



Cyanobacterial blooms in natural waters in southern Portugal: a water management perspective

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ABSTRACT: This synthesis of 3 studies from 2 regions of southern Portugal (Alentejo and Algarve) was part of a workshop focusing on cyanobacteria held at the SAME 10. The first study monitored impacts of the large Alqueva dam on the Guadiana estuary since 1996, revealing changes in sediment load, nutrient regime and phytoplankton succession. Prior to dam construction, dense cyanobacterial blooms occurred in the upper estuary during summer and fall. After dam construction, chlorophyll concentration, phytoplankton diversity and abundances of cyanobacteria decreased, contrary to predictions. Microcystins remained at low levels in the seston and undetectable in water samples, except during summer 2003 when the particulate fraction contained $1 \mu\text{g l}^{-1}$, while chlorophyll concentrations and abundances of potentially toxic cyanobacteria remained low. Algarve reservoirs studied since 2001 revealed differences in phytoplankton dynamics. In the western mesotrophic reservoirs (Bravura and Funcho), 40 to 50 % of surface samples contained cyanobacterial concentrations of $\geq 2000 \text{ cells ml}^{-1}$, while over 80 % of samples from the eastern oligotrophic reservoirs (Odeleite and Beliche) exceeded this value. Spring blooms were dominated by *Oscillatoriales* in Odeleite and Beliche and by *Chroococcales* in Bravura and Funcho. Bloom composition seemed to depend on water temperature and management strategies, while toxin concentrations reflected the increased biomass of toxic species. Finally, phytoplankton communities and microcystin production in 5 Alentejo freshwater reservoirs were studied from May to December 2005 and April to July 2006. Cyanobacterial blooms occurred, with varying intensities, not only during summer but also occasionally in winter. Microcystins were detected in 23 % of the samples ($n = 51$), but without correlation with cyanobacterial biomass. Although *Microcystis aeruginosa* seemed to be the major producer of microcystins, other potentially toxic species were found. In summary, the varying pattern of cyanobacterial bloom occurrence and toxicity requires a systematic approach to monitoring programs for adequate risk assessment.

KEY WORDS: Cyanobacteria · Microcystins · Phytoplankton · Estuary · Freshwater reservoirs

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INTRODUCTION

Shifts in the relative abundance of microalgae within phytoplankton communities define seasonal successions of organisms belonging to different taxa. Phytoplankton successions in natural waters of temperate regions have been well documented for decades

(Hutchinson 1967, Smetacek 1985, Cloern 1996). While a great variety of eukaryotic microalgae and cyanobacteria usually coexist in the same water body in late spring and early summer, this diversity may drop drastically towards the end of summer, giving way to the mass development of cyanobacterial populations (Hutchinson 1961, Rocha et al. 2002).

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Although this pattern of seasonal succession can be assumed to be a general trend, phytoplankton dynamics are affected by a more complex scenario of internal and external driving factors, in which different species interact not only with each other but also with the environment, rendering any attempt at prediction or modeling particularly challenging. In the last few decades, a lot of effort has been invested in discerning which environmental factors control bloom formation, especially with respect to toxic species such as cyanobacteria. Understanding the conditions that trigger toxic blooms of cyanobacteria in lakes, reservoirs and rivers is a complex quest, hindered by the lack of information on natural ecological relationships between plankton populations.

Blooms of cyanobacteria have been repeatedly associated with eutrophication processes (Berg et al. 1987, Carmichael et al. 1988, Druvietis 1997, Pinckney et al. 1998, Codd 2000, Chorus 2005). Factors such as high water retention time in lakes and reservoirs, increased temperature, low N:P ratio, as well as surface radiation and wind conditions have been reported to influence bloom development (Carmichael 1996, Kononen et al. 1996, Kawara et al. 1998, Chorus & Bartram 1999). Most species preferentially occur in stable, well stratified water bodies that provide optimal conditions for their vertical migration. Species like *Microcystis aeruginosa* and *Anabaena* spp. contain intracellular gas vesicles and become buoyant, accumulating on the water surface where they can be blown around by the wind, forming very dense stable scums that are easily detected by simple observation of the water reservoir from above (Dittmann & Wiegand 2006). In contrast, other species like *Planktothrix* spp. and *Oscillatoria* spp. are more sensitive to high light conditions and usually tend to accumulate in deeper water layers or disperse homogeneously in the water column (Dittmann & Wiegand 2006).

Cyanobacterial blooms can be harmful to humans due to the ability of several bloom-forming species to synthesize highly toxic secondary metabolites: cyanotoxins (Dittmann & Wiegand 2006). Hepatotoxic microcystins are the most commonly found and widespread cyanotoxins. They are associated with several bloom-forming genera of cyanobacteria, including *Microcystis*, *Anabaena*, *Anabaenopsis*, *Planktothrix*, *Nostoc*, *Hapalosiphon*, *Snowella*, *Woronichinia*, and *Oscillatoria* (Codd et al. 2005, Falconer 2005). However, not all cyanobacteria are capable of toxin production. Different strains may produce different types of toxins in different quantities and at different rates along their growth cycle, while others are non-toxigenic (Sivonen & Jones 1999, Codd et al. 2005). These characteristics explain why the occurrence of toxic blooms in natural environments is, apparently, unpredictable.

Current awareness of hazards from toxic cyanobacteria in different countries has made considerable progress due to cooperative programs between countries and regions that are aimed towards implementing existing regulations (Chorus 2005). The prevention of harmful cyanobacterial blooms has been based on the assessment and minimization of eutrophication conditions, thus falling within the scope of the EU Water Framework Directive, which calls for a 'good ecological status' of public water resources by 2015.

A general picture of the extent of cyanobacterial and cyanotoxin occurrence is emerging from scientific research, preliminary surveys and on-going monitoring in many countries (Chorus 2005). In Portugal, awareness of potential health risks has led to several research projects on freshwater and estuarine microbial ecology in the southern regions of Alentejo and Algarve. Furthermore, a working group, coordinated by the Portuguese General Directorate for Health, was formed in 1995–1996 to present a proposal for a national monitoring program of cyanobacteria and cyanotoxins in freshwater reservoirs. Since then, various laboratories have been carrying out regular screening of cyanobacteria in many freshwater reservoirs, alerting environmental and water management agencies to the more serious toxic occurrences and assessing the efficiency of water treatment processes in removing cyanobacteria and associated toxins.

