

The multiple faces of CXCL12 (SDF-1 α) in the regulation of immunity during health and disease

Nathan Karin¹

Department of Immunology, Bruce Rappaport Faculty of Medicine and Rappaport Family Institute for Research in the Medical Sciences, Technion–Israel Institute of Technology, Haifa, Israel

RECEIVED SEPTEMBER 8, 2009; REVISED MAY 5, 2010; ACCEPTED MAY 5, 2010. DOI: 10.1189/jlb.09090602

ABSTRACT

Chemokines are a group of small, structurally related molecules that regulate the trafficking of various types of leukocytes through interactions with a subset of 7-transmembrane G-protein-coupled receptors. As key chemoattractants of inflammatory leukocytes, chemokines have been marked as potential targets for neutralization in autoimmune diseases. Cancer cells also express chemokines, where they function as survival/growth factors and/or angiogenic factors that promote tumor development and angiogenesis. Accordingly, these functions make them attractive targets for therapy of these diseases. Recently, we reported that one of these chemokines CXCL12 (SDF-1 α) functions as an anti-inflammatory chemokine during autoimmune inflammatory responses and explored the mechanistic basis of this function. As a pleiotropic chemokine, CXCL12 participates in the regulation of tissue homeostasis, immune surveillance, autoimmunity, and cancer. This chemokine is constitutively expressed in the BM and various tissues, which enables it to regulate the trafficking and localization of immature and maturing leukocytes, including BM stem cells, neutrophils, T cells, and monocytic cells. We have shown recently that CXCL12 increases immunological tolerance in autoimmune diseases by polarizing Tregs and by doing so, restrains the progression of these diseases. This finding suggests a possible use of stabilized rCXCL12 as a potential drug for therapy of these diseases and targeted neutralization of CXCL12 for therapy of cancer diseases. The current review explores the different biological properties of CXCL12 and discusses the implications of CXCL12-based therapies for autoimmunity and cancer diseases. *J. Leukoc. Biol.* **88**: 463–473; 2010.

Introduction

Chemokines are a group of small (8–14 kDa), structurally related molecules that regulate the trafficking of various types of leukocytes through interactions with a subset of 7-transmembrane G-protein-coupled receptors [1–3]. Chemokines play fundamental roles in the development, homeostasis, and function of the immune system [1–3]. Although chemokines are produced primarily by immune cells, they are also produced by nonimmune cells and affect their biological function(s). For example, vascular endothelial cells produce chemokines that are involved in angiogenesis [1–3].

Based on the arrangement of the conserved cysteine residues, the chemokines are divided into four subgroups: C, CC, CXC, and CX3C [1–3]. The two major subgroups are the CXC and CC subgroups, in which the two cysteines are separated by a single amino acid (CXC) or are adjacent to each other (CC). The CXC chemokines are subdivided further into ELR and non-ELR types, according to the presence or absence of a ELR tripeptide sequence adjacent to the CXC motif [1–3].

Much attention has been paid to the key role of chemokines in inflammatory processes [4–10] and especially, inflammatory autoimmune diseases, such as MS and its experimental models [6, 8, 11–22], T1DM [23, 24] and RA [25–27], amongst others. As key chemoattractants of inflammatory leukocytes, chemokines have been marked as potential targets for neutralization in autoimmune diseases [6, 8, 11–22, 24–27]. Some of the results of these studies have been applied to human patients, thus far with limited success.

EAE is a T cell-mediated autoimmune disease of the CNS, and animals in which EAE is induced are used as an experimental model for MS. One of the chemokines that is likely to participate in the regulation of EAE is CXCL12 (SDF-1 α). This is a pleiotropic chemokine that participates in the regulation of tissue homeostasis, immune surveillance, inflammatory responses, and cancer development. Under noninflammatory conditions, CXCL12 attracts leukocytes to the CNS as a part of

Abbreviations: BM=bone marrow, EAE=experimental autoimmune encephalomyelitis, ELR=Glu-Leu-Arg, Foxp3=forkhead box p3, HEV=high endothelial venule, MS=multiple sclerosis, RA=rheumatoid arthritis, SDF-1 α =stromal cell-derived factor 1 α , T1DM=type I diabetes mellitus, Thnp=nonpolarized CD4⁺ T cell(s), Tr-1=regulatory T cell I, Treg=regulatory T cell

1. Correspondence: Department of Immunology, Bruce Rappaport Faculty of Medicine and Rappaport Family Institute for Research in the Medical Sciences, Technion–Israel Institute of Technology, 1 Efron St., Haifa 31096, Israel. E-mail: nkarin@tx.technion.ac.il

the immune surveillance of tissues, thereby functioning as a “proinflammatory” chemokine. In a recent study, we showed that when leukocytes rapidly enter a site of inflammation in the CNS during ongoing EAE, this chemokine shifts its potential proinflammatory function to become an anti-inflammatory chemokine [28]. This change skews the polarization of CD4⁺ antigen-specific effector T cells (also known as Th1 and Th17 cells), which normally promote inflammation, so that they become antigen-specific Tregs that produce a key anti-inflammatory cytokine, IL-10 [28], and restrain the development and progression of the disease. In light of these findings, the present article reviews the multiple faces of CXCL12 (SDF-1 α) in the regulation of immunity during health and disease and their therapeutic implications for cancer and autoimmunity.

THE ALTERNATIVE ROLE OF CHEMOKINES IN TISSUE HOMEOSTASIS, IMMUNE SURVEILLANCE, AUTOIMMUNITY, AND CANCER

Chemokines direct cellular movement and relocation, both of which are essential for many fundamental physiologic processes, which include embryonic development, neovascularization and angiogenesis, immunologic responses, wound healing, and organ repair. In embryonic life, together with other chemoattractive growth factors, CXCL12 directs the proliferation and differentiation of immature progenitor cells [29] by activation of the adhesion machinery, cytoskeleton rearrangement, the control of cell cycle, and the secretion of proteolytic enzymes [29]. In adults, chemokines have three major functions: They control tissue homeostasis; they direct the development of inflammatory responses, including inflammatory autoimmune diseases; and they function as growth/survival factors and as chemoattractants for cancer cells to support metastasis. For tissue homeostasis, chemokines are involved closely in the regulation of cell migration, particularly leukocytes, from the BM to blood and tissues, to ensure a dynamic balance of cell number and type in these organs under diverse conditions. For example, the interaction between the CC chemokine receptor, CCR2, which is expressed predominantly on BM-derived monocytes, and its ligands, particularly CCL2, is essential for directing the rapid recruitment of CCR2⁺ cells from the BM to a site of inflammation, where they are required to promote inflammatory processes [30]. Thus, CCR2-deficient mice are highly resistant to the induction of inflammatory autoimmunity [20]. Another example for the role of CXCL12 in tissue homeostasis is its constitutive expression in the BM and other tissues, which include the skin and heart, epithelial cells in human bile ducts, and brain endothelium [29]. This constitutive expression is responsible for regulating the trafficking and localization of immature and maturing leukocytes to these tissues. During steady-state conditions, these cells egress continuously from the BM reservoir to the circulation to replenish the blood with new cells throughout the lifespan of the individual [31]. In addition to its role in regulating BM stem cell homeostasis, CXCL12 is involved directly in neutrophil homeostasis under normal and stress

conditions. The number of circulating neutrophils is tightly regulated to ensure adequate protection of the host against microbial pathogens and concomitantly minimizing damage to noninflamed tissues. Neutrophil homeostasis in the blood is achieved by balancing neutrophil production, neutrophil release from the BM, and neutrophil clearance from the circulation. Accumulating evidence suggests that signaling by CXCL12, through its major receptor CXCR4, plays an important role in maintaining this homeostasis [32]. Very recently, Eash et al. [33] generated mice with a myeloid lineage-restricted deletion of CXCR4 to define the underlying mechanisms of CXCR4 signaling in neutrophil homeostasis. Similar to its effects on other leukocytes, particularly monocytes and T cells, CXCL12 directs the migration of neutrophils to organs, particularly immune-privileged organs, as part of the immune surveillance of the body. Based on observations that have been made in experimental models of autoimmune disease, whose results have been extended to humans, it is believed that the interaction between antigen-specific effector T cells and their target autoimmune antigen in T cell-mediated inflammatory autoimmune disease following tolerance breakdown begins with the activation of HEVs. Their activation then facilitates the extravasation of large numbers of lymphocytes and monocytes from the blood to the tissues to initiate the autoimmune process [34]. Inflammatory chemokines possess three key complementary features in this process: They attract various types of leukocytes, particularly lymphocytes, monocytes, and neutrophils to the inflammatory site; they activate adhesion molecules on HEV and leukocytes to allow rapid influx of cells to the autoimmune site [35–40]; and they polarize CD4⁺ T cells directly to become effector proinflammatory cells [26, 41]. Thus, not only inflammatory chemokines but also the key adhesion receptors that they activate, namely the α 4 β 1-integrin, VLA-4, have become a major target in the therapy of MS and other autoimmune diseases [42–44].

The role of chemokines in tumor biology is complex. It appears that many tumor cells produce a large variety of chemokines and their receptors [45–47]. The actions of chemokines in tumor biology can be divided into direct autocrine effects, in which tumor cells produce a given chemokine and its target receptor, and indirect effects, such as recruitment of tumor-associated macrophages, to support tumor development and angiogenesis [45, 48–51]. On their autocrine effects, chemokines function as growth/survival factors for cancer cells, as proangiogenic factors, and as chemoattractants for cancer cells to support metastasis. At least three major chemokines, CXCL8, CXCL12, and CCL2, all of which are produced by various solid tumors, such as malignant melanoma, liver and pancreatic tumors, colon carcinoma, breast cancer, and prostate cancer, function as autocrine growth factors [45–47]. It has been proposed that CXC chemokines with the ELR tripeptide sequence immediately adjacent to the amino-terminal of the CXC motif (ELR⁺ CXC chemokines), such as CXCL8, are angiogenic and that ELR⁻ CXC chemokines, such as CXCL10, are angiostatic [1–3]. Other data have shown that the ELR⁻ CXC chemokine, CXCL12, can act as a direct chemoattractant for endothelial cells in vitro and as an angiogenic factor in vivo [52–54]. CXCL12 is also thought to play a key role in at-

tracting tumor cells to their target organs to facilitate the formation of metastasis, such as the attraction of prostate cancer cells to the bones [55, 56]. On the indirect effects of chemokines, chemokines, such as CCL2, are thought to be highly important for attracting CCR2⁺ monocytic cells that assist tumor development, rather than destroying tumors, by producing angiogenic factors at the tumor site and producing cytokines, particularly IL-10, that suppresses anti-tumor immunity [45, 48, 49, 51, 57–62].

THE MAJOR PARADOX OF CHEMOKINES AS TARGETS FOR THERAPY OF INFLAMMATORY AUTOIMMUNE DISEASES

The underlying idea of antichemokine therapies for inflammatory autoimmune disease is that targeted neutralization of chemokines would restrain the autoimmune response by inhibiting the influx of inflammatory leukocytes to a site of autoimmune-induced inflammation. Reviewing the results of the many studies that have used this strategy reveals a major paradox. Although most of the 50 known chemokines can direct the migration of the same leukocytes, targeted neutralization of only one chemokine, such as CCL2, CCL3, CCL5, or CXCL10, is sufficient to suppress the entire inflammatory process [8, 20, 21, 41, 63–67]. Therefore, the question that begs an answer is why other chemokines that also attract the same type of leukocyte to the autoimmune site do not compensate for the absence of this single chemokine. In addition, it is not clear why neutralization of as few as eight to 10 of the 50 different chemokines can effectively suppress the attacks in autoimmune inflammatory diseases [8, 20, 21, 41, 63–67]. Hence, what are the attributes of this limited number of chemokines that make them so important in the regulation of inflammatory processes?

