-BLA pathway were caused by repeated treatments with fluvoxamine. Antidepressant drugs appear to reduce glutamatergic activity or glutamate receptor-related signal transduction (34). Thus, the suppression of paired-pulse facilitation by repeated treatments with fluvoxamine in the present study should decrease the excitability into the amygdala. This decreased excitability may contribute to the suppression of synaptic efficacy compared with a single treatment. Short-term synaptic plasticity, i.e., paired-pulse facilitation, has an important role in behavioral information processing in the hippocampal structures (35, 36). A more recent study also showed that short-term synaptic plasticity might link with the same as behavioral information processing in the “limbic-cortical” pathway; i.e., the hippocampo-mPFC pathway (37). Although further studies are needed to clarify the physiological functions of synaptic properties in the mPFC-BLA pathway, the findings in the present study suggest that repeated treatments with fluvoxamine suppressed an increased flow of excitatory activity into the amygdala, resulting in the therapeutic effects of SSRIs on anxiety and mood disorders. The results obtained here, furthermore, indicate that not only synaptic transmission, but also synaptic plasticity, could be influenced by repeated treatments with SSRI.

Several lines of evidence demonstrated that chronic antidepressive treatments could change the various synaptic properties in the rat hippocampal formation (38–41). We previously reported that long-term treatment with fluvoxamine enhanced the establishment of LTP in the hippocampo-mPFC pathway; i.e., the “limbic-cortical” projection (22). This pathway participates in asso-

ciative learning in animal behavioral tests (42, 43). Hypofunction of the prefrontal regions occurs in depressive patients with cognitive dysfunction. The enhancement of LTP seen at the hippocampal-mPFC synapses (i.e., the “limbic-cortical” pathway) may be associated with improvement in cognitive function, resulting in recovery from depressive states. However, repeated treatments with fluvoxamine produced suppression of synaptic transmission and short-term synaptic plasticity in the mPFC-BLA pathway (i.e., the “cortical-limbic” projection) in the present study, suggesting that decreases in information flows from the cortical areas to the limbic regions may be implicated in improvement of anxiety and mood disorders. These findings suggest that the different changes in synaptic properties in cortical-limbic communications induced by long-term administration of fluvoxamine may be involved in the therapeutic effects of SSRI in psychiatric disorders. In other words, our data that enhanced “limbic-cortical” LTP and suppressed “cortical-limbic” synaptic properties induced by repeated treatments with fluvoxamine are consistent with evidence by clinical studies of the effects of chronic antidepressants on both frontal hypofunction and limbic hyperfunction (12, 21, 44 – 47).

Acknowledgments

The authors would like to thank Meiji Seika Kaisha (Tokyo) for the gift of fluvoxamine. We are grateful to Ms. Asako Kaku for her technical support.