The southwest region of the Iberian Peninsula is a semi-arid region with a Mediterranean climate where the dry season usually extends from May to September and annual precipitation averages ca. 500 mm. The concentration of rainy days defines a torrential regime, where years of extended droughts alternate with years of intense rain and floods. Water reservoir management is strongly limited by water availability, leading to several situations of high hydraulic residence time while new dam construction further restricts river flow. All these conditions influence the development of cyanobacterial blooms and aggravate their environmental and economic impact.

This paper presents 3 different case studies from Alentejo and Algarve. The studies were presented at a workshop on harmful algal blooms, focusing primarily on cyanobacteria, organized within the SAME 10 held in September 2007 in Faro, Portugal. The first study focuses on the Guadiana River estuary, which runs through both regions. The second study compares cyanobacterial dynamics in 4 freshwater reservoirs of the Algarve region (Bravura, Funcho, Odeleite and Beliche), and the third relates microcystin production with the community composition of cyanobacteria in 5 reservoirs from the Alentejo region (Alqueva, Alvito, Enxoé, Odivelas and Roxo). Although describing different situations in terms of cyanobacterial occurrence

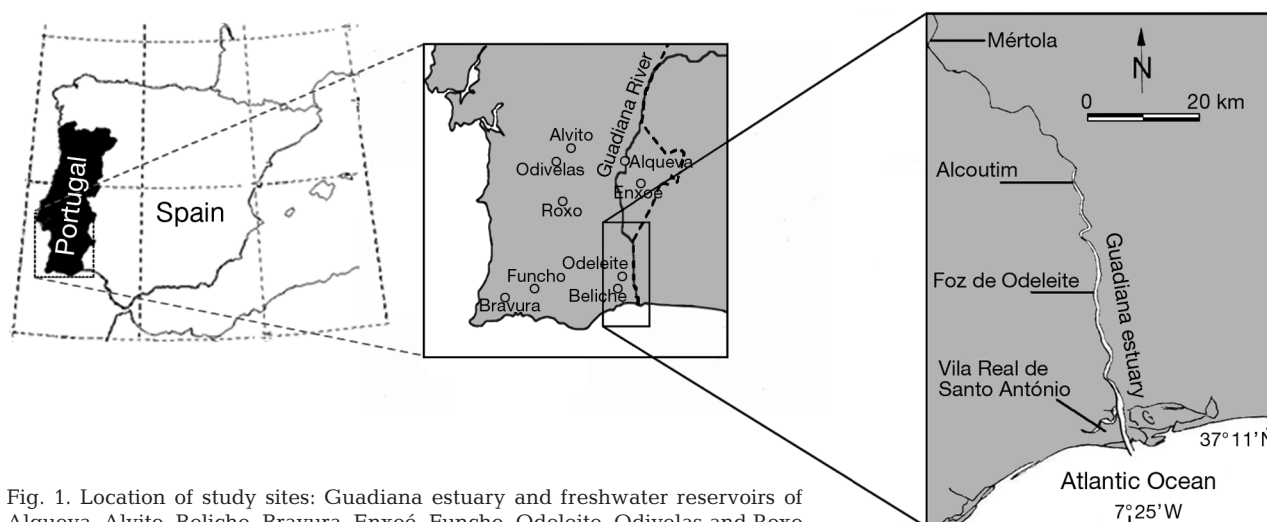


Fig. 1. Location of study sites: Guadiana estuary and freshwater reservoirs of Alqueva, Alvito, Beliche, Bravura, Enxoe, Funcho, Odeleite, Odivelas and Roxo

and toxicity, these 3 case studies represent complementary approaches to understanding the complexity of cyanobacterial blooms in Portuguese environments.

IMPACT OF DAM CONSTRUCTION ON THE GUADIANA ESTUARY

The Guadiana River, running along the southern border between Portugal and Spain (Fig. 1), has the fourth largest drainage basin of all Iberian rivers (67 840 km²), but a series of dams have severely restricted its freshwater flow by ca. 75%, and the construction of the large Alqueva dam increased flow regulation to up to 81% of the total catchment area (55 000 km²) starting in 2003.

Cyanobacterial blooms have been reported in the Guadiana River in association with seasons and/or years of low freshwater flow (Cabeçadas & Brogueira 1981, Oliveira 1985, 1991). Microbial ecology studies carried out from 1996 to 1998 showed a well-defined chlorophyll maximum in the upper estuary (Fig. 2), ca. 30 km upstream from the mouth of the river (Rocha et al. 2002, Domingues et al. 2005). This observation has been made in many temperate estuaries where estuarine circulation leads to the development of a maximum turbidity zone at the toe of the salt-wedge, upstream of which phytoplankton find optimal conditions for growth (Cloern 1987, 1996, 1999). Moreover, Rocha et al. (2002) indicated that high winter loads of nitrogen and phosphorus in the upper estuary were followed by silicate depletion during early spring caused by spring diatom blooms. This nutrient regime determined the phytoplankton succession from diatoms in early spring to chlorophytes, followed by cyanobacteria during summer and fall (Rocha et al. 2002, Domingues et al. 2005).

