

Original Article

Long noncoding RNA LINC01296 induces non-small cell lung cancer growth and progression through sponging miR-5095

Xuefei Hu*, Liang Duan*, Hongcheng Liu, Lei Zhang

*Department of Thoracic, Shanghai Pulmonary Hospital, Tongji University, Shanghai 200433, China. *Co-first authors.*

Received November 13, 2018; Accepted December 21, 2018; Epub February 15, 2019; Published February 28, 2019

Abstract: Long noncoding RNAs (lncRNAs) played in authentic biological cell roles such as cell apoptosis, cycle, differentiation, development, migration and invasion. However, the expression pattern and function of a new lncRNA LINC01296 in non-small cell lung cancer (NSCLC) are unknown and need to be studied. In our study, we indicate that the expression of LINC01296 was overexpressed in NSCLC samples compared to adjacent non-tumor tissues. Ectopic expression of LINC01296 promoted NSCLC cell proliferation and migration. Moreover, we demonstrated that LINC01296 has a potential binding site for miR-5095 by using online program tool StarBase. Overexpression of LINC01296 inhibited the expression of miR-5095 in the A549 cell. Furthermore, the miR-5095 expression was downregulated in the NSCLC tissues than in the adjacent non-tumor tissues. In addition, we found that there is a negative correlation between miR-5095 expression and LINC01296 level in the NSCLC tissues. Overexpression of miR-5095 suppressed NSCLC cell proliferation and migration. Finally, we demonstrate that ectopic expression of LINC01296 promoted cell proliferation and migration via inhibiting miR-5095 expression. These results suggested that LINC01296 might act a role as an oncogene in the tumorigenesis and development of NSCLC.

Keywords: Non-small cell lung cancer, lncRNA, LINC01296, miR-5095

Introduction

Lung cancer is primary cause of the mortality in both women and men and accounted for majority of tumor deaths worldwide [1-5]. Non-small cell lung cancer (NSCLC) is the most frequently type of lung tumor and accounts for about 85% of primary lung cancer [6-9]. Characteristically, cells of NSCLC could migrate to local samples and spread to distant tissues in the later stage [10-12]. Despite progression in chemotherapeutic and surgical interventions, the five-year survival of NSCLC cases remains discontent and recurrence rate of NSCLC patient is high, due to tumor metastasis [6, 13-16]. Thus, it is urgent to study the molecular mechanisms underlying initiation and development of NSCLC and find new therapeutic strategies for treatment of NSCLC.

Long noncoding RNAs (lncRNAs) are arbitrarily regarded as longer than about 200 nucleotides

(nts) with no or limited protein coding capacity [17-20]. Recently, some lncRNAs were revealed to be played in authentic biological cell roles such as cell apoptosis, cycle, differentiation, development, migration and invasion [21-24]. A number of lncRNAs were deregulated in diverse tumors including colorectal cancer, hepatocellular carcinoma, osteosarcoma, gastric cancer, ovarian carcinoma and also NSCLC [25-30]. More recently, a new LINC01296 was shown to be involved in several tumors development such as prostate cancer, colorectal cancer, bladder cancer, gastric cancer, esophageal squamous cell carcinoma (ESCC), cholangiocarcinoma and osteosarcoma [31-37]. For instance, Qiu et al. [33]. Found that LINC01296 was upregulated and the higher expression of LINC01296 was an independent predictor for colorectal cancer patients' prognosis. Seitz and colleagues used the lncRNA profiling to found that LINC01296 was a candidate oncogene in the bladder cancer [34]. Qin et al. [37] indicat-

LINC01296 induces non-small cell lung cancer growth and progression

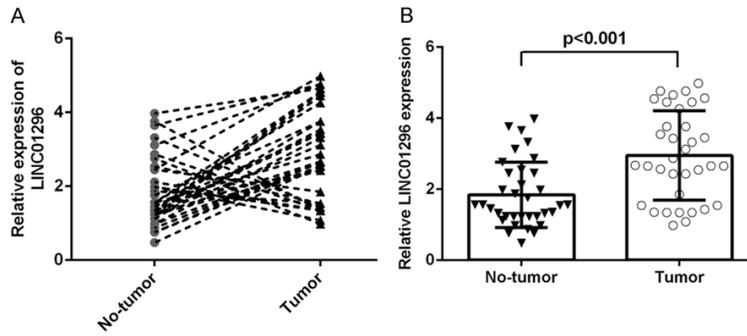


Figure 1. LINC01296 was overexpressed in NSCLC samples. A. The expression of LINC01296 was upregulated in 25 cases (25/35; 71.4%) compared to adjacent non-tumor tissues. The LINC01296 expression was determined by qRT-PCR. B. The LINC01296 expression was higher in the NSCLC tissues than in the adjacent non-tumor tissues.

ed that the LINC01296 expression was overexpressed in gastric cancer tissues and knock-down expression of LINC01296 inhibited gastric cancer cell migration, invasion and proliferation and induced cell apoptosis via regulating miR-122 expression. Zhang and colleagues showed that LINC01296 expression was overexpressed in cholangiocarcinoma and the LINC01296 expression was positively correlated with clinical stage and tumor severity [32]. LINC01296 knockdown inhibited cell migration, invasion and viability. They also demonstrated that LINC01296 induced cholangiocarcinoma growth and progression through inhibiting miR-5095. However, its expression pattern and function in NSCLC are unknown and need to be studied.

In this research, we indicate that the expression of LINC01296 was overexpressed in NSCLC samples compared to adjacent non-tumor tissues. Ectopic expression of LINC01296 promoted NSCLC cell proliferation and migration.