References

- 1 Sheehan DV, Raj BA, Trehan RR, Knapp EL. Serotonin in panic disorder and social phobia. *Int Clin Psychopharmacol.* 1993;8:63–77.
- 2 Lane R, Baldwin D, Preksorn S. The selective serotonin reuptake inhibitors: advantage, disadvantages and differences. *J Psychopharmacol.* 1995;9:163–178.
- 3 Ichimaru Y, Egawa T, Sawa A. 5-HT_{1A}-receptor subtype mediates the effect of fluvoxamine, a selective serotonin reuptake inhibitor, on marble-burying behavior in mice. *Jpn J Pharmacol.* 1995;68:65–70.
- 4 Egawa T, Ichimaru Y, Imanishi T, Sawa A. Neither the 5-HT_{1A}-nor the 5-HT₂-receptor subtype mediates the effect of fluvoxamine, a selective serotonin reuptake inhibitor, on forced-swimming-induced immobility in mice. *Jpn J Pharmacol.* 1995;68:71–75.
- 5 Duman RS, Heninger GR, Nestler EJ. A molecular and cellular theory of depression. *Arch Gen Psychiatry.* 1997;54:597–606.
- 6 Blier P, de Montigny C. Serotonin and drug-induced therapeutic responses in major depression, obsessive-compulsive and panic disorders. *Neuropsychopharmacology.* 1999;21:91S–98S.
- 7 Duman RS, Malberg J, Thome J. Neural plasticity to stress and antidepressant treatment. *Biol Psychiatry.* 1999;46:1181–1191.
- 8 Davidson RJ, Irwin W. The functional neuroanatomy of emotion and affective style. *Trends Cogn Neurosci.* 1999;3:11–21.
- 9 Drevets WC. Neuroimaging studies of mood disorders. *Biol Psychiatry.* 2000;48:813–829.
- 10 Carmichael ST, Price JL. Limbic connections of the orbital and medial prefrontal cortex in macaque monkeys. *J Comp Neurol.* 1995;363:615–641.
- 11 Dias R, Robbins TW, Roberts AC. Dissociation in prefrontal cortex of affective and attentional shifts. *Nature.* 1996;380:69–72.
- 12 Mayberg HS. Limbic-cortical dysregulation: a proposed model of depression. *J Neuropsychiatry Clin Neurosci.* 1997;9:471–481.
- 13 Drevets WC, Videen TO, Price JL, Preskorn SH, Carmichael ST, Raichle ME. A functional anatomical study of unipolar depression. *J Neurosci.* 1992;12:3628–3641.
- 14 Abercrombie HC, Schaefer SM, Larson CL, et al. Metabolic rate in the right amygdala predicts negative affect in depressed patients. *Neuroreport.* 1998;9:3301–3307.
- 15 Schneider F, Weiss U, Kessler C, et al. Subcortical correlates of differential classical conditioning of aversive emotional reactions in social phobia. *Biol Psychiatry.* 1999;45:863–871.
- 16 Rauch SL, Whalen PJ, Shin LM, et al. Exaggerated amygdala response to masked facial stimuli in posttraumatic stress disorder: a functional MRI study. *Biol Psychiatry.* 2000;47:769–776.
- 17 Sheline YI, Barch DM, Donnelly JM, Ollinger JM, Snyder AZ, Mintun MA. Increased amygdala response to masked emotional faces in depressed subjects resolves with antidepressant treatment: an fMRI study. *Biol Psychiatry.* 2001;50:651–658.
- 18 Thomas KM, Drevets WC, Dahl RE, et al. Amygdala response to fearful faces in anxious and depressed children. *Arch Gen Psychiatry.* 2001;58:1057–1063.
- 19 Dolan RJ, Bench CJ, Brown RG, Scott LC, Friston KJ, Frackowiak RSJ. Regional cerebral blood flow abnormalities in depressed patients with cognitive impairment. *J Neurol Neurosurg Psychiatry.* 1992;55:768–773.
- 20 Dolan RJ, Bench CJ, Brown RG, Scott LC, Frackowiak RSJ. Neuropsychological dysfunction in depression: the relationship to regional cerebral blood flow. *Psychol Med.* 1994;24:849–857.
- 21 Mayberg HS, Liotti M, Brannan SK, et al. Reciprocal limbic-cortical function and negative mood: converging PET findings in depression and normal sadness. *Am J Psychiatry.* 1999;156:675–682.
- 22 Ohashi S, Matsumoto M, Otani H, et al. Changes in synaptic plasticity in the rat hippocampo-medial prefrontal cortex pathway induced by repeated treatments with fluvoxamine. *Brain Res.* 2002;949:131–138.
- 23 Ottersen OP. Connections of the amygdala of the rat. IV: Corticoamygdaloid and intraamygdaloid connections as studied with axonal transport of horseradish peroxidase. *J Comp Neurol.* 1982;205:30–48.
- 24 McDonald AJ, Mascagni F, Guo L. Projections of the medial and lateral prefrontal cortices to the amygdala: a *Phaseolus vulgaris* leucoagglutinin study in the rat. *Neuroscience.* 1996;71:55–75.
- 25 Pitkänen A, Savander V, LeDoux JE. Organization of intra-amygdaloid circuitries in the rat: an emerging framework for understanding functions of the amygdala. *Trends Neurosci.* 1997;20:517–523.
- 26 Rosenkranz JA, Grace AA. Modulation of basolateral amygdala