Changes in freshwater flow lead to alterations in water quality and hydrography, thus affecting phytoplankton composition and succession. River flow after completion of the Alqueva dam was severely restricted even during winter months with high rainfall (Fig. 3). During the period of dam construction (1999 to 2000), the sediment load transported downstream increased dramatically, causing severe photolimitation for the phytoplankton and resulting in the absence of blooms and very low chlorophyll values throughout spring and summer. One exception occurred in April 2001, when chlorophyll reached a record high of 216 µg l⁻¹ due to an unknown chain-forming centric diatom coinciding with a sharp decrease in seston when dam construction was terminated (Fig. 4). Afterwards, during the filling period of the reservoir (2002 to 2003), nutrient concentrations as well as cyanobacterial abundance increased, while diatom abundance remained low (Domingues et al. 2007). After this period, total phytoplankton abundance and succes-

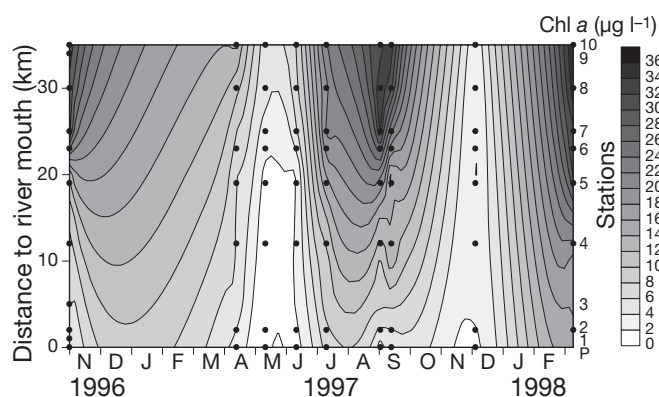


Fig. 2. Interpolation plots showing the longitudinal and seasonal distribution of surface chl a concentrations at selected stations in the Guadiana estuary (P: river mouth, 1: Vila Real St. Antonio, 6: Odeleite Stream, 10: Alcoutim)

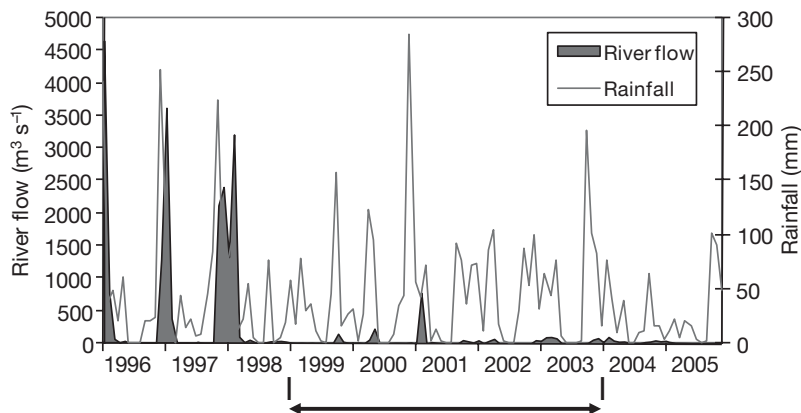


Fig. 3. Monthly river flow volume (Pulo do Lobo upstream from Mértola, mean values) and total monthly rainfall (Alcoutim) from 1996 to 2005 in the upper Guadiana estuary (data source: www.inag.pt/SNIRH/). The arrow marks the dam construction and filling period

sion followed the typical trend observed before dam filling. However, specific diversity and chlorophyll concentrations decreased from 2002 to 2005 (Domingues & Galvão 2007).

Microcosm studies undertaken in 2005 to explain the phytoplankton community structure based on nutrient regime indicated that nitrogen limitation could have influenced phytoplankton succession (R. B. Domingues unpubl. data). However, this type of experiment was not designed to simulate an important factor, namely estuarine circulation. In fact, the salinity regime in the estuary was affected during dam filling and construction due to the salt-wedge intrusion reaching further upstream, causing saltwater to be detected in the normally freshwater upper estuary (ca. 70 km upstream from the river mouth). The phytoplankton community found in the upper estuary, including cyanobacteria, was composed strictly of freshwater species with limited halotolerance. Therefore, an increase in salinity, particularly during late spring and early summer when maximum growth should occur, could have had a deleterious impact resulting in lower chlorophyll values and species numbers after construction of the Alqueva dam.

Microcystins were detected in 1999, 2000, 2002 (Caetano et al. 2001, Sobrino et al. 2004), 2004 and 2005 (H. M. Galvão unpubl. data) in concentrations well below the World Health Organization (WHO) guideline for drinking water of $1 \mu\text{g l}^{-1}$ (WHO 1998). Microcystin levels remained low, even during cyanobacterial blooms with abundances of potentially toxic species, such as *Microcystis aeruginosa*, reaching 10^6 to 10^8 cells ml^{-1}

(Caetano et al. 2001, Rocha et al. 2002). Microcystin concentrations higher than the WHO guideline were detected only once, on 25 July 2003, when values of $1.2 \mu\text{g l}^{-1}$ were reached in Mértola (70 km upstream from the river mouth) and $6.8 \mu\text{g l}^{-1}$ upstream from Mértola (C. Sobrino unpubl. data). Interestingly, these higher toxin levels were detected while total cyanobacteria counts, determined using epifluorescence microscopy (Haas 1982), were relatively low (maximum of 6×10^5 cells ml^{-1}) and potentially toxic genera such as *M. aeruginosa* registered low abundance. However, these low total counts could have reflected a shift in predominance from chroococcoid species towards filamentous forms such as *Oscillatoria* and *Anabaena* spp. with higher

specific biomass and potential toxin production. Furthermore, other samples taken on 30 July 2003 (M. R. Reis unpubl. data) contained high numbers of *M. aeruginosa* ($>6 \times 10^5$ cell ml^{-1}) and potentially toxic filamentous forms ($>1 \times 10^5$ cells ml^{-1}). Bathing and fishing in this area were forbidden by health authorities during July and August 2003 because high toxin concentrations persisted for several weeks.

Cyanobacterial blooms in freshwater reservoirs in the Algarve region

In the Algarve, strong demographic changes induced by tourism have led to the building of several freshwater reservoirs in order to ensure the water quantities required for activities such as agriculture, golf, landscaping and human consumption. Thus, from 2000 onwards, there was a shift from groundwater to surface water as a source of drinking water.