A partial explanation for this paradox could be that these chemokines have other biological actions that are associated with these autoimmune inflammatory diseases, as well as being chemoattractants. For example, it has been shown recently that CCR5 ligands are essential for the induction of costimulatory signals via stimulation of the CCR5 receptor on target effector CD4⁺ T cells and that these signals are essential to promote IL-2-dependent activation of these cells [68–70]. This may explain why targeted neutralization of CCR5 ligands is so effective in suppressing different autoimmune diseases, such as EAE [65, 71, 72], RA [25, 73], and T1DM, in different strains of mice [24].

EFFECTOR CD4⁺ T CELL SUBSETS

In 1986, Robert Coffman and Timothy Mosmann were the first to describe the division of CD4⁺ T cells into functional subsets based on cytokine production, which they termed Th1 and Th2 cells [74, 75]. Subsequently, other types of effector T cell and Treg subsets were identified. It has also been shown that during their activation, Thnp undergo epigenetic changes when differentiating into different subtypes. These epigenetic

changes are dependent on the cytokine milieu that is present within the microenvironment, in which they are undergoing clonal expansion [76, 77]. For example, in the presence of high levels of IL-12 and IL-18 and low levels of IL-4, Thnp will be polarized preferentially into IFN- γ ^{high}IL-4^{low}-producing Th1 cells that also produce substantial levels of IL-2, TNF- α , and other proinflammatory cytokines [78–81]. On the other hand, a milieu that is enriched with IL-4 and has low levels of IL-12 and IL-18 will drive the polarization of Thnp to become IL-4-producing Th2 cells [82, 83]. Although Th1 cells promote the inflammatory process, Th2 cells are associated with helping B cells switch their isotype and change their biological function [82, 83]. Thus, from the practical perspective, these observations suggest that anti-IL-12 and/or anti-IL-18 therapies may be beneficial in inflammatory autoimmunity. Indeed, the administration of neutralizing antibodies to IL-12 or IL-18 could suppress several experimentally induced autoimmune diseases effectively by shifting the Th1/Th2 cell balance in favor of Th2 cells [81, 84–87]. The drawback of this therapeutic strategy is the danger of substituting a Th1-driven inflammatory process with a Th2-directed allergic response [88].

A third subtype of CD4⁺ effector cell is the recently discovered IL-17-producing Th17 cell [89–93]. These cells are polarized initially by IL-6 and TGF- β and thereafter, by IL-21 in an autocrine way via the STAT3 pathway. This initial stepwise polarization increases their susceptibility to IL-23, which augments their rapid polarization [89–93]. IL-23-deficient mice are EAE-resistant, and therapies that neutralize this cytokine suppress EAE and other T cell-mediated autoimmune diseases [93–100]. It is also believed that Th1 and Th17 cells contribute independently to CD4⁺-directed inflammatory responses that include T cell-mediated autoimmunity by different mechanisms [99, 100].

Are there other subtypes of effector CD4⁺ T cells yet to be identified? The basic concept, which was proposed by Mosmann and colleagues, is that the Thnp could be divided into two exclusive types of effector cells, namely the IFN- γ ^{high}IL-4^{low} Th1 cells and the IL-4^{high}IFN- γ ^{low} Th2 cells, and that the lineage characteristics of each subtype are stable for as long as 20 years. Hence, the discovery of other types of effector T cells, such as Th17 cells [89–93], and the recent observations that challenge the dogma that these subsets are stable [101–103], which is discussed later in this review, have unwittingly opened a Pandora's box of complexity and controversy. As for other effector T cell subtypes, two new subtypes have been discovered recently: one subtype that produces IL-22 (Th22 cell) predominantly and a second subtype that produces IL-9 (Th9 cells) [104, 105]. Future investigations may well lead to the discovery of other new effector T cell subtypes.

TREG SUBSETS

Despite major functional differences among Th1, Th17, and Th2 cells, all are considered to be effector T cells that can promote inflammatory responses (Th1 and Th17 cells) or B cell-directed immunity (Th2 cells). What regulates their activities? As mentioned already, the CD4⁺ T cell subsets that control the function of effector T cells are known as Tregs, and

these subsets can be divided into two major categories: those that express Foxp3 (Foxp3⁺) and are known as natural Tregs [106, 107] and the antigen-specific Tregs, which are Foxp3⁻ and suppress the function of effector T cells by producing suppressor cytokines, such as TGF- β [108, 109], IL-10 [110], and possibly IL-35 [111]. Foxp3⁺ Tregs were discovered by Shimon Sakaguchi [106], who showed that their depletion leads to the spontaneous development of various autoimmune diseases, which could be reversed by adoptive transfer of natural Tregs from naïve donors. The relevance of these findings has been extended to humans. Similar to the T cell subsets in rodents, the human T cell subsets also include a significant portion of CD4⁺CD25⁺Foxp3⁺ T cells that functions as suppressor Tregs. Interestingly, human subjects who carry a functional mutation in the *FOXP3* gene spontaneously develop a severe autoimmune disorder that is called the immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome [112] and resembles the syndrome that develops when mice are depleted of CD4⁺CD25⁺Foxp3⁺ T cells [113]. For Foxp3⁻ Tregs, two major subtypes have been identified thus far. Almost 20 years ago, Howard Weiner and his group [108, 109] identified a novel type of antigen-specific Tregs that produces large amounts of the suppressor cytokine/growth factor TGF- β and small amounts of other cytokines and subsequently, named them Th3 cells. Weiner and his group [108, 109] showed that these Th3 cells seem to direct immunological tolerance within the gut, and their function could be induced by oral tolerance. In 1977, Groux et al. [110] identified another type of antigen-specific Tregs that produces mostly IL-10 and designated them as Tr-1 cells. It is believed that these Tr-1 cells are essential in the maintenance of active tolerance and in particular, for restraining the harmful activity of autoimmune T cells [114].

FLEXIBILITY IN THE POLARIZATION OF CD4⁺ T CELL SUBSETS

The traditional, consensual opinion about the mechanism of T cell polarization into subsets assumed that epigenetic changes lead to a well-defined characteristic of each CD4⁺ T cell subset with relative inflexibility [115]. For example, during their antigen-specific T cell activation, Thnp are polarized into Th1 cells in the presence of high levels of IL-12 and low levels of IL-4 [78]. Under these conditions, IL-12 activates a specific signal transduction cascade that leads to the synthesis of a T-box transcription factor, which controls the expression of the hallmark Th1 cytokine, IFN- γ [116]. According to the traditional dogma, these polarized CD4⁺ T cells would continue producing their typical cytokine profile as long as they continue to proliferate in response to their target antigen. For example, proliferating effector Th1 cells would continue producing high levels of IFN- γ and TNF- α and low levels of IL-4 as long as they are activated. Thereafter, these cells undergo apoptosis or become long-term memory cells [117]. Once becoming long-term memory cells, they arrest their typical cytokine production and enter a resting phase. Upon reactivation, it is believed that upon reactivation, they “remember” their basic cytokine profile that they obtained during their initial

polarization and continue producing this cytokine profile. Other examples are: Th2 cell polarization, which is directed initially by IL-4-induced signaling via the STAT6 and STAT5 transcription factors [118] and then by the GATA-3 transcription factor [119], and Th17 cells, which are polarized initially by IL-6 and TGF- β and thereafter, in an autocrine way by IL-21 via the STAT3 pathway and the major nuclear orphan receptor ROR γ t [120], and the resultant polarization increases their susceptibility to further polarization by IL-23 [89–93].

For Foxp3⁺ Tregs, the key transcription factor that characterizes the CD4⁺CD25⁺ Tregs is Foxp3, which is essential for their normal development and function [106, 121–123]. The dominant cytokine that drives the polarization of Foxp3⁻ to Foxp3⁺ CD4⁺ T cells is TGF- β [124, 125]. Less is known about the signal transduction events by which Foxp3⁻ IL-10-producing Tr-1 cells are polarized in the presence of IL-10 and IL-2 [110]. A recent study showed that this polarization is directed in human Tregs through a STAT5-responsive intronic enhancer in the IL-10 locus [126].

The results of several recent studies have now provided evidence that challenges the traditional dogma, which suggests that CD4⁺ T cells undergo epigenetic changes during T cell polarization and display limited flexibility. In 2007, Anderson et al. [101] showed that IFN- γ -producing Th1 cells could be repolarized into IL-10-producing Tr-1 cells in chronic, cutaneous leishmaniasis (see also a recent review by O’Garra and Vieira [103]). A more recent example is the role of another cytokine, IL-27, in the polarization of CD4⁺ T cells. Together with IL-12, IL-27 can polarize Th1 cells [127–129], and then together with IL-6, it can transform polarized Th1 cells, Th2 cells, and even Th17 cells into IL-10-producing Tr-1 cells [130]. These effects are dependent on the transcription factors STAT1 and STAT3. The polarization of CD4⁺ T cells into Th1 cells by IL-12 and IL-27 is dependent on STAT1 and STAT3, whereas the transformation of polarized Th1 cells by IL-6 and IL-27 is dependent on STAT3 [130–132]. Interestingly, no cytokines that are capable of reversing polarized Tr-1 cells into effector Th1 or Th17 cells have been found so far. We have shown recently that flexibility in skewing Th1 to Tr-1 could also be induced by CXCL12 via stimulation of the CXCR4 receptor on Th1 cells [28].

THE ROLE OF CHEMOKINES IN DIRECTING CD4⁺ T CELL POLARIZATION AND FUNCTION

The role of chemokines in directing the biological function of CD4⁺ T cells can be viewed from two perspectives: the differential effect of chemokines on the biological functions of polarized T cells and the ability of chemokines to polarize CD4⁺ T cells directly into selective subsets. Of the two roles, more is known about the role of chemokines on directing the biological properties of polarized T cells. In 1997, Sallusto and co-workers [133] showed that Th2 cells preferentially express the chemokine receptor, CCR3. In the following year, the same group extended this study to show that Th1 cells also express CXCR3, and Th2 cells preferentially express the chemokine receptors, CCR3 and CCR4 [134]. Th1 and Th2 cells express

CCR5 [133–135]. Subsequently, Hirahara et al. [136] reported that Foxp3⁺ Tregs also preferentially express CCR4. The direct consequence of these findings refers to the selective homing of various cell types as a result of their chemokine receptor expression. For example, Th17 cells predominantly express the chemokine receptor, CCR6, and its exclusive ligand CCL20 is constitutively expressed in the epithelial cells of choroid plexus in mice and humans to direct the homing of Th17 cells [137]. The results of a recent study have shown that an attenuated form of disease develops as a result of aberrant trafficking of Th17 cells when EAE is induced in CCR6-deficient mice [137]. The consequences of the CCL20-CCR6 interaction on the biological properties of Th17 cells are not known yet and still need to be studied. Another example is the CXCR3 ligand, CXCL10, which is preferentially expressed in the CNS during the accelerating phase of EAE and is associated with enhanced recruitment of CXCR3⁺ Th1 cells to the CNS [21].