Materials and methods

NSCLC tissues, cell lines cultured and plasmid DNA Transfection

Human NSCLC tissues samples and matched non-tumor samples were collected from Thoracic department of Shanghai Pulmonary Hospital, Tongji University (Shanghai, China). These samples were speedy frozen in the liquid nitrogen until use. Human NSCLC cell lines (H1299, SPC-A1, A549 and H23) and one normal bronchial epithelial cell line (16HBE) were purchased from the ATCC (American Type Culture Col-

lection, Manassas, USA). These cells were kept in RPMI-1640 complete medium. miR-5095 mimic and scramble, pcDNA-LINC01296 and control plasmid were collected from Invitrogen (Carlsbad, California). Lipofectamine 2000 transfection kit (Invitrogen, USA) was utilized for transfection.

RNA isolation and qRT-PCR

Total RNA from samples or tissues was extracted with TRIzol (Bio Basic, Inc., Toronto, Canada). Quantitative PCR (qRT-PCR) was conducted to measure the expression of lncRNA, miRNA and mRNA on the 7900HT Real-Time PCR System (Applied Biosystems) with SYBR Kit (Takara Bio Inc). GAPDH and U6 were taken as the internal control for mRNA or miRNA respectively. The $2^{-\Delta\Delta Ct}$ was done to detect RNAs. The primer sequences which were used in the study were shown as following: miR-5095, 5'-TACAGG CGTGAACCA-CC-3'; LINC01296 forward sequences, 5'-GAAGCAGTGGTGGGTTCC-3' and reverse sequences, 5'-GAGCAACACAGATGAACCGC-3'; GAPDH forward sequences, 5'-TCCTCTGACTTCAACAGCGACAC-3' and reverse sequences, 5'-CACCTGTGCTGTAGCCAAATTC-3'.

Dual luciferase reporter assay

A mixture containing pMIR-LINC01296 Mut or pMIR-LINC01296 WT plasmid, pRL-TK Renilla Plasmid, miR-5095 mimic or scramble were co-transfected into A540 cell by using Lipofectamine 2000 (Invitrogen). Renilla and Firefly luciferase activities were determined at 2 days post-transfection with Dual Luciferase Reporter (Promega). The Renilla luciferase activity was used as the control.

Cell growth assays and migration assay

For cell growth, cells were cultured in the 96-well plate. Cell Counting Kit-8 (CCK-8, Dojindo, Kumamoto, Japan) was done to measure the cell proliferation. Proliferation rate was determined at 0, 1, 2 and 3 day after treated. The absorbance at the 450 nm was determined by using Spectrophotometer (BioTek, Winooski, VT). For cell migration, wound healing assay

LINC01296 induces non-small cell lung cancer growth and progression

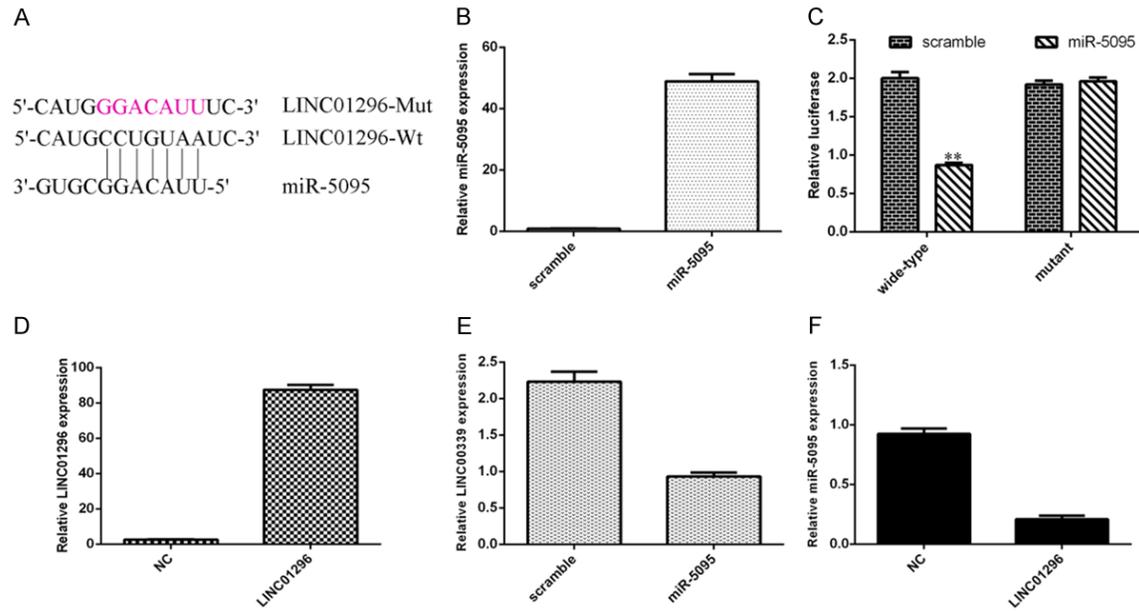


Figure 2. LINC01296 sponges miR-5095 in NSCLC cells. **A.** LINC01296 has a potential binding site for miR-5095 by using online program tool StarBase v3.0 (<http://starbase.sysu.edu.cn/>). **B.** The expression of miR-5095 was measured by qRT-PCR in the A549 cell. **C.** Ectopic expression of miR-5095 reduced the luciferase reporter activity carrying the 3'-UTR of LINC01296 but not the mutant of 3'-UTR of LINC01296. **D.** The expression of LINC01296 was upregulated in the A549 cell after treated with pcDNA-LINC01296 by qRT-PCR. **E.** Ectopic expression of miR-5095 suppressed the LINC01296 expression in the A549 cell. **F.** Overexpression of LINC01296 inhibited the expression of miR-5095 in the A549 cell. ** $P < 0.01$.

was performed. When the cell was reached confluence, the cell wound was created by pipette tip. Figures were taken subsequent to scratching and at 48 hours subsequent to scratching.