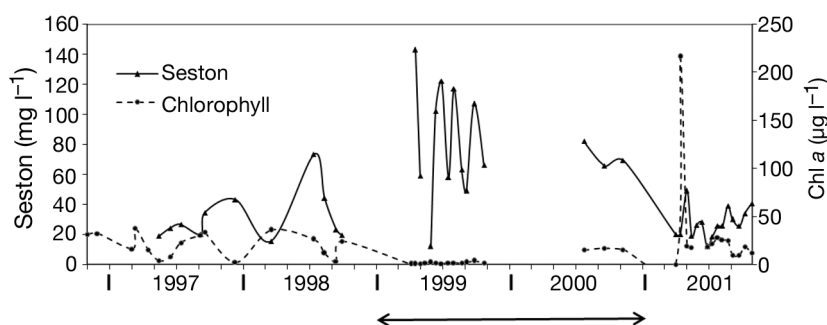


Fig. 4. Seston and chl a concentrations in surface water samples from 1997 to 2001 at the Alcoutim station in the upper Guadiana estuary. The arrow marks the dam construction period. Gaps in the data reflect periods without regular sampling

In western Algarve, the main reservoir used for drinking water production is Funcho (Fig. 1), while water removal from Bravura is seasonal, usually occurring from May to October. The eastern Algarve is supplied by water from Beliche, which is connected to the Odeleite reservoir by underwater sluices that regulate the flow from Odeleite to Beliche and which are usually opened in spring and closed in autumn. Basic information about location and dimension of the 4 main reservoirs in the Algarve is compiled in Table 1.

These reservoirs have been subject to standard physical, chemical and microbiological water quality monitoring since 1996 and, since 2001, to simultaneous monitoring of phytoplankton and cyanobacteria toxins. We organized all collected data into 2 types of database: one containing a long-term data series of physical and chemical variables and one with a shorter data series, but including biological data and diversity indices like Shannon diversity, evenness, Berger-Parker dominance (Berger & Parker 1970) and species richness (Magurran 1988). For the present study, multivariate analysis of these data sets was carried out in order to assess the main factors triggering cyanobacteria blooms.

Relative abundances of cyanobacteria and microcystin concentrations obtained over a 2.5 yr period in the western and eastern Algarve are illustrated in Figs. 5 & 6, respectively. In the western reservoirs, 40 to 50 % of the water samples contained cyanobacterial concentrations higher than the WHO alert level 1 for raw waters (≥ 2000 cells ml^{-1}). However, more than 80 % of the samples in Odeleite and Beliche exceeded this alert level. Furthermore, 13 to 17 % of all analyzed samples presented concentrations of $>20\,000$ cells ml^{-1} , but WHO alert level 2 ($\geq 100\,000$ cells ml^{-1}) was only occasionally exceeded in Funcho in 2001, 2002 and 2005, when *Microcystis* spp. reached $>10^6$ cells ml^{-1} . Microcystin concentrations were always negligible with respect to human health risk, only exceeding WHO guideline value of $1\,\mu\text{g l}^{-1}$ in scums of a winter bloom detected in Funcho in 2002, and in bottom samples of a spring bloom detected in Beliche in 2004.

Cyanobacterial blooms occurred during the dry as well as during the rainy season, and no evident relationship between weather conditions and bloom occurrence was found. Data analyses of monthly water column profiles for temperature and oxygen concentrations revealed strong differences in the stratification of the 4 reservoirs (M. R. Reis unpubl. data), probably related to the different water management strategies. While Funcho reservoir behaved as a warm monomictic lake with winter destratification occurring only from December to February, the Bravura water column tended to destratify 2 mo earlier and stratify 2 to 3 mo

Table 1. Location and dimensions of Algarve and Alentejo freshwater reservoirs

Reservoir	Stream	Watershed	Catchment area (km^2)	Latitude ($^{\circ}\text{N}$)	Longitude ($^{\circ}\text{W}$)	Year of closure	Max. water column height (m)	Total volume ($\times 10^6\,\text{m}^3$)	Flooded surface (ha)	Mean annual precipitation (mm)
Algarve										
Bravura	Ribeira de Odiáxere	Ribeiras do Algarve	76.58	$37^{\circ}12'6''$	$8^{\circ}41'52''$	1952	23	35	285	821
Funcho	Arade	Arade	212.59	$37^{\circ}15'46''$	$8^{\circ}22'57''$	1993	31	48	360	744
Odeleite	Ribeira de Odeleite	Guadiana	347.51	$37^{\circ}19'52''$	$7^{\circ}29'11''$	1996	30	130	720	722
Beliche	Ribeira de Beliche	Guadiana	98.47	$37^{\circ}16'35''$	$7^{\circ}30'33''$	1986	30	48	292	644
Alentejo										
Alqueva	Guadiana	Guadiana	55 289.00	$38^{\circ}11'50''$	$7^{\circ}29'43''$	2002	22	4150	25 000	593
Alvito	Ribeira do Alvito	Sado	210.77	$38^{\circ}16'44''$	$7^{\circ}54'37''$	1968	25	132	1480	640
Odivelas	Ribeira de Odivelas	Sado	431.88	$38^{\circ}11'05''$	$8^{\circ}06'55''$	1972	12	96	973	622
Enxoé	Ribeira de Enxoé	Guadiana	61.15	$37^{\circ}59'38''$	$7^{\circ}27'55''$	1998	10	10	–	555
Roxo	Ribeira do Roxo	Sado	351.1	$37^{\circ}55'47''$	$8^{\circ}04'55''$	1967	14	96	1378	559