In addition to directing the migratory properties of CD4⁺ T cells, the selective interaction of chemokines and their receptors affects their biological properties. For example, the chemokine receptor, CCR8, is preferentially expressed on CD45⁺ Th2 cells and Foxp3⁺ Tregs [138], and Foxp3⁺ Tregs suppress effector T cell functions by various mechanisms [107], and Th2 cells produce IL-4, which also restrains Th1 activities [80]. It has been suggested that Th2 and Foxp3⁺ T cells are not only attracted to the site of inflammation via the CCR8 receptor but also, that the interaction of CCL1 with its exclusive CCR8 receptor potentiates their anti-inflammatory activities [138], although this possibility has yet to be explored. Another example is the interaction between the CCR5 receptor that is expressed on Th1 cells and Th2 cells and its three different ligands, CCL3 (MIP- α), CCL4 (MIP- β), and CCL5 (RANTES) [1]. Recently, Molon et al. [68] reported that signals that are mediated by CCL5 by stimulation of the CCR5 receptor are required for effective activation of the CCR5 receptor, thus serving as costimulatory signals in T cell activation. Subsequently, Camargo et al. [70] showed that the interaction between the CCR5 receptor and its ligands induces signals via translocation of the transcription factor, NFAT, which results in IL-2 production by CD4⁺ T cells.

Less is known about the role of chemokines in the polarization of CD4⁺ T cells. CXCR3 is a chemokine receptor that is expressed mainly on CD4⁺ cells and in particular, Th1 cells [134], and three ligands bind to this receptor: CXCL9 (monokine induced by IFN- γ), CXCL10 (IFN-inducible protein 10), and CXCL11 (IFN-inducible T cell- α chemoattractant). In 1998, Gangur et al. [139] reported that the addition of CXCL10 to proliferating human T cells polarizes them into IFN- γ ^{high}IL-4^{low} Th1 cells. Based on this information, we have shown that indeed, CXCL10 not only attracts CXCR3⁺ T cells to the site of inflammation but also polarizes them into effector Th1 cells in two different experimental models of inflammatory autoimmune disease, namely EAE and adjuvant-induced arthritis [26, 41]. Thus, targeted neutralization of CXCL10 reduces the severity of ongoing EAE and adjuvant-induced arthritis by shifting the Th1/Th2 cell balance in favor of the Th2 cells [26, 41]. More recently, Flanagan et al. [140] reported that the lymphoid chemokine, CCL21, polarizes na-

ive T cells into Th1 cells. In our recent publication, we reported that chemokines may polarize some CD4⁺ subsets, and this finding supports the notion that competence is not limited only to effector cell polarization but also for skewing the polarization of effector T cells into Tregs [28].

THE MULTIPLE FACES OF CXCL12 DURING ONGOING INFLAMMATORY AUTOIMMUNITY

The CXC chemokine, CXCL12, which is also known as SDF-1 [141], was identified originally as a growth factor for murine pre-B cells [142]. It exists in two alternative splice variants: SDF-1 α and SDF-1 β . It is expressed constitutively by various cells and exhibits chemoattractive activity for monocytes, BM neutrophils, and early-stage B cell precursors, as well as being a highly efficient and potent chemoattractant for T cells and a costimulator of their activation [143]. Furthermore, CXCL12 induces adhesion of T cells to ICAM-1 (CD54) [144] by up-regulating the binding activity of LFA-1 (CD11a/CD18) and modulates the adhesion of α 4 β 7 integrin-mediated lymphocytes to mucosal addressin cell adhesion molecule-1 and fibronectin [145]. In light of these findings, it is thought that CXCL12 plays an important role in the attraction of T cells to specific sites. Furthermore, it was also suggested that CXCL12 could play a proinflammatory role in various autoimmune diseases, particularly RA and nephritis, in murine lupus erythematosus and therefore, could be a valid target for neutralization in these diseases [35, 146]. Our findings showing that CXCL12 functions as an anti-inflammatory chemokine during the inflammatory process challenge this concept [28].

In the healthy CNS, CXCL12 serves as a survival and migratory factor for neural and oligodendrocyte precursors that express the CXCR4 receptor [147]. The expression of CXCL12 within the CNS is up-regulated in the astrocytes of the MS brain; hence, its role in the regulation of this disease is not fully understood [148–150].

Our working hypothesis has been that targeted neutralization of CXCL12 during EAE would suppress the disease. In initial experiments, we tried to neutralize CXCL12 at different time-points after active EAE was induced and found contradictory effects. When a CXCL12-Ig fusion protein was administered to mice after the induction but before the onset of symptoms of EAE, we showed that the onset of disease was delayed for 2–3 days (Moran Meiron, Yaniv Zohar, and N. Karin, unpublished data). In contrast, there was rapid remission of the disease when the fusion protein was administered after the onset of disease [28].

The delay of the onset of disease following early administration of the CXCL12-Ig fusion protein could be explained by CXCL12 acting as a chemoattractant of leukocytes that are essential for the regular “policing” of the CNS. It is likely that endogenous CXCL12, which is produced within the CNS by astrocytes and other cells, is involved in modulating the migration of those leukocytes that are essential for the regular policing of the CNS. The development of progressive, multifocal leukoencephalopathy following anti- α 4-integrin (VLA-4) ther-

apy [151] may serve as an example for the importance of policing the CNS by T cells and macrophages.

However, according our observations, when the inflammatory process within the CNS later enters into an accelerating phase, CXCL12 functions as an anti-inflammatory chemokine that directs the polarization of CD4⁺ T cells and macrophages to become IL-10^{high}-producing Tregs [28]. The implication of these results for humans is that CXCL12 could be used as a potential drug only during advanced stages of inflammatory autoimmunity because of its pleiotropic characteristics.

The results of several studies have shown that C-C chemokines may affect the polarization of antigen-specific T cells by altering the Th1/Th2 cell balance [6, 152, 153]. However, the results of our recent study have shown that a chemokine can direct the polarization of IL-10-producing Tregs [28], thereby introducing a new concept that proposes chemokines as potential anti-inflammatory proteins. Are there other chemokines that polarize Tr-1 cells? We have observed recently that of the three CXCR3 ligands, CXCL9, CXCL10, and CXCL11, only CXCL11, which competes effectively with CXCL10 in binding their common CXCR3 receptor, antagonizes the function of the other CXCR3 ligands by skewing CD4⁺ T cell polarization into IL-10-producing Tr-1 cells (Yaniv Zohar, Gizi Wildbaum, Nir Netzer, and N. Karin, manuscript in preparation). CXCR3-deficient mice can develop an extremely severe form of EAE and T1DM [154, 155]. We do not exclude the possibility that the exacerbation of these diseases in experimental models is, in part, a result of the lack of protective, anti-inflammatory signals, which are mediated by the CXCL11-CXCR3 interactions.

CXCL12 POLARIZES ANTIGEN-SPECIFIC TR-1 CELLS BY TWO COMPLEMENTARY MECHANISMS

Antigen-specific Tr-1 cells were first identified by Groux et al. [110], who showed that the addition of IL-10 and IL-2 to cultured, primary T cells caused them to proliferate in response to their target antigen and polarize them into Tr-1 cells. Under these working conditions (in vitro) and at an inflammatory site (in vivo), IL-10 is produced by two major types of cells: the APCs that include dendritic cells (also known as D2 cells) and monocytes (also known as M2 cells) [61, 156] and the IL-10-producing Tr-1 cells [110]. We have shown that CXCL12 via stimulation of the CXCR4 receptor induces IL-10 production in macrophages and also acts directly on CD4⁺ T cells that are being activated. We, therefore, suggest that the outcome of CXCL12-induced skewing of CD4⁺ T cells to become Tr-1 cells is a result of two complementary pathways: a direct effect on the CD4⁺ T cells and an indirect effect that is mediated by antigen-presenting T cells to produce IL-1, which together with IL-2, polarizes Tr-1 cells (Fig. 1).

IMPLICATIONS FOR THERAPY OF AUTOIMMUNITY, GRAFT REJECTION, AND CANCER

Various tumors and in particular, androgen-dependent tumors, such as prostate cancer, breast cancer, and ovarian cancer cells,

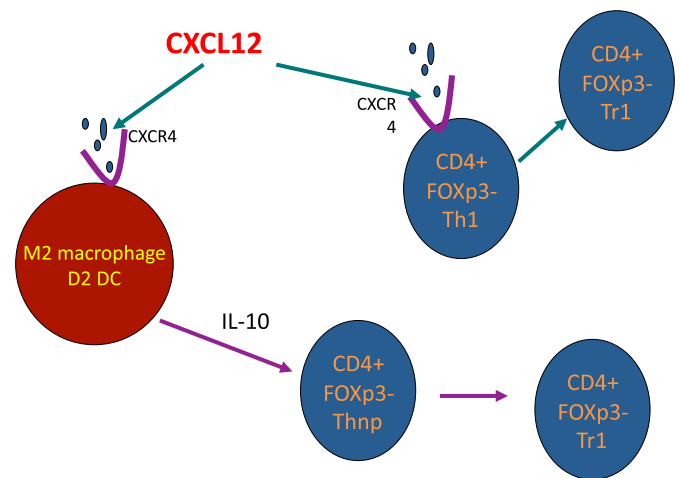


Figure 1. Direct and indirect polarization of Tr-1 by CXCL12. CXCL12 polarizes Tr-1 cells by two complementary independent mechanisms. A direct effect on CXCR4⁺ CD4⁺ T cells, including effector Th1 cells, and an indirect effect by eliciting IL-10 production in M2 macrophages and D2 dendritic cells (DC) that then polarize antigen-specific Tr-1.

produce CXCL12 predominantly and express its two receptors, CXCR4 and CXCR7 [157–161]. In these types of tumors, CXCL12 functions, in an autocrine manner, as a survival/growth factor, as well as a chemoattractant of tumor cells, and in particular, during tumor spread to the bones [53, 158, 159, 162–166]. Aside from its role as a survival/growth factor and a chemoattractant of tumor cells, CXCL12 suppresses anti-tumor immunity by promoting IL-10 production by CXCR4⁺ tumor-associated macrophages that are recruited at the tumor site. The relevance of this hypothesis has been explored in humans by Zou et al. [157], who showed that CXCL12 recruits and directs the function of plasmacytoid precursor dendritic cells so that they become IL-10-producing cells that are capable of suppressing immunity in human ovarian neoplasms. These results reinforce the idea that anti-CXCL12-based therapies could be beneficial for some types of cancers.

As mentioned previously, CXCL12 is essential for the homing of BM stem cells to the BM, and this action has potential implications for the treatment of some types of cancer. The ability of CXCL12 to attract BM-derived cells to the BM is mediated by the CXCR4 receptor [31]. Thus, a molecule that selectively blocks the CXCR4 receptor (AMD3100, Pleixafor) has been approved recently as a drug (in combination with GM-CSF) for hematopoietic stem cell mobilization from the BM to the blood, where they could be collected for autologous human stem cell transplantation [167]. This approach has been approved for intervention in two relevant cancer diseases: multiple myeloma and non-Hodgkin's lymphoma [167].