Statistical analyses

Numerical data were shown as mean \pm standard deviation (SD). The difference among the treatment groups was analyzed by Student's t test. P at < 0.05 was considered to be statistically significant. The SPSS 8.0 (SPSS, Inc.) was used in this study.

Results

LINC01296 was overexpressed in NSCLC samples

To study the role of lncRNA LINC01296 in NSCLC, we firstly measured LINC01296 expression in 35 NSCLC samples and adjacent non-tumor samples through performing qRT-PCR. As indicated in the **Figure 1A**, the expression of LINC01296 was upregulated in 25 cases (25/35; 71.4%) compared to adjacent non-tumor tissues. Furthermore, the LINC01296 ex-

pression was higher in the NSCLC tissues than in the adjacent non-tumor tissues (**Figure 1B**).

LINC01296 sponges miR-5095 in NSCLC cells

The online program tool StarBase v3.0 (<http://starbase.sysu.edu.cn/>) was performed to predict miRNA-lncRNA interactions; we demonstrated that LINC01296 has a potential binding site for miR-5095 (**Figure 2A**). We then showed that the expression of miR-5095 was upregulated in the A549 cell after treated with miR-5095 mimic by qRT-PCR (**Figure 2B**). Next, dual luciferase reporter was done to validate the binding between LINC01296 and miR-5095. We demonstrated that ectopic expression of miR-5095 reduced the luciferase reporter activity carrying the 3'-UTR of LINC01296 but not the mutant of 3'-UTR of LINC01296 (**Figure 2C**). In addition, we indicated that the expression of LINC01296 was upregulated in the A549 cell after treated with pcDNA-LINC01296 by qRT-PCR (**Figure 2D**). Ectopic expression of miR-5095 suppressed the LINC01296 expression in the A549 cell (**Figure 2E**). Overexpression of LINC01296 inhibited the expression of miR-5095 in the A549 cell (**Figure 2F**).

LINC01296 induces non-small cell lung cancer growth and progression

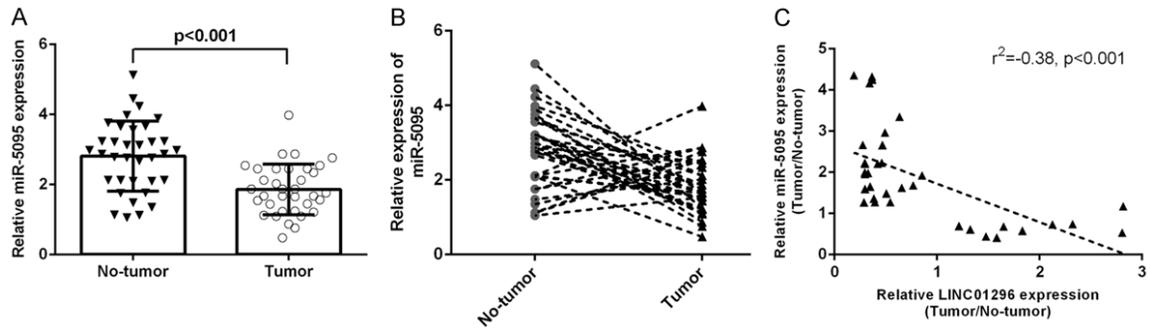


Figure 3. miR-5095 was downregulated in NSCLC samples. A. The expression of miR-5095 was downregulated in 26 cases (26/35; 74.3%) compared to adjacent non-tumor tissues. The miR-5095 expression was determined by qRT-PCR. B. The miR-5095 expression was lower in the NSCLC tissues than in the adjacent non-tumor tissues. C. There is a negative correlation between miR-5095 expression and LINC01296 level in the NSCLC tissues.