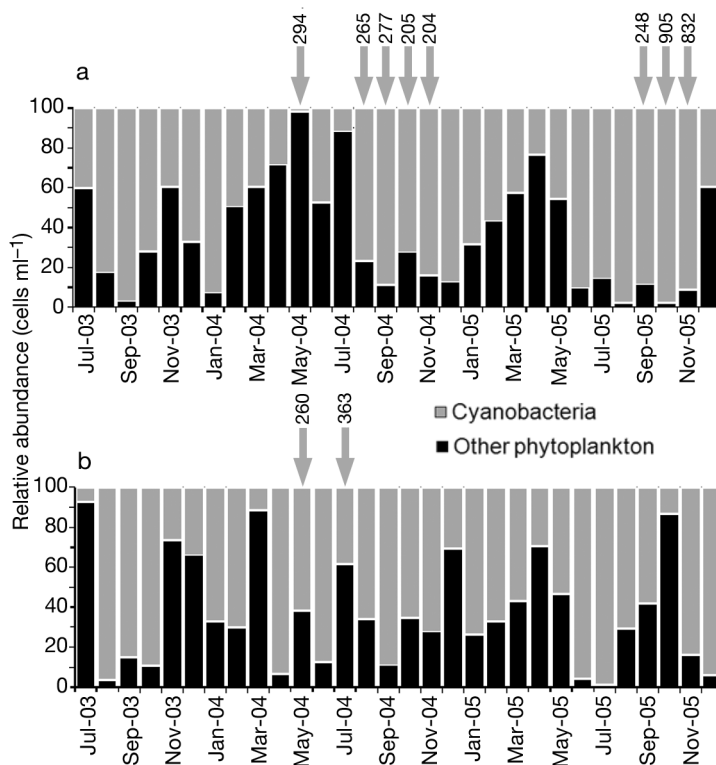
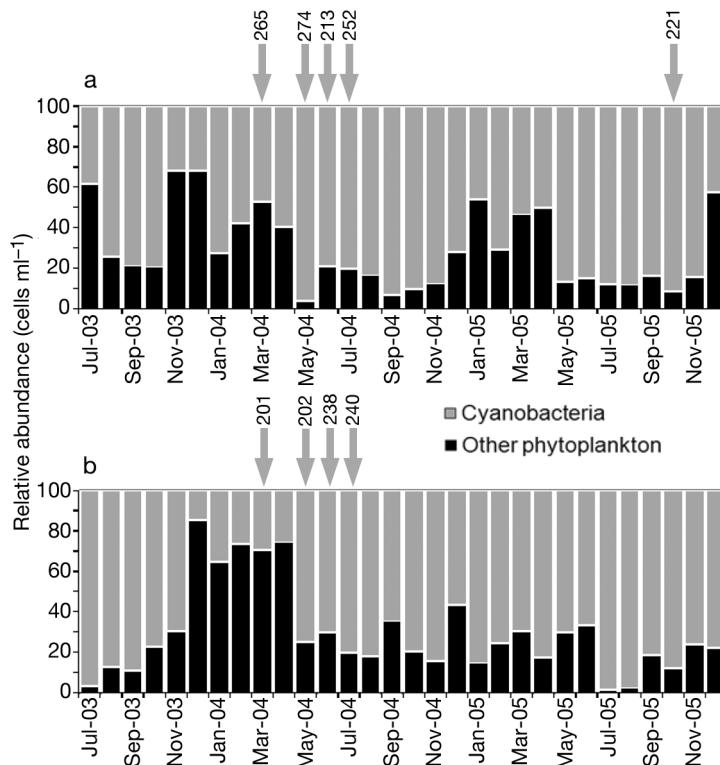


Fig. 5. Relative abundances of cyanobacteria in the Western Algarve reservoirs, (a) Bravura and (b) Funcho. Arrows show samples positive for microcystins; values beside arrows represent microcystin concentrations (ng l^{-1})



later. This difference could only be explained by the artificial reinforcement of the Funcho thermocline, caused by year-round cold water removal from the hypolimnion for drinking water production.

Concerning the 2 eastern reservoirs, management of the underground channel between Odeleite and Beliche artificially induced a polymictic behavior that can also be associated with conditions favouring blooms of filamentous cyanobacteria through the resuspension of bottom sediment and nutrients. This hydraulic regulation may also have caused differences in the observed phytoplankton succession. The eastern reservoirs (Odeleite and Beliche) systematically developed mixed cyanobacterial blooms of *Nostocales* and *Oscillatoriales* in late spring, which were then gradually replaced by *Chroococcales*. However, in 2005, the channel between Odeleite and Beliche was opened earlier because of a severe drought. Nutrients and akinetes were resuspended in cold waters unfavourable for *Nostocales* blooms. During this drought, *Chroococcales* dominated spring blooms even in the eastern reservoirs.

Based on the dominance of cyanobacteria, all 4 reservoirs could be considered eutrophic, but based on soluble reactive phosphorus (SRP) and dissolved inorganic nitrogen (DIN) concentrations, the western reservoirs (Bravura and Funcho) would be classified as slightly mesotrophic and the eastern (Odeleite and Beliche) as oligotrophic. Frequent cyanobacterial blooms occurring in oligotrophic reservoirs are in contradiction with most published reports, which correlate bloom formation with eutrophication or low N:P ratios (Chorus 2001).

Fig. 7 illustrates how blooms and associated DIN:SRP ratios in Funcho were related to the Redfield ratio of 16N:1P. In fact, DIN never exceeded the desirable limit for a lake in an arid region (US Environmental Protection Agency standard, www.epa.gov/waterscience/criteria/nutrient) in the eastern reservoirs. In the western reservoirs, this value was only exceeded occasionally in winter. In view of potential SRP-limitation, cyanobacterial blooms during summer were generally not linked to SRP levels, except for the 2002 winter bloom.