As discussed above, CXCL12 has pleiotropic actions in inflammatory autoimmune disease. It directs monocyte and T cell recruitment to restricting sites for immune surveillance, and by doing so, it stimulates effector T cells to combat potential infections at these immune-restricted sites. Nevertheless, it acts in an opposing manner during severe inflammation, where it polarizes IL-10-producing Tregs and IL-10-producing macrophages to suppress

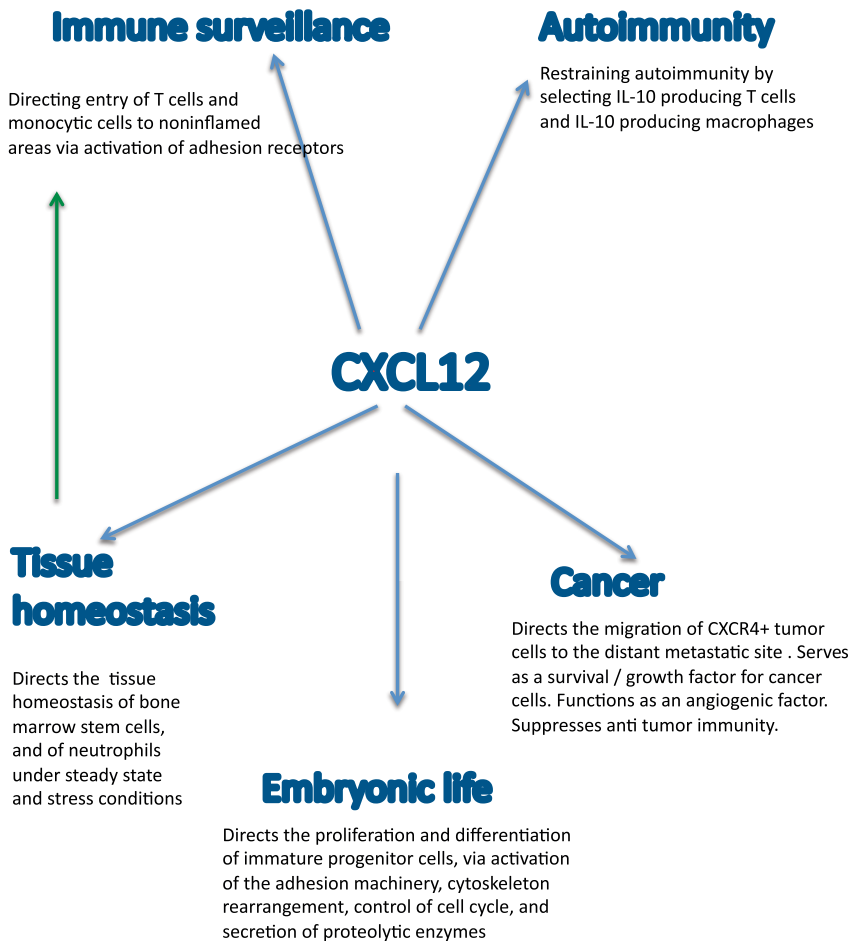


Figure 2 . Multiple faces of CXCL12.

the inflammatory process [28]. This finding suggests that CXCL12-based therapies for autoimmune diseases may be considered in the treatment of ongoing diseases but not for prophylactic use.

THE MULTIPLE FACES OF CXCL12 IN TISSUE HOMEOSTASIS, IMMUNE SURVEILLANCE, AND CANCER: WHAT SHOULD BE TAKEN IN CONSIDERATION WHEN PLANNING LONG-TERM THERAPEUTIC INTERVENTIONS?

The aims of administering a CXCL12-Ig fusion protein for treating inflammatory autoimmunity are twofold: to direct the polarization of antigen-specific T cells so that they become IL-10-producing Tr-1 cells and to polarize macrophages into IL-10-producing macrophages. The short half-life of chemokines suggests that a stabilized form of CXCL12, such as CXCL12-Ig-fusion protein, could be used in the therapy of inflammatory autoimmunity [28]. Which of the inflammatory autoimmune diseases could be a suitable candidate for CXCL12-Ig-based fusion protein therapy? Based on the underlying mechanism of disease, autoimmune diseases can be separated into those in which the direct attack of self-components is mediated by effector T cells, such as MS; inflammatory bowel diseases, such as Crohn's disease and ulcerative

colitis, T1DM, amongst others; and antibody-mediated autoimmunity, such as systemic lupus erythematosus. Tentatively, CXCL12-based therapy should be preferentially directed against effector T cell-mediated autoimmunity. Hence, it should be noted that effector T cells also direct and assist B cells to produce autoantibodies in antibody-directed autoimmunity. Thus, CXCL12-Ig-fusion protein therapy could also be beneficial for these diseases.

Other points that should not be overlooked are the other activities of this pleiotropic chemokine, which includes its role in tissue homeostasis, immune regulation of cancer, and immune surveillance. As mentioned previously, the major functions of CXCL12 in the maintenance of tissue homeostasis are directing the homing of BM stem cells and neutrophil homeostasis to ensure adequate protection against microbial pathogens [32]. Whether repeated administration of a CXCL12-Ig fusion protein would affect these functions has not yet been established. Another point of concern is whether this type of therapy would affect the Yin-Yang regulation of autoimmunity and cancer, in which the immune system protects the host from pathogens and emerging cancer cells and at the same time, minimizes local damage to tissue. Thus, shifting the balance between effector response and tolerance to favor increased tolerance to self-antigens may promote unwanted tolerance to tumor growth [168, 169]. Soluble peptide therapies [170–172] can be used to overcome this obstacle, as they can induce antigen-

specific tolerance, which could then be amplified by CXCL12-Ig fusion protein therapy. It should be noted, however, that such therapy might also result in undesirable side-effects, such as causing the production of antigen-specific IgE, which in turn, can lead to an allergic response [88].

CONCLUSIONS

This review focuses on exploring the various biological properties of CXCL12 as summarized in **Fig. 2**. In particular, its role in directing and regulating immunity. In this review, we describe the distinct functions of this chemokine in noninflammatory/low-inflammatory conditions and severe inflammation. In noninflammatory or low-inflammatory conditions, CXCL12 attracts monocytic cells and T cells to various tissues as a part of immune policing. In severe inflammation and at tumor sites, CXCL12 functions as an anti-inflammatory chemokine that skews the polarization of antigen-specific Tregs and IL-10-producing dendritic cells/monocytic cells to restrain the inflammatory process in inflammatory diseases [28] and suppress anti-tumor immunity in cancerous diseases [157]. Although these functions are beneficial for combating autoimmunity or restraining an aggressive, infectious, inflammatory process, they are undesirable in cancerous diseases, as they suppress immunity against cancer cells. It is therefore likely that therapies aiming at neutralizing the biological function of CXCL12 could be used in the therapy of various cancer diseases, whereas the administration of stabilized CXCL12 could be considered for therapy of ongoing inflammatory autoimmune diseases.

ACKNOWLEDGMENT

The author thanks Dr. Arie Bomzon [ConsulWrite (www.consulwrite.com)] for his editorial assistance in preparing this review.