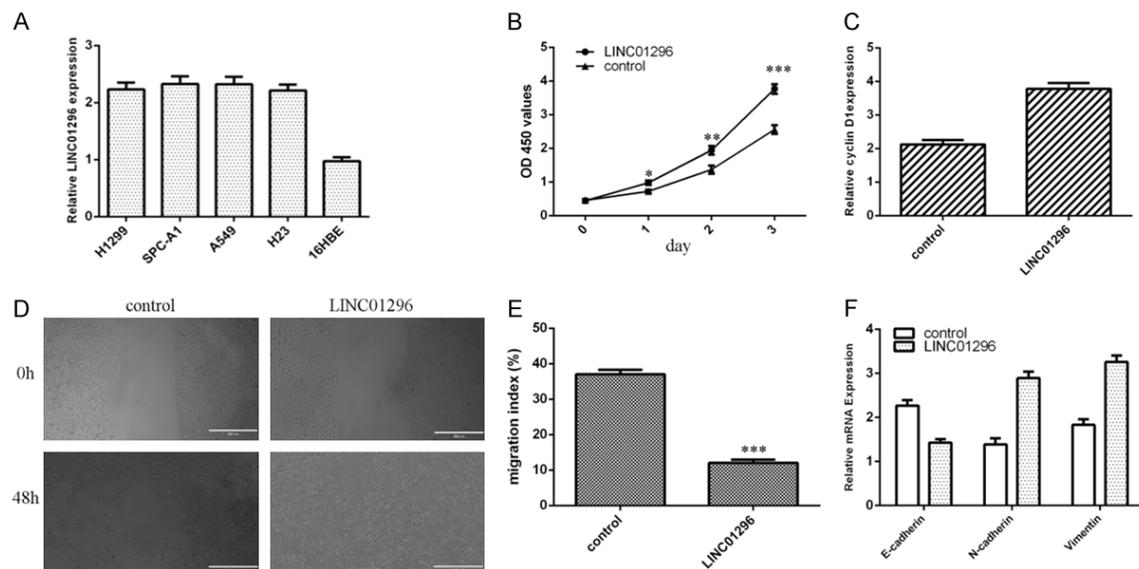


Figure 4. Ectopic expression of LINC01296 promoted cell proliferation and migration. (A) The expression of LINC01296 in the four NSCLC cell lines (H1299, SPC-A1, A549 and H23) and one normal bronchial epithelial cell line (16HBE) was detected by qRT-PCR. (B) Ectopic expression of LINC01296 induced the A549 cell proliferation by CCK-8 analysis. (C) Elevated expression of LINC01296 increased the cyclin D1 expression in the A549 cell. (D) Overexpression of LINC01296 promoted the A549 cell migration. (E) The relative migration index was shown (F) LINC01296 overexpression suppressed the expression of E-cadherin and induced the expression of N-cadherin and Vimentin in the A549 cell. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$.

miR-5095 was downregulated in NSCLC samples

To study the role of miR-5095 in NSCLC, we then detected miR-5095 expression in 35 NSCLC samples and adjacent non-tumor samples through performing qRT-PCR. As shown in the **Figure 3A**, the expression of miR-5095 was downregulated in 26 cases (26/35; 74.3%) compared to adjacent non-tumor tissues. Furthermore, the miR-5095 expression was lower in the NSCLC tissues than in the adjacent non-tumor tissues (**Figure 3B**). In addition, there is a

negative correlation between miR-5095 expression and LINC01296 level in the NSCLC tissues (**Figure 3C**).

Ectopic expression of LINC01296 promoted cell proliferation and migration

We then found that the expression of LINC01296 was upregulated in the four NSCLC cell lines (H1299, SPC-A1, A549 and H23) compared to one normal bronchial epithelial cell line (16HBE) (**Figure 4A**). Ectopic expression of LINC01296 induced the A549 cell proliferation

LINC01296 induces non-small cell lung cancer growth and progression

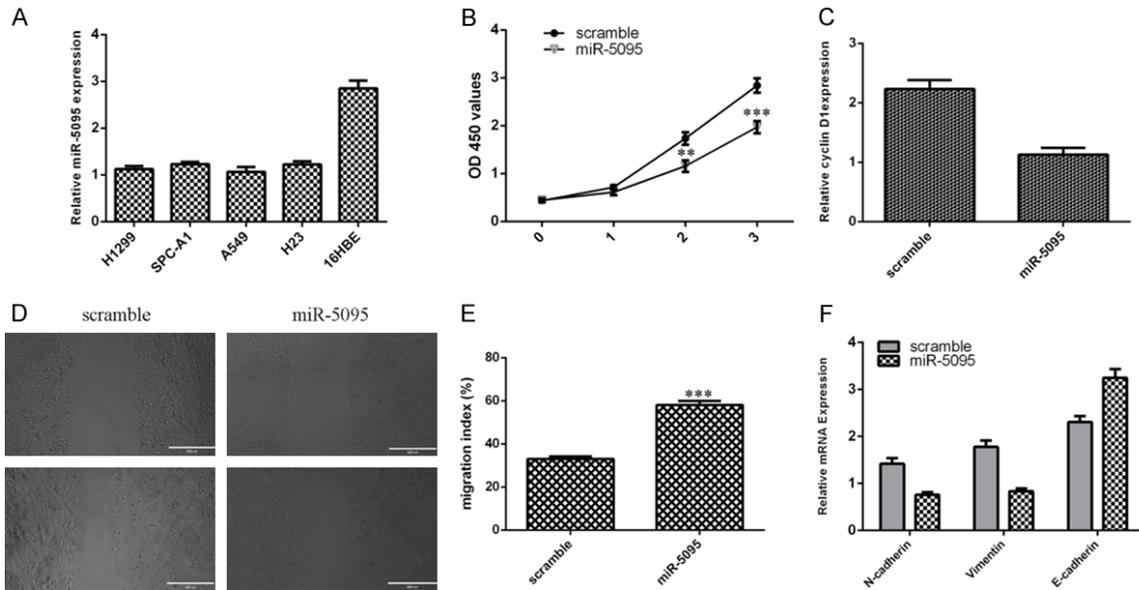


Figure 5. Overexpression of miR-5095 suppressed cell proliferation and migration. A. The expression of miR-5095 in the four NSCLC cell lines (H1299, SPC-A1, A549 and H23) and one normal bronchial epithelial cell line (16HBE) was detected by qRT-PCR. B. Ectopic expression of miR-5095 suppressed the A549 cell proliferation via CCK-8 analysis. C. Elevated expression of miR-5095 suppressed the cyclin D1 expression in the A549 cell. D. Overexpression of miR-5095 decreased the A549 cell migration. E. The relative migration index was shown. F. The mRNA expression of E-cadherin, N-cadherin and Vimentin in the A549 cell was detected by qRT-PCR. ** $P < 0.01$ and *** $P < 0.001$.

by CCK-8 analysis (Figure 4B). Moreover, elevated expression of LINC01296 increased the cyclin D1 expression in the A549 (Figure 4C). Furthermore, overexpression of LINC01296 promoted the A549 cell migration (Figure 4D and 4E). LINC01296 overexpression suppressed the expression of E-cadherin and induced the expression of N-cadherin and Vimentin in the A549 cell (Figure 4F).