- neuronal firing and afferent drive by dopamine receptor activation in vivo. *J Neurosci*. 1999;19:11027–11039.
- 27 Paxinos G, Watson C. *The rat brain in stereotaxic coordinates*. 2nd ed. New York: Academic Press; 1986.
- 28 Ohashi S, Matsumoto M, Togashi H, et al. SSRI-induced changes in synaptic efficacy in the cortico-limbic systems in rats. *Soc Neurosci Abstr*. 2001;27:275.
- 29 Fuller RW. Uptake inhibitors increase extracellular serotonin concentration measured by brain microdialysis. *Life Sci*. 1994;55:163–167.
- 30 LeDoux JE. Emotion circuits in the brain. *Annu Rev Neurosci*. 2000;23:155–184.
- 31 Schafe GE, Nader K, Blair HT, LeDoux JE. Memory consolidation of Pavlovian fear conditioning: a cellular and molecular perspective. *Trends Neurosci*. 2001;24:540–546.
- 32 Medina JF, Christopher RJ, Mauk MD, LeDoux JE. Parallels between cerebellum- and amygdala-dependent conditioning. *Nat Rev Neurosci*. 2002;3:122–131.
- 33 Zucker RS. Short-term synaptic plasticity. *Annu Rev Neurosci*. 1989;12:13–31.
- 34 Krystal JH, Sanacora G, Blumberg H, et al. Glutamate and GABA systems as targets for novel antidepressant and mood-stabilizing treatments. *Mol Psychiatry*. 2002;7:S71–S81.
- 35 Cao F, Leung LS. Behavior-dependent paired-pulse responses in the hippocampal CA1 region. *Exp Brain Res*. 1991;87:553–561.
- 36 Silva AJ, Rosahl TW, Chapman PF, et al. Impaired learning in mice with abnormal short-lived plasticity. *Curr Biol*. 1996;6:1509–1518.
- 37 Izaki Y, Takita M, Nomura M. Local properties of CA1 region in hippocampo-prefrontal synaptic plasticity in rats. *Neuroreport*. 2002;13:469–472.
- 38 Bijak M, Tokarski K, Maj J. Repeated treatment with antidepressant drugs induces subsensitivity to the excitatory effect of 5-HT₄ receptor activation in the rat hippocampus. *Naunyn Schmiedeberg's Arch Pharmacol*. 1997;355:14–19.
- 39 Stewart CA, Reid IC. Repeated ECS and fluoxetine administration have equivalent effects on hippocampal synaptic plasticity. *Psychopharmacology (Berl)*. 2000;148:217–223.
- 40 Levkovitz Y, Grisaru N, Segal M. Transcranial magnetic stimulation and antidepressive drugs share similar cellular effects in rat hippocampus. *Neuropsychopharmacology*. 2001;24:608–616.
- 41 Zahorodna A, Tokarski K, Bijak M. Imipramine but not 5-HT_{1A} receptor agonists or neuroleptics induces adaptive changes in hippocampal 5-HT_{1A} and 5-HT₄ receptors. *Eur J Pharmacol*. 2002;443:51–57.
- 42 Doyère V, Burette F, Negro CR-D, Laroche S. Long-term potentiation of hippocampal afferents and efferents to prefrontal cortex: implications for associative learning. *Neuropsychologia*. 1993;31:1031–1053.
- 43 Laroche S, Davis S, Jay TM. Plasticity at hippocampal to prefrontal cortex synapses: dual roles in working memory and consolidation. *Hippocampus*. 2000;10:438–446.
- 44 Bench CJ, Frackowiak RSJ, Dolan RJ. Changes in regional cerebral blood flow on recovery from depression. *Psychol Med*. 1995;25:247–251.
- 45 Buchsbaum MS, Wu J, Siegel BV, et al. Effect of sertraline on regional metabolic rate in patients with affective disorders. *Biol Psychiatry*. 1997;41:15–22.
- 46 Mayberg HS, Brannan SK, Tekell JL, et al. Regional metabolic effects of fluoxetine in major depression: serial changes and relationship to clinical response. *Biol Psychiatry*. 2000;48:830–843.
- 47 Kennedy SH, Evans KR, Krüger S, et al. Changes in regional brain glucose metabolism measured with positron tomography after paroxetine treatment of major depression. *Am J Psychiatry*. 2001;158:899–905.