REFERENCES

- Zlotnik, A., Yoshida, O. (2000) Chemokines: a new classification system and their role in immunity. *Immunity* **12**, 121–127.
- Proudfoot, A. E. (2002) Chemokine receptors: multifaceted therapeutic targets. *Nat. Rev. Immunol.* **2**, 106–115.
- Luster, A. D. (1998) Chemokines—chemotactic cytokines that mediate inflammation. *N. Engl. J. Med.* **338**, 436–445.
- Ben-Baruch, A., Michiel, D. F., Oppenheim, J. J. (1995) Signals and receptors involved in recruitment of inflammatory cells. *J. Biol. Chem.* **270**, 11703–11706.
- Loetscher, P., Seitz, M., Clark-Lewis, I., Baggiolini, M., Moser, B. (1994) Monocyte chemotactic proteins MCP-1, MCP-2, and MCP-3 are major attractants for human CD4⁺ and CD8⁺ T lymphocytes. *FASEB J.* **8**, 1055–1060.
- Huang, D. R., Wang, J., Kivisakk, P., Rollins, B. J., Ransohoff, R. M. (2001) Absence of monocyte chemoattractant protein 1 in mice leads to decreased local macrophage recruitment and antigen-specific T helper cell type 1 immune response in experimental autoimmune encephalomyelitis. *J. Exp. Med.* **193**, 713–726.
- Gong, J. H., Ratkay, L. G., Waterfield, J. D., Clark-Lewis, I. (1997) An antagonist of monocyte chemoattractant protein 1 (MCP-1) inhibits arthritis in the MRL-lpr mouse model. *J. Exp. Med.* **186**, 131–137.
- Karpus, W. J., Lukacs, N. W., McRae, B. L., Strieter, R. M., Kunkel, S. L., Miller, S. D. (1995) An important role for the chemokine macrophage inflammatory protein-1 α in the pathogenesis of the T cell-mediated autoimmune disease, experimental autoimmune encephalomyelitis. *J. Immunol.* **155**, 5003–5010.
- Kennedy, K. J., Smith, W. S., Miller, S. D., Karpus, W. J. (1997) Induction of antigen-specific tolerance for the treatment of ongoing, relapsing autoimmune encephalomyelitis: a comparison between oral and peripheral tolerance. *J. Immunol.* **159**, 1036–1044.
- Rollins, B. J. (1996) Monocyte chemoattractant protein 1: a potential regulator of monocyte recruitment in inflammatory disease. *Mol. Med. Today* **2**, 198–204.
- Izhak, L., Wildbaum, G., Zohar, Y., Anunu, R., Klapper, L., Elkeles, A., Seagal, J., Yefenof, E., Ayalon-Soffer, M., Karin, N. (2009) A novel recombinant fusion protein encoding a 20-amino acid residue of the third extracellular (E3) domain of CCR2 neutralizes the biological activity of CCL2. *J. Immunol.* **183**, 732–739.
- Liu, M. T., Keirstead, H. S., Lane, T. E. (2001) Neutralization of the chemokine CXCL10 reduces inflammatory cell invasion and demyelination and improves neurological function in a viral model of multiple sclerosis. *J. Immunol.* **167**, 4091–4097.
- Liu, M. T., Armstrong, D., Hamilton, T. A., Lane, T. E. (2001) Expression of Mig (monokine induced by interferon- γ) is important in T lymphocyte recruitment and host defense following viral infection of the central nervous system. *J. Immunol.* **166**, 1790–1795.
- Ransohoff, R. M., Kivisakk, P., Kidd, G. (2003) Three or more routes for leukocyte migration into the central nervous system. *Nat. Rev. Immunol.* **3**, 569–581.
- Rottman, J. B., Slavin, A. J., Silva, R., Weiner, H. L., Gerard, C. G., Hancock, W. W. (2000) Leukocyte recruitment during onset of experimental allergic encephalomyelitis is CCR1 dependent. *Eur. J. Immunol.* **30**, 2372–2377.
- Sorensen, T. L., Tani, M., Jensen, J., Pierce, V., Lucchinetti, C., Folcik, V. A., Qin, S., Rottman, J., Sellebjerg, F., Strieter, R. M., Frederiksen, J. L., Ransohoff, R. M. (1999) Expression of specific chemokines and chemokine receptors in the central nervous system of multiple sclerosis patients. *J. Clin. Invest.* **103**, 807–815.
- Balashov, K. E., Rottman, J. B., Weiner, H. L., Hancock, W. W. (1999) CCR5(+) and CXCR3(+) T cells are increased in multiple sclerosis and their ligands MIP-1 α and IP-10 are expressed in demyelinating brain lesions. *Proc. Natl. Acad. Sci. USA* **96**, 6873–6878.
- Huang, D., Han, Y., Rani, M. R., Glabinski, A., Trebst, C., Sorensen, T., Tani, M., Wang, J., Chien, P., O'Bryan, S., Bielecki, B., Zhou, Z. L., Majumder, S., Ransohoff, R. M. (2000) Chemokines and chemokine receptors in inflammation of the nervous system: manifold roles and exquisite regulation. *Immunol. Rev.* **177**, 52–67.
- Godiska, R., Chantry, D., Dietsch, G. N., Gray, P. W. (1995) Chemokine expression in murine experimental allergic encephalomyelitis. *J. Neuroimmunol.* **58**, 167–176.
- Izikson, L., Klein, R. S., Charo, I. F., Weiner, H. L., Luster, A. D. (2000) Resistance to experimental autoimmune encephalomyelitis in mice lacking the CC chemokine receptor (CCR)2. *J. Exp. Med.* **192**, 1075–1080.
- Fife, B. T., Kennedy, K. J., Paniagua, M. C., Lukacs, N. W., Kunkel, S. L., Luster, A. D., Karpus, W. J. (2001) CXCL10 (IFN- γ -inducible protein-10) control of encephalitogenic CD4⁺ T cell accumulation in the central nervous system during experimental autoimmune encephalomyelitis. *J. Immunol.* **166**, 7617–7624.
- Fife, B. T., Paniagua, M. C., Lukacs, N. W., Kunkel, S. L., Karpus, W. J. (2001) Selective CC chemokine receptor expression by central nervous system-infiltrating encephalitogenic T cells during experimental autoimmune encephalomyelitis. *J. Neurosci. Res.* **66**, 705–714.
- Shehadeh, N., Pollack, S., Wildbaum, G., Zohar, Y., Shafat, I., Makhoul, R., Daod, E., Hakim, F., Perlman, R., Karin, N. (2009) Selective autoantibody production against CCL3 is associated with human type 1 diabetes mellitus and serves as a novel biomarker for its diagnosis. *J. Immunol.* **182**, 8104–8109.
- Cameron, M. J., Arreaza, G. A., Grattan, M., Meagher, C., Sharif, S., Burdick, M. D., Strieter, R. M., Cook, D. N., Delovitch, T. L. (2000) Differential expression of CC chemokines and the CCR5 receptor in the pancreas is associated with progression to type 1 diabetes. *J. Immunol.* **165**, 1102–1110.
- Youssef, S., Maor, G., Wildbaum, G., Grabie, N., Gour-Lavie, A., Karin, N. (2000) C-C chemokine-encoding DNA vaccines enhance breakdown of tolerance to their gene products and treat ongoing adjuvant arthritis. *J. Clin. Invest.* **106**, 361–371.
- Salomon, I., Netzer, N., Wildbaum, G., Schiff-Zuck, S., Maor, G., Karin, N. (2002) Targeting the function of IFN- γ -inducible protein 10 suppresses ongoing adjuvant arthritis. *J. Immunol.* **169**, 2685–2693.
- Pierer, M., Rethage, J., Seibl, R., Lauener, R., Brentano, F., Wagner, U., Hantzel, H., Michel, B. A., Gay, R. E., Gay, S., Kyburz, D. (2004) Chemokine secretion of rheumatoid arthritis synovial fibroblasts stimulated by Toll-like receptor 2 ligands. *J. Immunol.* **172**, 1256–1265.
- Meiron, M., Zohar, Y., Anunu, R., Wildbaum, G., Karin, N. (2008) CXCL12 (SDF-1 α) suppresses ongoing experimental autoimmune encephalomyelitis by selecting antigen-specific regulatory T cells. *J. Exp. Med.* **205**, 2643–2655.
- Dar, A., Kollet, O., Lapidot, T. (2006) Mutual, reciprocal SDF-1/CXCR4 interactions between hematopoietic and bone marrow stromal cells regulate human stem cell migration and development in NOD/SCID chimeric mice. *Exp. Hematol.* **34**, 967–975.
- Serbina, N. V., Pamer, E. G. (2006) Monocyte emigration from bone marrow during bacterial infection requires signals mediated by chemokine receptor CCR2. *Nat. Immunol.* **7**, 311–317.