Overexpression of miR-5095 suppressed cell proliferation and migration

Then, we demonstrated that the expression of miR-5095 was downregulated in the four NSCLC cell lines (H1299, SPC-A1, A549 and H23) compared to one normal bronchial epithelial cell line (16HBE) (Figure 5A). Ectopic expression of miR-5095 suppressed the A549 cell proliferation via CCK-8 analysis (Figure 5B). Moreover, elevated expression of miR-5095 suppressed the cyclin D1 expression in the A549 (Figure 5C). Furthermore, overexpression of miR-5095 decreased the A549 cell migration (Figure 5D and 5E). miR-5095 overexpression enhanced the expression of E-cadherin and inhibited the expression of N-cadherin and Vimentin in the A549 cell (Figure 5F).

miR-5095 reversed the promoting effects of LINC01296 on NSCLC cells

To investigate the important role of miR-5095 binding in LINC01296-inducing NSCLC progression, we transfected miR-5095 mimic in LINC01296 overexpressing A549 cell. We indicated that elevated expression of miR-5095 attenuated the inducing effect on cell proliferation of A549 cell via CCK-8 assay (Figure 6A). In addition, our data suggested that overexpression of miR-5095 suppressed the cyclin D1 expression in the LINC01296-overexpressing A549 cell (Figure 6B). Ectopic expression of miR-5095 enhanced the expression of E-cadherin and inhibited the expression of N-cadherin and Vimentin in the LINC01296-overexpressing A549 cell (Figure 6C). Elevated expression of miR-5095 attenuated the promoting effect on cell migration of A549 cell (Figure 6D and 6E).

Discussion

In our study, we indicate that the expression of LINC01296 was overexpressed in NSCLC samples compared to adjacent non-tumor tissues. Ectopic expression of LINC01296 promoted NSCLC cell proliferation and migration.

LINC01296 induces non-small cell lung cancer growth and progression

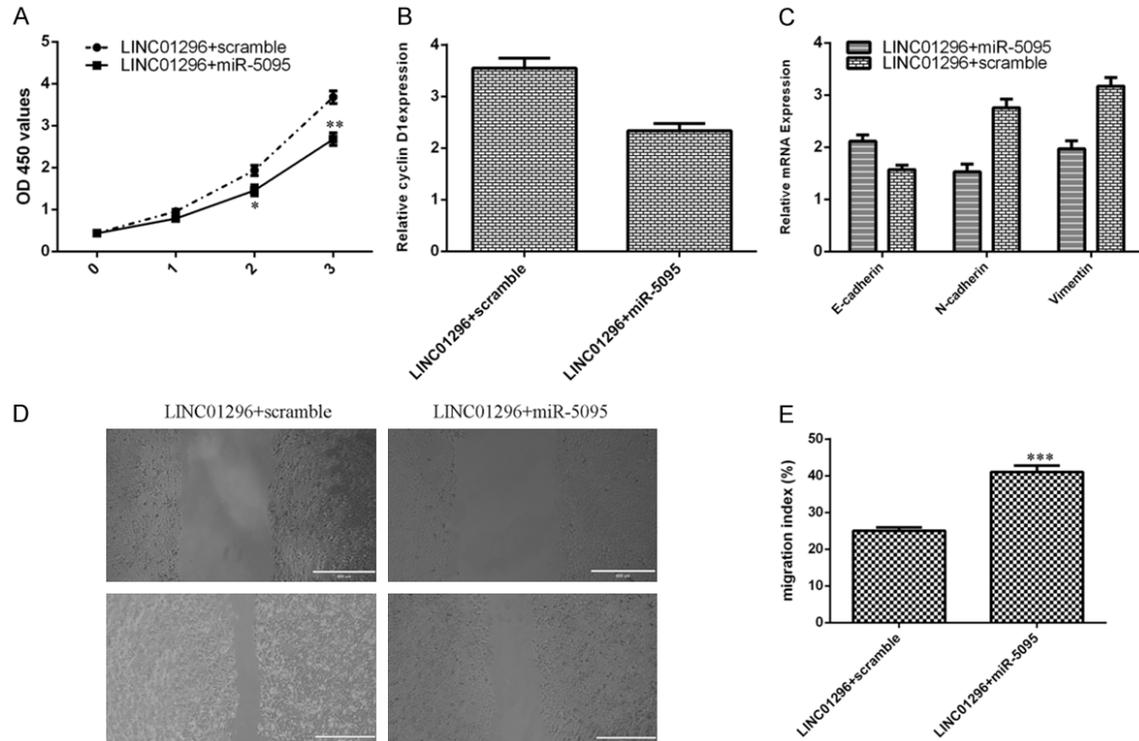


Figure 6. miR-5095 reversed the promoting effects of LINC01296 on NSCLC cells. A. Elevated expression of miR-5095 attenuated the inducing effect on cell proliferation of A549 cell via CCK-8 assay. B. The mRNA expression of cyclin D1 was detected by qRT-PCR. C. The mRNA expression of E-cadherin, N-cadherin and Vimentin in the A549 cell was detected by qRT-PCR. D. Elevated expression of miR-5095 attenuated the promoting effect on cell migration of A549 cell. E. The relative migration index was shown. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$.