Fig. 6. Relative abundances of cyanobacteria in the Eastern Algarve reservoirs (a) Odeleite and (b) Beliche. Arrows show samples positive for microcystins; values beside arrows represent microcystin concentrations (ng l^{-1})

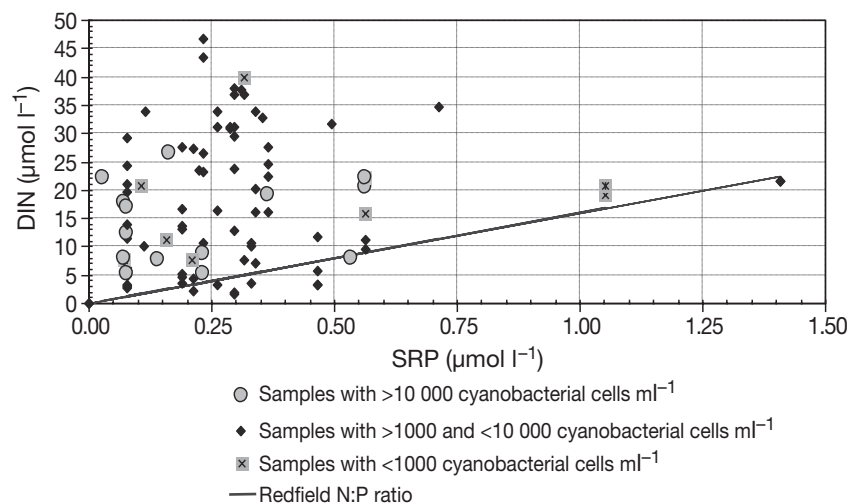


Fig. 7. Relation between cyanobacterial cell concentration and molar proportions of dissolved inorganic nitrogen (DIN) to soluble reactive phosphorus (SRP) in the Funcho reservoir in western Algarve (July 2001 to June 2004). The line indicates the 16N:1P Redfield ratio

Exclusive use of physical and chemical data in multivariate analysis of bloom dynamics was not sufficient to discriminate bloom conditions (Reis 2005). This led to the inclusion of biological variables in multivariate analysis of bloom dynamics. Using methods such as distance based redundancy analysis (db-RDA), summer *Chroococcales* blooms could be associated with increased temperature while the winter *Aphanizomenon flos-aquae* bloom was associated with increased SRP. Trend analysis through dynamic factor analysis (Zuur et al. 2003) applied to biological variables revealed no seasonal pattern and a tendency towards decreasing frequency and density of cyanobacterial blooms. This could be related to the fact that these reservoirs are relatively young water bodies that have not yet developed a stable food web with effective top-down control (Reis 2005).

Cyanobacterial blooms and associated microcystins in freshwater reservoirs in the Alentejo region

The occurrence of microcystins in Portuguese freshwater resources has been mostly associated with blooms of *Microcystis aeruginosa*, although other potentially toxic species also occur (Vasconcelos 1993, 1994, 2001, Vasconcelos et al. 1996, Pereira et al. 2001, Vasconcelos & Pereira 2001, Moreno et al. 2003, Saker et al. 2005). A wide range of microcystin concentrations have been encountered in many Portuguese rivers and reservoirs, though most reports are based on the sporadic analysis of environmental samples taken at the time of a bloom event, rather than on a systematic survey of aquatic ecosystems.

In this study, cyanobacteria and microcystins were monitored in 5 Portuguese freshwater reservoirs over 2 consecutive time periods, from May to December of 2005 and from April to July of 2006. This was intended to evaluate changes in microcystin concentrations and persistence with respect to variations in cyanobacterial assemblages and biomass throughout the sampling period. Table 1 lists some basic information about each of the 5 reservoirs included in the study (Alqueva, Alvito, Enxoé, Odivelas and Roxo), while Fig. 1 shows their location.

Results concerning phytoplankton abundance and microcystin concentrations found in each reservoir during the monitoring period are shown in Fig. 8. The occurrence of cyanobacteria seemed to follow a seasonal pattern in most reservoirs, with major peaks

being detected during summer periods and lower cell densities during cold periods. Although this appeared to be a general trend, the different reservoirs also showed varying patterns in terms of bloom-forming species abundance, biomass and toxicity. In Alqueva, a noticeable shift in phytoplankton composition towards cyanobacterial dominance was observed from July to October 2005. After being replaced by chlorophytes and cryptophytes during spring 2006, cyanobacteria dominated the phytoplankton again in July 2006. *Microcystis aeruginosa* and *Anabaena circinalis* were the predominant bloom-forming species, accounting for 11 to 100% and 3 to 58% of total cyanobacterial biomass, respectively. However, although both potentially toxic species reached high cell densities, microcystins were not detected throughout the sampling period.

A different pattern was observed in the Alvito reservoir, where microcystins were detected in several samples containing high counts of cyanobacteria. Although different bloom-forming species were found, toxin concentrations were better correlated with *Microcystis aeruginosa* biomass than with total cyanobacterial biomass. In fact, *Anabaena circinalis* dominated the cyanobacterial assemblage from July to August 2005, subsequently being replaced as a dominant species by *Aphanizomenon flos-aquae*, which formed an extremely dense bloom in September. During this period, the relative contribution of *M. aeruginosa* to the overall cyanobacterial community never exceeded 4% of total cell counts, and microcystins remained below $1.10 \mu\text{g l}^{-1}$. However, an increase in toxin concentration in October was associated with a change in bloom