31. Peled, A., Petit, I., Kollet, O., Magid, M., Ponomarev, T., Byk, T., Nagler, A., Ben-Hur, H., Many, A., Shultz, L., Lider, O., Alon, R., Zipori, D., Lapidot, T. (1999) Dependence of human stem cell engraftment and repopulation of NOD/SCID mice on CXCR4. *Science* **283**, 845–848.
32. Christopher, M. J., Link, D. C. (2007) Regulation of neutrophil homeostasis. *Curr. Opin. Hematol.* **14**, 3–8.
33. Eash, K. J., Means, J. M., White, D. W., Link, D. C. (2009) CXCR4 is a key regulator of neutrophil release from the bone marrow under basal and stress granulopoiesis conditions. *Blood* **113**, 4711–4719.
34. Karin, N., Szafer, F., Mitchell, D., Gold, D. P., Steinman, L. (1993) Selective and nonselective stages in homing of T lymphocytes to the central nervous system during experimental allergic encephalomyelitis. *J. Immunol.* **150**, 4116–4124.
35. Blades, M. C., Ingegnoli, F., Wheller, S. K., Manzo, A., Wahid, S., Panayi, G. S., Perretti, M., Pitzalis, C. (2002) Stromal cell-derived factor 1 (CXCL12) induces monocyte migration into human synovium transplanted onto SCID mice. *Arthritis Rheum.* **46**, 824–836.
36. Rossi, D., Zlotnik, A. (2000) The biology of chemokines and their receptors. *Annu. Rev. Immunol.* **18**, 217–242.
37. Lloyd, A. R., Oppenheim, J. J., Kelvin, D. J., Taub, D. D. (1996) Chemokines regulate T cell adherence to recombinant adhesion molecules and extracellular matrix proteins. *J. Immunol.* **156**, 932–938.
38. Del Pozo, M. A., Sanchez-Mateos, P., Nieto, M., Sanchez-Madrid, F. (1995) Chemokines regulate cellular polarization and adhesion receptor redistribution during lymphocyte interaction with endothelium and extracellular matrix. Involvement of cAMP signaling pathway. *J. Cell Biol.* **131**, 495–508.
39. Vaddi, K., Newton, R. C. (1994) Regulation of monocyte integrin expression by β -family chemokines. *J. Immunol.* **153**, 4721–4732.
40. Carlos, T. M., Harlan, J. M. (1994) Leukocyte-endothelial adhesion molecules. *Blood* **84**, 2068–2101.
41. Wildbaum, G., Netzer, N., Karin, N. (2002) Plasmid DNA encoding IFN- γ -inducible protein 10 redirects antigen-specific T cell polarization and suppresses experimental autoimmune encephalomyelitis. *J. Immunol.* **168**, 5885–5892.
42. Yednock, T. A., Cannon, C., Fritz, L. C., Sanchez-Madrid, F., Steinman, L., Karin, N. (1992) Prevention of experimental autoimmune encephalomyelitis by antibodies against $\alpha 4 \beta 1$ integrin. *Nature* **356**, 63–66.
43. Von Andrian, U. H., Engelhardt, B. (2003) $\alpha 4$ Integrins as therapeutic targets in autoimmune disease. *N. Engl. J. Med.* **348**, 68–72.
44. Miller, D. H., Khan, O. A., Sheremata, W. A., Blumhardt, L. D., Rice, G. P., Libonati, M. A., Willmer-Hulme, A. J., Dalton, C. M., Miskiel, K. A., O'Connor, P. W. (2003) A controlled trial of natalizumab for relapsing multiple sclerosis. *N. Engl. J. Med.* **348**, 15–23.
45. Balkwill, F. (2004) Cancer and the chemokine network. *Nat. Rev. Cancer* **4**, 540–550.
46. Homey, B., Muller, A., Zlotnik, A. (2002) Chemokines: agents for the immunotherapy of cancer? *Nat. Rev. Immunol.* **2**, 175–184.
47. Loberg, R. D., Day, L. L., Harwood, J., Ying, C., St John, L. N., Giles, R., Neeley, C. K., Pienta, K. J. (2006) CCL2 is a potent regulator of prostate cancer cell migration and proliferation. *Neoplasia* **8**, 578–586.
48. Lewis, C. E., Pollard, J. W. (2006) Distinct role of macrophages in different tumor microenvironments. *Cancer Res.* **66**, 605–612.
49. Lin, E. Y., Li, J. F., Gnatovskiy, L., Deng, Y., Zhu, L., Grzesik, D. A., Qian, H., Xue, X. N., Pollard, J. W. (2006) Macrophages regulate the angiogenic switch in a mouse model of breast cancer. *Cancer Res.* **66**, 11238–11246.
50. Porta, C., Subhra Kumar, B., Larghi, P., Rubino, L., Mancino, A., Sica, A. (2007) Tumor promotion by tumor-associated macrophages. *Adv. Exp. Med. Biol.* **604**, 67–86.
51. Sica, A., Allavena, P., Mantovani, A. (2008) Cancer related inflammation: the macrophage connection. *Cancer Lett.* **267**, 204–215.
52. Vander Cappellen, J., Van Damme, J., Struyf, S. (2008) The role of CXC chemokines and their receptors in cancer. *Cancer Lett.* **267**, 226–244.
53. Wang, J., Shiozawa, Y., Wang, Y., Jung, Y., Pienta, K. J., Mehra, R., Loberg, R., Taichman, R. S. (2008) The role of CXCR7/RDC1 as a chemokine receptor for CXCL12/SDF-1 in prostate cancer. *J. Biol. Chem.* **283**, 4283–4294.
54. Miao, Z., Luker, K. E., Summers, B. C., Berahovich, R., Bhojani, M. S., Rehemtulla, A., Kleer, C. G., Essner, J. J., Nasevicius, A., Luker, G. D., Howard, M. C., Schall, T. J. (2007) CXCR7 (RDC1) promotes breast and lung tumor growth in vivo and is expressed on tumor-associated vasculature. *Proc. Natl. Acad. Sci. USA* **104**, 15735–15740.
55. Sun, Y. X., Schneider, A., Jung, Y., Wang, J., Dai, J., Wang, J., Cook, K., Osman, N. I., Koh-Paige, A. J., Shim, H., Pienta, K. J., Keller, E. T., McCauley, L. K., Taichman, R. S. (2005) Skeletal localization and neutralization of the SDF-1(CXCL12)/CXCR4 axis blocks prostate cancer metastasis and growth in osseous sites in vivo. *J. Bone Miner. Res.* **20**, 318–329.
56. Singh, S., Singh, U. P., Grizzle, W. E., Lillard Jr., J. W. (2004) CXCL12-CXCR4 interactions modulate prostate cancer cell migration, metalloproteinase expression and invasion. *Lab. Invest.* **84**, 1666–1676.
57. Ono, M. (2008) Molecular links between tumor angiogenesis and inflammation: inflammatory stimuli of macrophages and cancer cells as targets for therapeutic strategy. *Cancer Sci.* **99**, 1501–1506.
58. Fujita, K., Ewing, C. M., Sokoll, L. J., Elliott, D. J., Cunningham, M., De Marzo, A. M., Isaacs, W. B., Pavlovich, C. P. (2008) Cytokine profiling of prostatic fluid from cancerous prostate glands identifies cytokines associated with extent of tumor and inflammation. *Prostate* **68**, 872–882.
59. Loberg, R. D., Ying, C., Craig, M., Yan, L., Snyder, L. A., Pienta, K. J. (2007) CCL2 as an important mediator of prostate cancer growth in vivo through the regulation of macrophage infiltration. *Neoplasia* **9**, 556–562.
60. Condeelis, J., Pollard, J. W. (2006) Macrophages: obligate partners for tumor cell migration, invasion, and metastasis. *Cell* **124**, 263–266.
61. Mantovani, A., Sozzani, S., Locati, M., Allavena, P., Sica, A. (2002) Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol.* **23**, 549–555.
62. Mantovani, A., Bottazzi, B., Colotta, F., Sozzani, S., Ruco, L. (1992) The origin and function of tumor-associated macrophages. *Immunol. Today* **13**, 265–270.
63. Fukumoto, N., Shimaoka, T., Fujimura, H., Sakoda, S., Tanaka, M., Kita, T., Yonehara, S. (2004) Critical roles of CXC chemokine ligand 16/scavenger receptor that binds phosphatidylserine and oxidized lipoprotein in the pathogenesis of both acute and adoptive transfer experimental autoimmune encephalomyelitis. *J. Immunol.* **173**, 1620–1627.
64. Youssef, S., Wildbaum, G., Karin, N. (1999) Prevention of experimental autoimmune encephalomyelitis by MIP-1 α and MCP-1 naked DNA vaccines. *J. Autoimmun.* **13**, 21–29.
65. Youssef, S., Wildbaum, G., Maor, G., Lanir, N., Gour-Lavie, A., Grabie, N., Karin, N. (1998) Long-lasting protective immunity to experimental autoimmune encephalomyelitis following vaccination with naked DNA encoding C-C chemokines. *J. Immunol.* **161**, 3870–3879.
66. Karpus, W. J., Kennedy, K. J., Kunkel, S. L., Lukacs, N. W. (1998) Monocyte chemoattractant protein 1 regulates oral tolerance induction by inhibition of T helper cell 1-related cytokines. *J. Exp. Med.* **187**, 733–741.
67. Karpus, W. J., Kennedy, K. J. (1997) MIP-1 α and MCP-1 differentially regulate acute and relapsing autoimmune encephalomyelitis as well as Th1/Th2 lymphocyte differentiation. *J. Leukoc. Biol.* **62**, 681–687.
68. Molon, B., Gri, G., Bettella, M., Gomez-Mouton, C., Lanzavecchia, A., Martinez, A. C., Manes, S., Viola, A. (2005) T cell costimulation by chemokine receptors. *Nat. Immunol.* **6**, 465–471.
69. Contento, R. L., Boularan, C., Pozzan, T., Manes, S., Marullo, S., Viola, A. (2008) CXCR4-CCR5: a couple modulating T cell functions. *Proc. Natl. Acad. Sci. USA* **105**, 10101–10106.
70. Camargo, J. F., Quinones, M. P., Mummidi, S., Srinivas, S., Gaitan, A. A., Begum, K., Jimenez, F., VanCompernelle, S., Unutmaz, D., Ahuja, S. S., Ahuja, S. K. (2009) CCR5 expression levels influence NFAT translocation, IL-2 production, and subsequent signaling events during T lymphocyte activation. *J. Immunol.* **182**, 171–182.
71. Matsui, M., Weaver, J., Proudfoot, A. E., Wujek, J. R., Wei, T., Richer, E., Trapp, B. D., Rao, A., Ransohoff, R. M. (2002) Treatment of experimental autoimmune encephalomyelitis with the chemokine receptor antagonist Met-RANTES. *J. Neuroimmunol.* **128**, 16–22.
72. Jia, Y., Li, H., Chen, W., Li, M., Lv, M., Feng, P., Hu, H., Zhang, L. (2006) Prevention of murine experimental autoimmune encephalomyelitis by in vivo expression of a novel recombinant immunotoxin DT390-RANTES. *Gene Ther.* **13**, 1351–1359.
73. Barnes, D. A., Tse, J., Kaufhold, M., Owen, M., Hesselgesser, J., Strieter, R., Horuk, R., Perez, H. D. (1998) Polyclonal antibody directed against human RANTES ameliorates disease in the Lewis rat adjuvant-induced arthritis model. *J. Clin. Invest.* **101**, 2910–2919.
74. Mosmann, T. R., Cherwinski, H., Bond, M. W., Giedlin, M. A., Coffman, R. L. (1986) Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. *J. Immunol.* **136**, 2348–2357.
75. Mosmann, T. R., Coffman, R. L. (1989) TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. *Annu. Rev. Immunol.* **7**, 145–173.
76. O'Garra, A., Steinman, L., Gijbels, K. (1997) CD4+ T-cell subsets in autoimmunity. *Curr. Opin. Immunol.* **9**, 872–883.
77. Abbas, A. K., Murphy, K. M., Sher, A. (1996) Functional diversity of helper T lymphocytes. *Nature* **383**, 787–793.
78. Seder, R. A., Gazzinelli, R., Sher, A., Paul, W. E. (1993) IL-12 acts directly on CD4+ T cells to enhance priming for IFN- γ production and diminishes IL-4 inhibition of such priming. *Proc. Natl. Acad. Sci. USA* **90**, 10188–10192.
79. Trinchieri, G. (1993) Interleukin-12 and its role in the generation of Th1 cells. *Immunol. Today* **14**, 335–337.
80. O'Garra, A., Murphy, K. (1994) Role of cytokines in determining T-lymphocyte function. *Curr. Opin. Immunol.* **6**, 458–466.
81. Wildbaum, G., Youssef, S., Grabie, N., Karin, N. (1998) Neutralizing antibodies to IFN- γ -inducing factor prevent experimental autoimmune encephalomyelitis. *J. Immunol.* **161**, 6368–6374.
82. Castro, A. G., Hauser, T. M., Cocks, B. G., Abrams, J., Zurawski, S., Churakova, T., Zonin, F., Robinson, D., Tangye, S. G., Aversa, G., Nichols, K. E., de Vries, J. E., Lanier, L. L., O'Garra, A. (1999) Molecular and functional characterization of mouse signaling lymphocytic activation molecule (SLAM): differential expression and responsiveness in Th1 and Th2 cells. *J. Immunol.* **163**, 5860–5870.
83. Stevens, T. L., Bossie, A., Sanders, V. M., Fernandez-Botran, R., Coffman, R. L., Mosmann, T. R., Vitetta, E. S. (1988) Regulation of antibody