Moreover, we demonstrated that LINC01296 has a potential binding site for miR-5095 by using online program tool StarBase. Overexpression of LINC01296 inhibited the expression of miR-5095 in the A549 cell. Furthermore, the miR-5095 expression was downregulated in the NSCLC tissues than in the adjacent non-tumor tissues. In addition, we found that there is a negative correlation between miR-5095 expression and LINC01296 level in the NSCLC tissues. Overexpression of miR-5095 suppressed NSCLC cell proliferation and migration. Finally, we demonstrate that ectopic expression of LINC01296 promoted cell proliferation and migration via inhibiting miR-5095 expression. These results suggested that LINC01296 might act a role as an oncogene in the tumorigenesis and development of NSCLC.

LINC01296 was shown to be involved in several tumors development such as colorectal cancer, bladder cancer, prostate cancer, gastric cancer, cholangiocarcinoma, esophageal squamous cell carcinoma (ESCC) and osteosarcoma [31-37]. For instance, Qiu et al. [33]. Found that

LINC01296 was overexpressed and the higher expression of LINC01296 was an independent predictor for colorectal cancer patients' prognosis. Seitz and colleagues performed the lncRNA profiling to found that LINC00958 was a candidate oncogene in the bladder cancer [34]. Qin et al. [37]. Showed that the LINC01296 expression was overexpressed in gastric cancer tissues and knockdown expression of LINC01296 inhibited gastric cancer cell migration, invasion and proliferation and induced cell apoptosis via regulating miR-122 expression. Zhang and colleagues found that LINC01296 expression was overexpressed in cholangiocarcinoma and the LINC01296 expression was positively correlated with clinical stage and tumor severity [32]. LINC01296 knockdown inhibited cell migration, invasion and viability. Yu et al. [36]. showed that LINC01296 expression was upregulated in osteosarcoma and the higher expression of LINC01296 was associated with poor survival. Overexpression of LINC01296 promoted osteosarcoma cell invasion, migration and proliferation. However, the expression pattern and functional role of LIN-

LINC01296 induces non-small cell lung cancer growth and progression

C01296 remains unclear. In this research, we firstly measured the LINC01296 expression in 35 NSCLC samples and adjacent non-tumor samples through performing qRT-PCR. We found that the expression of LINC01296 was upregulated in NSCLC tissues compared to adjacent non-tumor tissues. Ectopic expression of LINC01296 promoted NSCLC cell proliferation and migration. Furthermore, we demonstrated that elevated expression of LINC01296 induced the cyclin D1 expression in the A549 cell. Furthermore, LINC01296 overexpression suppressed the expression of E-cadherin and induced the expression of N-cadherin and Vimentin in the A549 cell.

Binding relationship between miR-5095 and LINC01296 was confirmed by our research, which helps us to further comprehend LINC01296's promoting of lung NSCLC progression via interacting with miR-5095. Previous studies also found that lncRNA act as sponges for miRNA [38, 39]. For example, Chi et al. [40] demonstrated that lncRNA RP11-79H23.3 acts as a ceRNA to modulate PTEN expression via sponging miR-107 expression in the bladder Cancer development. Li et al. [41]. Showed that lncRNA LINC00978 induced NSCLC cell invasion and proliferation through inhibiting miR-6754-5p. Moreover, Zhang et al. [32]. demonstrated that LINC01296 induced cholangiocarcinoma growth and progression through sponging miR-5095. We furtherly demonstrated that ectopic expression of miR-5095 reduced the luciferase reporter activity carrying the 3'-UTR of LINC01296 but not the mutant of 3'-UTR of LINC01296 In addition, ectopic expression of LINC01296 inhibited miR-5095 expression in the A549 cell. The miR-5095 expression was lower in the NSCLC tissues than in the adjacent non-tumor tissues. There is a negative correlation between miR-5095 expression and LINC01296 level in the NSCLC tissues. Overexpression of miR-5095 suppressed cell proliferation and migration. Finally, we demonstrate that ectopic expression of LINC01296 promoted cell proliferation and migration via inhibiting miR-5095 expression.

In summary, we found that the expression of LINC01296 was overexpressed in NSCLC samples compared to adjacent non-tumor tissues. Ectopic expression of LINC01296 promoted NSCLC cell proliferation and migration partly via inhibiting miR-5095 expression.

Acknowledgements

Supported by Medical guidance project of Shanghai Committee on Science and Technology (No. 134119a3400) and Rising Frontiers projects of ShenKang centre (SHDC12016106).

Disclosure of conflict of interest

None.

Address correspondence to: Drs. Hongcheng Liu and Lei Zhang, Department of Thoracic, Shanghai Pulmonary Hospital, Tongji University, Shanghai 200433, China. E-mail: lhczs@163.com (HCL); 13816121971@163.com (LZ)