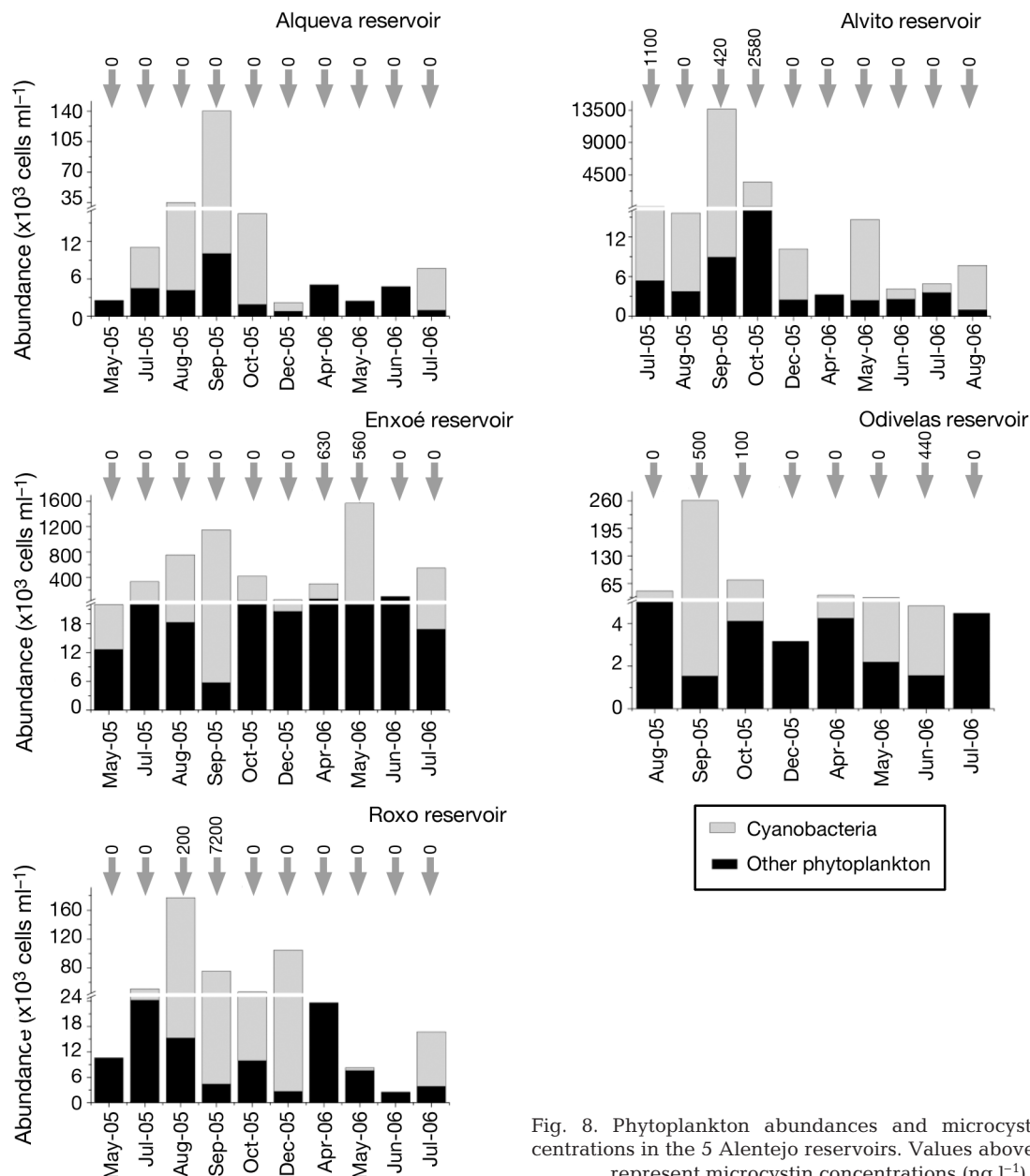


Fig. 8. Phytoplankton abundances and microcystin concentrations in the 5 Alentejo reservoirs. Values above arrows represent microcystin concentrations (ng l⁻¹)

composition, with abundances shifting towards *M. aeruginosa* dominance. Although cyanobacteria were present in much lower total cell counts, microcystins reached a maximum of 2.58 µg l⁻¹ during this period.

In Enxoé, 2 phytoplankton blooms were observed throughout the sampling period. The first one occurred in September 2005, following a steady increase in cyanobacterial biomass throughout the summer period. The second bloom occurred in May 2006, when an extremely high abundance of cyanobacteria occurred after a period of low cell counts during winter. Both blooms were dominated by *Planktothrix* spp. However, microcystins were only detected during the second bloom. Interestingly, no cyanobacteria were

found in June 2006, just one month after the major peak in toxic cyanobacteria. Subsequently, *Planktothrix* spp. dominated again in July and microcystins were below detection limits, similar to the pattern observed in the previous year. In fact, microcystin concentrations did not even co-vary with *Planktothrix* spp. cell counts during the 2006 toxic bloom, when no other potentially toxic species were found.

In Odivelas, 2 distinct cyanobacterial blooms also occurred, but with a different species composition. In August 2005, 3 different species of *Aphanizomenon* spp. predominated, contributing ca. 40% of the total cyanobacterial counts. Other species, such as *Anabaena* spp. and *Microcystis aeruginosa*, were also

detected, but with much lower relative abundances (12 and 14 % of total cyanobacterial counts, respectively). One month later, predominance shifted sharply towards *M. aeruginosa*, with abundances increasing from 6500 cells ml⁻¹ in August to 131 000 cells ml⁻¹ in September, representing ca. 50 % of total cyanobacterial cell counts. Microcystin concentrations detected in the Odivelas reservoir were associated with high *M. aeruginosa* cell counts and reached a maximum of 0.5 µg l⁻¹ in September 2005. This *M. aeruginosa* bloom was followed in October by an *Aphanizomenon* spp. bloom, during which *M. aeruginosa* abundance decreased to <1300 cells ml⁻¹. Cyanobacteria were not observed during winter, but dominated the phytoplankton in the following spring with a different species composition. In fact, cyanobacterial blooms were mainly composed of *Microcystis wesenbergii* from April to May 2006, while microcystins were not detected.

In the Roxo reservoir, high cyanobacterial counts occurred from July to December 2005. Two cyanobacteria species dominated successively: *Microcystis incerta* in July and August (ca. 60 % of the total cyanobacterial cell counts), followed by *Planktothrix* sp. from September until December. High levels of microcystins were only detected at the onset of the *Planktothrix* sp. bloom in September 2005. In 2006, cyanobacterial blooms did not occur until July, when *Planktothrix* sp. reached abundances of 15 500 cells ml⁻¹.

DISCUSSION

The 3 studies presented here used different yet complementary approaches towards understanding the complexity of cyanobacterial bloom dynamics in different ecosystems subject to different sets of varying environmental constraints, which represent a major challenge for implementing adequate water management strategies.

The first study addressed the impact of a recently constructed dam on phytoplankton succession and cyanobacterial blooms in the Guadiana upper estuary and focussed on changes in hydrography and sediment load as well as light and nutrient regime. The second study examined cyanobacterial bloom dynamics in different freshwater reservoirs in the Algarve region in an attempt to discern general patterns in bloom occurrence as well as triggering factors. The third study evaluated seasonal changes in microcystin concentrations in relation to shifts in cyanobacteria species dominance, in terms of abundance and biomasses, in 5 freshwater reservoirs in the Alentejo region. Taken as a whole, these studies emphasize the need for a comprehensive approach towards understanding cyano-

bacterial bloom dynamics and toxin production in natural environments in order to correctly assess potential risk and outline adequate monitoring programs.