- isotype secretion by subsets of antigen-specific helper T cells. *Nature* **334**, 255–258.
84. Leonard, J. P., Waldburger, K. E., Goldman, S. J. (1995) Prevention of experimental autoimmune encephalomyelitis by antibodies against interleukin 12. *J. Exp. Med.* **181**, 381–386.
 85. Butler, D. M., Malfait, A. M., Maini, R. N., Brennan, F. M., Feldmann, M. (1999) Anti-IL-12 and anti-TNF antibodies synergistically suppress the progression of murine collagen-induced arthritis. *Eur. J. Immunol.* **29**, 2205–2212.
 86. Gracie, J. A., Forsey, R. J., Chan, W. L., Gilmour, A., Leung, B. P., Greer, M. R., Kennedy, K., Carter, R., Wei, X. Q., Xu, D., Field, M., Foulis, A., Liew, F. Y., McInnes, I. B. (1999) A proinflammatory role for IL-18 in rheumatoid arthritis. *J. Clin. Invest.* **104**, 1393–1401.
 87. Joosten, L. A., van De Loo, F. A., Lubberts, E., Helsen, M. M., Netea, M. G., van Der Meer, J. W., Dinarello, C. A., van Den Berg, W. B. (2000) An IFN- γ -independent proinflammatory role of IL-18 in murine streptococcal cell wall arthritis. *J. Immunol.* **165**, 6553–6558.
 88. Pedotti, R., Mitchell, D., Wedemeyer, J., Karpuz, M., Chabas, D., Hattab, E. M., Tsai, M., Galli, S. J., Steinman, L. (2001) An unexpected version of horror autotoxicus: anaphylactic shock to a self-peptide. *Nat. Immunol.* **2**, 216–222.
 89. Aggarwal, S., Ghilardi, N., Xie, M. H., de Sauvage, F. J., Gurney, A. L. (2003) Interleukin-23 promotes a distinct CD4 T cell activation state characterized by the production of interleukin-17. *J. Biol. Chem.* **278**, 1910–1914.
 90. Bettelli, E., Kuchroo, V. K. (2005) IL-12- and IL-23-induced T helper cell subsets: birds of the same feather flock together. *J. Exp. Med.* **201**, 169–171.
 91. Hunter, C. A. (2005) New IL-12-family members: IL-23 and IL-27, cytokines with divergent functions. *Nat. Rev. Immunol.* **5**, 521–531.
 92. Bettelli, E., Carrier, Y., Gao, W., Korn, T., Strom, T. B., Oukka, M., Weiner, H. L., Kuchroo, V. K. (2006) Reciprocal developmental pathways for the generation of pathogenic effector TH17 and regulatory T cells. *Nature* **441**, 235–238.
 93. Bettelli, E., Oukka, M., Kuchroo, V. K. (2007) T(H)-17 cells in the circle of immunity and autoimmunity. *Nat. Immunol.* **8**, 345–350.
 94. Cua, D. J., Sherlock, J., Chen, Y., Murphy, C. A., Joyce, B., Seymour, B., Lucian, L., To, W., Kwan, S., Churakova, T., Zurawski, S., Wickowski, M., Lira, S. A., Gorman, D., Kastelein, R. A., Sedgwick, J. D. (2003) Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. *Nature* **421**, 744–748.
 95. Langrish, C. L., Chen, Y., Blumenschein, W. M., Mattson, J., Basham, B., Sedgwick, J. D., McClanahan, T., Kastelein, R. A., Cua, D. J. (2005) IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J. Exp. Med.* **201**, 233–240.
 96. Lubberts, E., Koenders, M. I., van den Berg, W. B. (2005) The role of T cell interleukin-17 in conducting destructive arthritis: lessons from animal models. *Arthritis Res. Ther.* **7**, 29–37.
 97. McGeachy, M. J., Anderson, S. M. (2005) Cytokines in the induction and resolution of experimental autoimmune encephalomyelitis. *Cytokine* **32**, 81–84.
 98. Thakker, P., Leach, M. W., Kuang, W., Benoit, S. E., Leonard, J. P., Marusic, S. (2007) IL-23 is critical in the induction but not in the effector phase of experimental autoimmune encephalomyelitis. *J. Immunol.* **178**, 2589–2598.
 99. Kroenke, M. A., Carlson, T. J., Andjelkovic, A. V., Segal, B. M. (2008) IL-12- and IL-23-modulated T cells induce distinct types of EAE based on histology, CNS chemokine profile, and response to cytokine inhibition. *J. Exp. Med.* **205**, 1535–1541.
 100. Luger, D., Silver, P. B., Tang, J., Cua, D., Chen, Z., Iwakura, Y., Bowman, E. P., Sgambellone, N. M., Chan, C. C., Caspi, R. R. (2008) Either a Th17 or a Th1 effector response can drive autoimmunity: conditions of disease induction affect dominant effector category. *J. Exp. Med.* **205**, 799–810.
 101. Anderson, C. F., Oukka, M., Kuchroo, V. J., Sacks, D. (2007) CD4(+)CD25(–)Foxp3(–) Th1 cells are the source of IL-10-mediated immune suppression in chronic cutaneous leishmaniasis. *J. Exp. Med.* **204**, 285–297.
 102. Josefowicz, S. Z., Rudensky, A. (2009) Control of regulatory T cell lineage commitment and maintenance. *Immunity* **30**, 616–625.
 103. O'Garra, A., Vieira, P. (2007) T(H)1 cells control themselves by producing interleukin-10. *Nat. Rev. Immunol.* **7**, 425–428.
 104. Jager, A., Dardalhon, V., Sobel, R. A., Bettelli, E., Kuchroo, V. K. (2009) Th1, Th17, and Th9 effector cells induce experimental autoimmune encephalomyelitis with different pathological phenotypes. *J. Immunol.* **183**, 7169–7177.
 105. Fujita, H., Nogales, K. E., Kikuchi, T., Gonzalez, J., Carucci, J. A., Krueger, J. G. (2009) Human Langerhans cells induce distinct IL-22-producing CD4+ T cells lacking IL-17 production. *Proc. Natl. Acad. Sci. USA* **106**, 21795–21800.
 106. Sakaguchi, S. (2005) Naturally arising Foxp3-expressing CD25+CD4+ regulatory T cells in immunological tolerance to self and non-self. *Nat. Immunol.* **6**, 345–352.
 107. Shevach, E. M. (2009) Mechanisms of Foxp3+ T regulatory cell-mediated suppression. *Immunity* **30**, 636–645.
 108. Chen, Y., Kuchroo, V. K., Inobe, J., Hafler, D., Weiner, H. L. (1994) Regulatory T-cell clones induced by oral tolerance: suppression of autoimmune encephalomyelitis. *Science* **265**, 1237–1240.
 109. Khoury, S. J., Hancock, W. W., Weiner, H. L. (1992) Oral tolerance to myelin basic protein and natural recovery from experimental autoimmune encephalomyelitis are associated with downregulation of inflammatory cytokines and differential upregulation of transforming growth factor β , interleukin 4, and prostaglandin E expression in the brain. *J. Exp. Med.* **176**, 1355–1364.
 110. Groux, H., O'Garra, A., Bigler, M., Rouleau, M., Antonenko, S., de Vries, J. E., Roncarolo, M. G. (1997) A CD4+ T-cell subset inhibits antigen-specific T-cell responses and prevents colitis. *Nature* **389**, 737–742.
 111. Collison, L. W., Workman, C. J., Kuo, T. T., Boyd, K., Wang, Y., Vignali, K. M., Cross, R., Schy, D., Blumberg, R. S., Vignali, D. A. (2007) The inhibitory cytokine IL-35 contributes to regulatory T-cell function. *Nature* **450**, 566–569.
 112. Bennett, C. L., Christie, J., Ramsdell, F., Brunkow, M. E., Ferguson, P. J., Whitesell, L., Kelly, T. E., Saulsbury, F. T., Chance, P. F., Ochs, H. D. (2001) The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat. Genet.* **27**, 20–21.
 113. Lahl, K., Loddenkemper, C., Drouin, C., Freyer, J., Arnason, J., Eberl, G., Hamann, A., Wagner, H., Huehn, J., Sparwasser, T. (2007) Selective depletion of Foxp3+ regulatory T cells induces a scurfy-like disease. *J. Exp. Med.* **204**, 57–63.
 114. Wildbaum, G., Netzer, N., Karin, N. (2002) Tr1 cell-dependent active tolerance blunts the pathogenic effects of determinant spreading. *J. Clin. Invest.* **110**, 701–710.
 115. Avni, O., Lee, D., Macian, F., Szabo, S. J., Glimcher, L. H., Rao, A. (2002) T(H) cell differentiation is accompanied by dynamic changes in histone acetylation of cytokine genes. *Nat. Immunol.* **3**, 643–651.
 116. Szabo, S. J., Kim, S. T., Costa, G. L., Zhang, X., Fathman, C. G., Glimcher, L. H. (2000) A novel transcription factor, T-bet, directs Th1 lineage commitment. *Cell* **100**, 655–669.
 117. Sallusto, F., Lenig, D., Forster, R., Lipp, M., Lanzavecchia, A. (1999) Two subsets of memory T lymphocytes with distinct homing potentials and effector functions. *Nature* **401**, 708–712.
 118. Zhu, J., Cote-Sierra, J., Guo, L., Paul, W. E. (2003) Stat5 activation plays a critical role in Th2 differentiation. *Immunity* **19**, 739–748.
 119. Zheng, W., Flavell, R. A. (1997) The transcription factor GATA-3 is necessary and sufficient for Th2 cytokine gene expression in CD4 T cells. *Cell* **89**, 587–596.
 120. Ivanov, I. I., Zhou, L., Littman, D. R. (2007) Transcriptional regulation of Th17 cell differentiation. *Semin. Immunol.* **19**, 409–417.
 121. Vieira, P. L., Christensen, J. R., Minae, S., O'Neill, E. J., Barrat, F. J., Boonstra, A., Barthlott, T., Stockinger, B., Wraith, D. C., O'Garra, A. (2004) IL-10-secreting regulatory T cells do not express Foxp3 but have comparable regulatory function to naturally occurring CD4+CD25+ regulatory T cells. *J. Immunol.* **172**, 5986–5993.
 122. Hori, S., Nomura, T., Sakaguchi, S. (2003) Control of regulatory T cell development by the transcription factor Foxp3. *Science* **299**, 1057–1061.
 123. Fontenot, J. D., Gavin, M. A., Rudensky, A. Y. (2003) Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat. Immunol.* **4**, 330–336.
 124. Marie, J. C., Letterio, J. J., Gavin, M., Rudensky, A. Y. (2005) TGF- β 1 maintains suppressor function and Foxp3 expression in CD4+CD25+ regulatory T cells. *J. Exp. Med.* **201**, 1061–1067.
 125. Zhou, L., Lopes, J. E., Chong, M. M., Ivanov, I. I., Min, R., Victora, G. D., Shen, Y., Du, J., Rubtsov, Y. P., Rudensky, A. Y., Ziegler, S. F., Littman, D. R. (2008) TGF- β -induced Foxp3 inhibits T(H)17 cell differentiation by antagonizing ROR γ t function. *Nature* **453**, 236–240.
 126. Tsuji-Takayama, K., Suzuki, M., Yamamoto, M., Harashima, A., Okochi, A., Otani, T., Inoue, T., Sugimoto, A., Toraya, T., Takeuchi, M., Yamasaki, F., Nakamura, S., Kibata, M. (2008) The production of IL-10 by human regulatory T cells is enhanced by IL-2 through a STAT5-responsive intronic enhancer in the IL-10 locus. *J. Immunol.* **181**, 3897–3905.
 127. Goldberg, R., Zohar, Y., Wildbaum, G., Geron, Y., Maor, G., Karin, N. (2004) Suppression of ongoing experimental autoimmune encephalomyelitis by neutralizing the function of the p28 subunit of IL-27. *J. Immunol.* **173**, 6465–6471.
 128. Pflanz, S., Timans, J. C., Cheung, J., Rosales, R., Kanzler, H., Gilbert, J., Hibbert, L., Churakova, T., Travis, M., Vaisberg, E., Blumenschein, W. M., Mattson, J. D., Wagner, J. L., To, W., Zurawski, S., McClanahan, T. K., Gorman, D. M., Bazan, J. F., de Waal Malefyt, R., Rennick, D., Kastelein, R. A. (2002) IL-27, a heterodimeric cytokine composed of EBI3 and p28 protein, induces proliferation of naive CD4(+) T cells. *Immunity* **16**, 779–790.
 129. Robinson, D. S., O'Garra, A. (2002) Further checkpoints in Th1 development. *Immunity* **16**, 755–758.
 130. Stumhofer, J. S., Silver, J. S., Laurence, A., Porrett, P. M., Harris, T. H., Turka, L. A., Ernst, M., Saris, C. J., O'Shea, J. J., Hunter, C. A. (2007) Interleukins 27 and 6 induce STAT3-mediated T cell production of interleukin 10. *Nat. Immunol.* **8**, 1363–1371.
 131. Awasthi, A., Carrier, Y., Peron, J. P., Bettelli, E., Kamanaka, M., Flavell, R. A., Kuchroo, V. K., Oukka, M., Weiner, H. L. (2007) A dominant