References

- [1] Chen J, Zhang F, Wang JJ, Hu LJ, Chen J, Xu G and Wang YM. LncRNA LINC01512 promotes the progression and enhances oncogenic ability of lung adenocarcinoma. *J Cell Biochem* 2017; 118: 3102-3110.
- [2] She K, Yan H, Huang J, Zhou H and He J. miR-193b availability is antagonized by lncRNA-SNHG7 for FAIM2-induced tumour progression in non-small cell lung cancer. *Cell Prolif* 2018; 51.
- [3] Zhuang Y, Jiang HY, Li H, Dai JH, Liu Y, Li Y, Miao LY, Cai HR, Xiao YL, Xia HP, Wang YS and Shi MK. Down-regulation of long non-coding RNA AFAP1-AS1 inhibits tumor cell growth and invasion in lung adenocarcinoma. *Am J Transl Res* 2017; 9: 2997-3005.
- [4] Sun Y, Zhao J, Yin X, Yuan X, Guo J and Bi J. miR-297 acts as an oncogene by targeting GPC5 in lung adenocarcinoma. *Cell Prolif* 2016; 49: 636-643.
- [5] Huang CS, Liu SG, Wang HJ, Zhang ZC, Yang Q and Gao F. LncRNA PVT1 overexpression is a poor prognostic biomarker and regulates migration and invasion in small cell lung cancer. *Am J Transl Res* 2016; 8: 5025-5034.
- [6] Yao S, Zhao T and Jin H. Expression of MicroRNA-325-3p and its potential functions by targeting HMGB1 in non-small cell lung cancer. *Biomed Pharmacother* 2015; 70: 72-79.
- [7] Zhang ZY, Fu SL, Xu SQ, Zhou X, Liu XS, Xu YJ, Zhao JP and Wei S. By downregulating Ku80, hsa-miR-526b suppresses non-small cell lung cancer. *Oncotarget* 2015; 6: 1462-1477.
- [8] Zhu X, Gao G, Chu K, Yang X, Ren S, Li Y, Wu H, Huang Y and Zhou C. Inhibition of RAC1-GEF DOCK3 by miR-512-3p contributes to suppression of metastasis in non-small cell lung cancer. *Int J Biochem Cell Biol* 2015; 61: 103-114.
- [9] Zhang L, Xu B, Qiang Y, Huang H, Wang C, Li D and Qian J. Overexpression of deubiquitinating

LINC01296 induces non-small cell lung cancer growth and progression

- enzyme USP28 promoted non-small cell lung cancer growth. *J Cell Mol Med* 2015; 19: 799-805.
- [10] Zhang J, Xu L, Yang Z, Lu H, Hu D, Li W, Zhang Z, Liu B and Ma S. MicroRNA-10b indicates a poor prognosis of non-small cell lung cancer and targets E-cadherin. *Clin Transl Oncol* 2015; 17: 209-214.
- [11] Zhang B, Liu T, Wu T, Wang Z, Rao Z and Gao J. microRNA-137 functions as a tumor suppressor in human non-small cell lung cancer by targeting SLC22A18. *Int J Biol Macromol* 2015; 74: 111-118.
- [12] Yu X, Wei F, Yu J, Zhao H, Jia L, Ye Y, Du R, Ren X and Li H. Matrix metalloproteinase 13: a potential intermediate between low expression of microRNA-125b and increasing metastatic potential of non-small cell lung cancer. *Cancer Genet* 2015; 208: 76-84.
- [13] Yu SH, Zhang CL, Dong FS and Zhang YM. miR-99a suppresses the metastasis of human non-small cell lung cancer cells by targeting AKT1 signaling pathway. *J Cell Biochem* 2015; 116: 268-276.
- [14] Ye XW, Yu H, Jin YK, Jing XT, Xu M, Wan ZF and Zhang XY. miR-138 inhibits proliferation by targeting 3-phosphoinositide-dependent protein kinase-1 in non-small cell lung cancer cells. *Clin Respir J* 2015; 9: 27-33.
- [15] Ye M, Zhang J, Miao Q and Yao L. Curcumin promotes apoptosis by activating the p53-miR-192-5p/215-XIAP pathway in non-small cell lung cancer. *Cancer Lett* 2015; 357: 196-205.
- [16] Xu LM, Li LQ, Li J, Li HW, Shen QB, Ping JL, Ma ZH, Zhong J and Dai LC. Upregulation of MiR-1280 expression in non-small cell lung cancer tissues. *Chin Med J (Engl)* 2015; 128: 670-673.
- [17] Wu ZH, He YY, Li DL, Fang X, Shang T, Zhang HK and Zheng XT. Long noncoding RNA MEG3 suppressed endothelial cell proliferation and migration through regulating miR-21. *Am J Transl Res* 2017; 9: 3326-3335.
- [18] Zheng JL, Yi D, Liu Y, Wang MQ, Zhu YL and Shi HZ. Long noncoding RNA UCA1 regulates neural stem cell differentiation by controlling miR-1/Hes1 expression. *Am J Transl Res* 2017; 9: 3696-3704.
- [19] Zhang Y, Dang YW, Wang X, Yang X, Zhang R, Lv ZL and Chen G. Comprehensive analysis of long non-coding RNA PVT1 gene interaction regulatory network in hepatocellular carcinoma using gene microarray and bioinformatics. *Am J Transl Res* 2017; 9: 3904-3917.
- [20] Zhang SL, Dong XX, Ji TY, Chen GX and Shan L. Long non-coding RNA UCA1 promotes cell progression by acting as a competing endogenous RNA of ATF2 in prostate cancer. *Am J Transl Res* 2017; 9: 366-375.
- [21] Wang XM, Lu XB, Geng ZS, Yang GY and Shi Y. LncRNA PTCSC3/miR-574-5p governs cell proliferation and migration of papillary thyroid carcinoma via Wnt/-Catenin signaling. *J Cell Biochem* 2017; 118: 4745-4752.
- [22] Wang XB, Lv GH, Li J, Wang B, Zhang QS and Lu C. LncRNA-RP11-296A18.3/miR-138/HIF1A Pathway regulates the proliferation ECM synthesis of human nucleus pulposus cells (HNPCs). *J Cell Biochem* 2017; 118: 4862-4871.
- [23] Tong GL, Wu X, Cheng BR, Li L, Li X, Li Z, Nong QH, Chen XQ, Liu YJ and Wang SB. Knockdown of HOXA-AS2 suppresses proliferation and induces apoptosis in colorectal cancer. *Am J Transl Res* 2017; 9: 4545-4552.
- [24] Tan XY, Huang ZG and Li XG. Long non-coding RNA MALAT1 interacts With miR-204 to modulate human hilar cholangiocarcinoma proliferation, migration, and invasion by targeting CXCR4. *J Cell Biochem* 2017; 118: 3643-3653.
- [25] Li J, Lian YF, Yan CS, Cai ZL, Ding J, Ma ZH, Peng P and Wang KM. Long non-coding RNA FOXP4-AS1 is an unfavourable prognostic factor and regulates proliferation and apoptosis in colorectal cancer. *Cell Prolif* 2017; 50.
- [26] Chen LS, Yao HB, Wang K and Liu XF. Long non-coding RNA MALAT1 regulates ZEB1 expression by sponging miR-143-3p and promotes hepatocellular carcinoma progression. *J Cell Biochem* 2017; 118: 4836-4843.
- [27] Zhao HX, Hou WG, Tao JG, Zhao YL, Wan G, Ma C and Xu HB. Upregulation of lncRNA HNF1A-AS1 promotes cell proliferation and metastasis in osteosarcoma through activation of the Wnt/beta-catenin signaling pathway. *Am J Transl Res* 2016; 8: 3503-3512.
- [28] Li P, Xue WJ, Feng Y and Mao QS. Long non-coding RNA CASC2 suppresses the proliferation of gastric cancer cells by regulating the MAPK signaling pathway. *Am J Transl Res* 2016; 8: 3522-3529.
- [29] Qiu JJ, Lin YY, Ding JX, Feng WW, Jin HY and Hua KQ. Long non-coding RNA ANRIL predicts poor prognosis and promotes invasion/metastasis in serous ovarian cancer. *Int J Oncol* 2015; 46: 2497-2505.
- [30] Xu R, Mao YQ, Chen KB, He W, Shi WJ and Han Y. The long noncoding RNA ANRIL acts as an oncogene and contributes to paclitaxel resistance of lung adenocarcinoma A549 cells. *Oncotarget* 2017; 8: 39177-39184.
- [31] Wu J, Cheng G, Zhang C, Zheng YX, Xu HX, Yang HW and Hua LX. Long noncoding RNA LINC01296 is associated with poor prognosis