Initially, the impact of the Alqueva dam on phytoplankton dynamics was predicted to increase the predominance of cyanobacteria with a higher potential toxicity (Rocha et al. 2002). Instead, both cyanobacterial abundance and numbers of toxic species tended to decrease in 2003, after the period of dam construction and filling. Cyanobacterial peak abundances during summer and fall remained well below the maximum cell counts observed prior to dam construction in 1996 and 1997 (10⁷ to 10⁸ cells ml⁻¹), probably due to the interaction of several environmental factors such as photolimitation resulting from increased sediment load and haloinhibition due to deeper intrusion of saltwater in the upper estuary.

Data from the Guadiana estuary collected since 1996 revealed that, in the upper estuary, the seasonal succession of phytoplankton populations generally led to a predominance of cyanobacteria in the summer and fall, which could be explained by the nutrient regime (Rocha et al. 2002). Thus, silica depletion caused the demise of diatom blooms followed by increased abundance of chlorophytes in mid-spring. By the end of spring, nitrogen limitation favoured the predominance of small unicellular cyanobacteria from early summer to mid-fall. However, the construction of the Alqueva dam, which further restricted freshwater flow, caused changes in the nutrient regime with increased silica concentrations (Domingues et al. 2007), sediment load and saltwater intrusion in the upper estuary. In fact, estuarine circulation has probably been affected to such a degree that both the maximum turbidity zone and associated chlorophyll maximum no longer constitute well-defined features in the estuary. Unfortunately, slackwater runs (sampling following tidal excursion upstream during the slackwater phase), which enable proper delineation of longitudinal and vertical profiles, have not been regularly performed since 1998 due to lack of funding. However, a more recent project, financed by the Portuguese National Science Foundation (FCT), resumed this sampling program in 2008.

The freshwater reservoir studies demonstrated that different types of cyanobacterial blooms had diverse origins, while no clear seasonal pattern for cyanobacterial abundances was found. In the Algarve reservoirs, multivariate analysis was a useful tool to interpret reservoir dynamics with respect to water quality (Reis 2005). Cyanobacterial dominance in those reservoirs was not always associated with the nutrient regime, but summer blooms were related to high water temperatures. Water management strategies and water removal level affected water column stratifica-

tion, thus affecting phytoplankton dynamics and cyanobacterial blooms. In the western reservoirs, the enhancement of water column stratification, caused by the removal of cold water from the hypolimnion, led to the export of nutrients and produced a chemostat-like system for the growth of *Chroococcales*. In fact, the maintenance of a warm water surface layer positively selected for cyanobacteria and produced prolonged summer blooms. In the eastern reservoirs, mixing of artificial spring water caused by water level management strategies was associated with late spring blooms of *Nostocales* and *Oscillatoriales*, probably due to nutrients and akinete resuspension.

Meteorological conditions such as temperature and radiation could not be associated with total cyanobacterial abundances but only with particular bloom-forming species. Furthermore, trend analyses revealed that the abundance of bloom species decreased in both oligotrophic and mesotrophic reservoirs, perhaps due to increased top-down control. Finally, SRP limitation did not seem to affect summer blooms of cyanobacteria. Time series analyses (Legendre et al. 1985, Zuur et al. 2003) could be a useful tool for determining selective mechanisms which affect phytoplankton composition and dominance of cyanobacterial species. However, existing time series lack important ecological data such as competition, viral abundance, predation rates and selective predator abundance, which hinders the predictive accuracy of models for phytoplankton dynamics.

Independent of environmental constraints, the Alentejo reservoir study showed that microcystin concentrations in natural waters were not always related to cyanobacterial biomass or even to the abundance of potentially toxic species. Microcystin concentrations found in Alvito and Odivelas, for example, were related to *Microcystis aeruginosa* cell counts, suggesting that primarily this species was responsible for toxin production in these reservoirs. However, no microcystins were detected in Alqueva, despite the presence of *M. aeruginosa*. In Roxo, microcystins were only detected in high levels during the onset of the *Planktothrix* sp. bloom in September 2005. However, no microcystins were found in the following months, despite the prevalence of high abundances of *Planktothrix* sp. In Enxoe, microcystins were only detected in one *Planktothrix* sp. bloom, yet toxin concentrations did not follow *Planktothrix* sp. cell counts during the bloom.

Variations in toxin production between and within cyanobacterial species are well known from laboratory studies based on isolated strains (Rapala et al. 1997, Kameyama et al. 2004, Welker et al. 2004, Saker et al. 2005). Furthermore, blooms may be composed of cyanobacterial assemblages producing different toxins

at different rates, while the same bloom-forming species can occur as both toxic and non-toxic forms, which are impossible to differentiate based on morphological examination. In fact, all reservoirs included in this study showed very sharp changes in cyanobacterial abundance, whereas marked differences in toxicity were observed both between blooms and within the same bloom. These observations emphasize the irregular pattern of cyanobacterial blooms and cyanotoxin production. Consequently, there is a pressing need for the implementation of systematic monitoring programs of cyanobacteria and cyanotoxins in natural waters in order to minimize potential health risks to animal and human populations resulting from exposure through drinking water and recreational activities.

CONCLUSIONS

The need for better integration of ecological aspects in water quality assessment is generally accepted, but the European Union Directive (<http://ec.europa.eu/environment/water/water-framework/>) that encompasses measures for protection of the ecological quality of waters is far from being fully implemented. Systematic approaches to reservoir ecology should lead to the establishment of ecological water quality criteria, which in turn should help to outline adequate monitoring programs with specific time-scale adjustments, depending on the ecosystem. Ecological water quality criteria should be useful not only in promoting environmental conservation and rehabilitation but also as a tool for raw water management, allowing, for instance, drinking water treatment plants to more efficiently adjust their technologies for the removal of *Cyanobacteria* and cyanotoxins.

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