- function for interleukin 27 in generating interleukin 10-producing anti-inflammatory T cells. *Nat. Immunol.* **8**, 1380–1389.
132. Fitzgerald, D. C., Zhang, G. X., ElBehi, M., Fonseca-Kelly, Z., Li, H., Yu, S., Saris, C. J., Gran, B., Ciric, B., Rostami, A. (2007) Suppression of autoimmune inflammation of the central nervous system by interleukin 10 secreted by interleukin 27-stimulated T cells. *Nat. Immunol.* **8**, 1372–1379.
 133. Sallusto, F., Mackay, C. R., Lanzavecchia, A. (1997) Selective expression of the eotaxin receptor CCR3 by human T helper 2 cells. *Science* **277**, 2005–2007.
 134. Sallusto, F., Lenig, D., Mackay, C. R., Lanzavecchia, A. (1998) Flexible programs of chemokine receptor expression on human polarized T helper 1 and 2 lymphocytes. *J. Exp. Med.* **187**, 875–883.
 135. Sallusto, F., Lanzavecchia, A., Mackay, C. R. (1998) Chemokines and chemokine receptors in T-cell priming and Th1/Th2-mediated responses. *Immunol. Today* **19**, 568–574.
 136. Hirahara, K., Liu, L., Clark, R. A., Yamanaka, K., Fuhlbrigge, R. C., Kupper, T. S. (2006) The majority of human peripheral blood CD4⁺CD25^{high}Foxp3⁺ regulatory T cells bear functional skin-homing receptors. *J. Immunol.* **177**, 4488–4494.
 137. Reboldi, A., Coisne, C., Baumjohann, D., Benvenuto, F., Bottinelli, D., Lira, S., Uccelli, A., Lanzavecchia, A., Engelhardt, B., Sallusto, F. (2009) C-C chemokine receptor 6-regulated entry of TH-17 cells into the CNS through the choroid plexus is required for the initiation of EAE. *Nat. Immunol.* **10**, 514–523.
 138. Soler, D., Chapman, T. R., Poisson, L. R., Wang, L., Cote-Sierra, J., Ryan, M., McDonald, A., Badola, S., Fedyk, E., Coyle, A. J., Hodge, M. R., Kolbeck, R. (2006) CCR8 expression identifies CD4 memory T cells enriched for FOXP3⁺ regulatory and Th2 effector lymphocytes. *J. Immunol.* **177**, 6940–6951.
 139. Gangur, V., Simons, F. E., Hayglass, K. T. (1998) Human IP-10 selectively promotes dominance of polyclonally activated and environmental antigen-driven IFN- γ over IL-4 responses. *FASEB J.* **12**, 705–713.
 140. Flanagan, K., Moroziewicz, D., Kwak, H., Horig, H., Kaufman, H. L. (2004) The lymphoid chemokine CCL21 costimulates naive T cell expansion and Th1 polarization of non-regulatory CD4⁺ T cells. *Cell. Immunol.* **231**, 75–84.
 141. D'Apuzzo, M., Rolink, A., Loetscher, M., Hoxie, J. A., Clark-Lewis, I., Melchers, F., Baggiolini, M., Moser, B. (1997) The chemokine SDF-1, stromal cell-derived factor 1, attracts early stage B cell precursors via the chemokine receptor CXCR4. *Eur. J. Immunol.* **27**, 1788–1793.
 142. Nagasawa, T., Kikutani, H., Kishimoto, T. (1994) Molecular cloning and structure of a pre-B-cell growth-stimulating factor. *Proc. Natl. Acad. Sci. USA* **91**, 2305–2309.
 143. Nanki, T., Lipsky, P. E. (2000) Cutting edge: stromal cell-derived factor-1 is a costimulator for CD4⁺ T cell activation. *J. Immunol.* **164**, 5010–5014.
 144. Campbell, J. J., Hedrick, J., Zlotnik, A., Siani, M. A., Thompson, D. A., Butcher, E. C. (1998) Chemokines and the arrest of lymphocytes rolling under flow conditions. *Science* **279**, 381–384.
 145. Wright, N., Hidalgo, A., Rodriguez-Frade, J. M., Soriano, S. F., Mellado, M., Parmo-Cabanas, M., Briskin, M. J., Teixeira, J. (2002) The chemokine stromal cell-derived factor-1 α modulates α 4 β 7 integrin-mediated lymphocyte adhesion to mucosal addressin cell adhesion molecule-1 and fibronectin. *J. Immunol.* **168**, 5268–5277.
 146. Balabanian, K., Couderc, J., Bouchet-Delbos, L., Amara, A., Berrebi, D., Foussat, A., Baleux, F., Portier, A., Durand-Gasselin, I., Coffman, R. L., Galanaud, P., Peuchmaur, M., Emilie, D. (2003) Role of the chemokine stromal cell-derived factor 1 in autoantibody production and nephritis in murine lupus. *J. Immunol.* **170**, 3392–3400.
 147. Dziembowska, M., Tham, T. N., Lau, P., Vitry, S., Lazarini, F., Dubois-Dalq, M. (2005) A role for CXCR4 signaling in survival and migration of neural and oligodendrocyte precursors. *Glia* **50**, 258–269.
 148. Ambrosini, E., Remoli, M. E., Giacomini, E., Rosicarelli, B., Serafini, B., Lande, R., Aloisi, F., Coccia, E. M. (2005) Astrocytes produce dendritic cell-attracting chemokines in vitro and in multiple sclerosis lesions. *J. Neuropathol. Exp. Neurol.* **64**, 706–715.
 149. Calderon, T. M., Eugenini, E. A., Lopez, L., Kumar, S. S., Hesselgesser, J., Raine, C. S., Berman, J. W. (2006) A role for CXCL12 (SDF-1 α) in the pathogenesis of multiple sclerosis: regulation of CXCL12 expression in astrocytes by soluble myelin basic protein. *J. Neuroimmunol.* **177**, 27–39.
 150. Krumbholz, M., Theil, D., Cepok, S., Hemmer, B., Kivisakk, P., Ransohoff, R. M., Hofbauer, M., Farina, C., Derfuss, T., Hartle, C., Newcombe, J., Hohlfeld, R., Meinl, E. (2006) Chemokines in multiple sclerosis: CXCL12 and CXCL13 up-regulation is differentially linked to CNS immune cell recruitment. *Brain* **129**, 200–211.
 151. Yousry, T. A., Major, E. O., Ryschewitsch, C., Fahl, G., Fischer, S., Hou, J., Curfman, B., Miszkil, K., Mueller-Lenke, N., Sanchez, E., Barkhof, F., Radue, E. W., Jager, H. R., Clifford, D. B. (2006) Evaluation of patients treated with natalizumab for progressive multifocal leukoencephalopathy. *N. Engl. J. Med.* **354**, 924–933.
 152. Karpus, W. J., Kennedy, K. J. (1997) MIP-1 α and MCP-1 differentially regulate acute and relapsing experimental autoimmune encephalomyelitis as well as Th1/Th2 lymphocyte differentiation. *J. Leukoc. Biol.* **62**, 681–687.
 153. Lukacs, N. W., Chensue, S. W., Karpus, W. J., Lincoln, P., Keefer, C., Strieter, R. M., Kunkel, S. L. (1997) C-C chemokines differentially alter interleukin-4 production from lymphocytes. *Am. J. Pathol.* **150**, 1861–1868.
 154. Frigerio, S., Jun, T., Lu, B., Gerard, C., Zumsteg, U., Hollander, G. A., Piali, L. (2002) β Cells are responsible for CXCR3-mediated T-cell infiltration in insulinitis. *Nat. Med.* **8**, 1414–1420.
 155. Liu, L., Huang, D., Matsui, M., He, T. T., Hu, T., Demartino, J., Lu, B., Gerard, C., Ransohoff, R. M. (2006) Severe disease, unaltered leukocyte migration, and reduced IFN- γ production in CXCR3^{-/-} mice with experimental autoimmune encephalomyelitis. *J. Immunol.* **176**, 4399–4409.
 156. Tebo, J. M., Kim, H. S., Gao, J., Armstrong, D. A., Hamilton, T. A. (1998) Interleukin-10 suppresses IP-10 gene transcription by inhibiting the production of class I interferon. *Blood* **92**, 4742–4749.
 157. Zou, W., Machelon, V., Coulomb-LHermin, A., Borvak, J., Nome, F., Isaeva, T., Wei, S., Krzyzysk, B., Durand-Gasselin, I., Gordon, A., Pustilnik, T., Curiel, D. T., Galanaud, P., Capron, F., Emilie, D., Curiel, T. J. (2001) Stromal-derived factor-1 in human tumors recruits and alters the function of plasmacytoid precursor dendritic cells. *Nat. Med.* **7**, 1339–1346.
 158. Kukreja, P., Abdel-Mageed, A. B., Mondal, D., Liu, K., Agrawal, K. C. (2005) Up-regulation of CXCR4 expression in PC-3 cells by stromal-derived factor-1 α (CXCL12) increases endothelial adhesion and transendothelial migration: role of MEK/ERK signaling pathway-dependent NF- κ B activation. *Cancer Res.* **65**, 9891–9898.
 159. Taichman, R. S., Cooper, C., Keller, E. T., Pienta, K. J., Taichman, N. S., McCauley, L. K. (2002) Use of the stromal cell-derived factor-1/CXCR4 pathway in prostate cancer metastasis to bone. *Cancer Res.* **62**, 1832–1837.
 160. Darash-Yahana, M., Pikarsky, E., Abramovitch, R., Zeira, E., Pal, B., Karpus, R., Beider, K., Avniel, S., Kasem, S., Galun, E., Peled, A. (2004) Role of high expression levels of CXCR4 in tumor growth, vascularization, and metastasis. *FASEB J.* **18**, 1240–1242.
 161. Sun, Y. X., Wang, J., Shelburne, C. E., Lopatin, D. E., Chinnaiyan, A. M., Rubin, M. A., Pienta, K. J., Taichman, R. S. (2003) Expression of CXCR4 and CXCL12 (SDF-1) in human prostate cancers (PCa) in vivo. *J. Cell. Biochem.* **89**, 462–473.
 162. Akashi, T., Koizumi, K., Nagakawa, O., Fuse, H., Saiki, I. (2006) Androgen receptor negatively influences the expression of chemokine receptors (CXCR4, CCR1) and ligand-mediated migration in prostate cancer DU-145. *Oncol. Rep.* **16**, 831–836.
 163. Begley, L., Monteleon, C., Shah, R. B., Macdonald, J. W., Macoska, J. A. (2005) CXCL12 overexpression and secretion by aging fibroblasts enhance human prostate epithelial proliferation in vitro. *Aging Cell* **4**, 291–298.
 164. Engl, T., Relja, B., Marian, D., Blumenberg, C., Muller, I., Beecken, W. D., Jones, J., Ringel, E. M., Bereiter-Hahn, J., Jonas, D., Blaheta, R. A. (2006) CXCR4 chemokine receptor mediates prostate tumor cell adhesion through α 5 and β 3 integrins. *Neoplasia* **8**, 290–301.
 165. Muller, A., Homey, B., Soto, H., Ge, N., Catron, D., Buchanan, M. E., McClanahan, T., Murphy, E., Yuan, W., Wagner, S. N., Barrera, J. L., Mohar, A., Verastegui, E., Zlotnik, A. (2001) Involvement of chemokine receptors in breast cancer metastasis. *Nature* **410**, 50–56.
 166. Sun, Y. X., Fang, M., Wang, J., Cooper, C. R., Pienta, K. J., Taichman, R. S. (2007) Expression and activation of α (v) β (3) integrins by SDF-1/CXCL12 increases the aggressiveness of prostate cancer cells. *Prostate* **67**, 61–73.
 167. DiPersio, J. F., Uy, G. L., Yasothan, U., Kirkpatrick, P. (2009) Plerixafor. *Nat. Rev. Drug Discov.* **8**, 105–106.
 168. Galgani, M., Di Giacomo, A., Matarese, G., La Cava, A. (2009) The Yin and Yang of CD4(+) regulatory T cells in autoimmunity and cancer. *Curr. Med. Chem.* **16**, 4626–4631.
 169. Nurieva, R. I., Liu, X., Dong, C. (2009) Yin-Yang of costimulation: crucial controls of immune tolerance and function. *Immunol. Rev.* **229**, 88–100.
 170. Karin, N., Mitchell, D. J., Brocke, S., Ling, N., Steinman, L. (1994) Reversal of experimental autoimmune encephalomyelitis by a soluble peptide variant of a myelin basic protein epitope: T cell receptor antagonism and reduction of interferon γ and tumor necrosis factor α production. *J. Exp. Med.* **180**, 2227–2237.
 171. Brocke, S., Gijbels, K., Allegratta, M., Ferber, I., Piercy, C., Blankenstein, T., Martin, R., Utz, U., Karin, N., Mitchell, D., Veromaa, T., Waisman, A., Gaur, A., Conlon, P., Ling, N., Fairchild, P. J., Wraith, D. C., O'Garra, A., Fathman, C. G., Steinman, L. (1996) Treatment of experimental encephalomyelitis with a peptide analogue of myelin basic protein. *Nature* **379**, 343–346.
 172. Karin, N., Binah, O., Grabie, N., Mitchell, D. J., Felzen, B., Solomon, M. D., Conlon, P., Gaur, A., Ling, N., Steinman, L. (1998) Short peptide-based tolerogens without self-antigenic or pathogenic activity reverse autoimmune disease. *J. Immunol.* **160**, 5188–5194.

KEY WORDS:
 regulatory chemokines • Tr-1 cells • cancer • autoimmunity • experimental autoimmune encephalomyelitis (EAE) • M2 macrophages • D2 dendritic cells