LINC01296 induces non-small cell lung cancer growth and progression

- in prostate cancer and promotes cancer-cell proliferation and metastasis. *Onco Targets Ther* 2017; 10: 1843-1852.
- [32] Zhang DW, Li HY, Xie JP, Jiang DC, Cao LQ, Yang XW, Xue P and Jiang XF. Long noncoding RNA LINC01296 promotes tumor growth and progression by sponging miR-5095 in human cholangiocarcinoma. *Int J Oncol* 2018; 52: 1777-1786.
- [33] Qiu JJ and Yan JB. Long non-coding RNA LINC01296 is a potential prognostic biomarker in patients with colorectal cancer. *Tumor Biol* 2015; 36: 7175-7183.
- [34] Seitz AK, Christensen LL, Christensen E, Faarkrog K, Ostenfeld MS, Hedegaard J, Nordentoft I, Nielsen MM, Palmfeldt J, Thomson M, Jensen MTS, Nawroth R, Maurer T, Orntoft TF, Jensen JB, Damgaard CK and Dyrskjot L. Profiling of long non-coding RNAs identifies LINC00958 and LINC01296 as candidate oncogenes in bladder cancer. *Sci Rep* 2017; 7: 395.
- [35] Wang B, Liang T and Li J. Long noncoding RNA LINC01296 is associated with poor prognosis in ESCC and promotes ESCC cell proliferation, migration and invasion. *Eur Rev Med Pharmacol Sci* 2018; 22: 4524-4531.
- [36] Yu X, Pang L, Yang T and Liu P. lncRNA LINC01296 regulates the proliferation, metastasis and cell cycle of osteosarcoma through cyclin D1. *Oncol Rep* 2018; 40: 2507-2514.
- [37] Qin Q, Yin Z, Li Y, Wang B and Zhang M. Long intergenic noncoding RNA 01296 aggravates gastric cancer cells progress through miR-122/MMP-9. *Biomed Pharmacother* 2018; 97: 450-457.
- [38] Ye KS, Wang SK, Zhang HH, Han H, Ma B and Nan W. Long noncoding RNA GAS5 suppresses cell growth and epithelial-mesenchymal transition in osteosarcoma by regulating the miR-221/ARHI pathway. *J Cell Biochem* 2017; 118: 4772-4781.
- [39] Liu J, Song ZW, Feng C, Lu YL, Zhou Y, Lin Y and Dong CY. The long non-coding RNA SUMO1P3 facilitates breast cancer progression by negatively regulating miR-320a. *Am J Transl Res* 2017; 9: 5594-5602.
- [40] Chi H, Yang R, Zheng X, Zhang L, Jiang R and Chen J. LncRNA RP11-79H23.3 functions as a competing endogenous RNA to regulate PTEN expression through sponging hsa-miR-107 in the development of bladder cancer. *Int J Mol Sci* 2018; 19.
- [41] Li X, Ren Y and Zuo T. Long noncoding RNA LINC00978 promotes cell proliferation and invasion in non-small cell lung cancer by inhibiting miR-6754-5p. *Mol Med Rep* 2018; 18: 4725-4732.