

# Clinical implications of basic research: The role of hypocretin/orexin neurons in the central autonomic network

*Clinical & Translational Neuroscience*  
July-December 2018: 1–11  
© The Author(s) 2018  
Article reuse guidelines:  
sagepub.com/journals-permissions  
DOI: 10.1177/2514183X18789327  
journals.sagepub.com/home/ctn



Stefano Bastianini and Alessandro Silvani

## Abstract

Narcolepsy type 1 (NT1) and, to a lesser extent, neurodegenerative diseases, such as Alzheimer's disease, Parkinson's disease, dementia with Lewy bodies and multiple system atrophy, entail the loss of the hypothalamic neurons that release the hypocretin/orexin (H/O) neuropeptides. NT1 has been associated with autonomic anomalies including alterations in temperature regulation and cardiovascular control, particularly during sleep. A spectrum of autonomic dysfunctions also characterizes neurodegenerative diseases. The central autonomic network (CAN) is an interconnected set of brain structures that are critical for the control of autonomic preganglionic neurons. The H/O neurons include pre-autonomic neurons that directly target preganglionic sympathetic neurons in the intermediolateral column of the spinal cord and parasympathetic neurons in the dorsal motor nucleus of the vagus nerve. The H/O neurons also project to and modulate the activity of other CAN structures that include pre-autonomic neurons, such as the rostral ventromedial medulla and caudal raphe nuclei, the rostral ventrolateral medulla and the hypothalamic paraventricular nucleus. In addition, the H/O neurons project to and modulate the activity of neurons in the nucleus of the solitary tract in the medulla, which receives and relays visceral afferent information, and in higher order structures of the CAN, such as the dorsomedial nucleus of the hypothalamus and the extended amygdala. The H/O neurons should, therefore, be regarded as a key component of the CAN. Functional alterations of the CAN due to H/O neuron deficiency might contribute to autonomic anomalies in patients with neurodegenerative diseases and are likely to underlie autonomic anomalies in patients with NT1.

## Keywords

Orexins, hypocretins, narcolepsy, cataplexy, sympathetic, parasympathetic, Alzheimer's disease, Parkinson's disease

## Introduction

Orexin A and B (O-A and O-B, also called hypocretin 1 and 2, H-1 and H-2, respectively) are two hypothalamic neuropeptides independently discovered in 1998 by two different research groups, which employed hypothesis-free molecular biology techniques on rats.<sup>1,2</sup> These neuropeptides are synthesized and released by neurons in a restricted area of the tuberal region of the hypothalamus, caudal to the paraventricular nucleus (PVN).<sup>3</sup> The loss of hypocretin/orexin (H/O) neurons has been associated with the pathophysiology of narcolepsy with cataplexy,<sup>4</sup> which is presently referred to as narcolepsy type 1 (NT1).<sup>5</sup> Less severe losses of H/O neurons occur in neurodegenerative diseases including Alzheimer's disease (AD),<sup>6</sup> Parkinson's disease (PD),<sup>7</sup> dementia with Lewy bodies (DLB)<sup>8</sup> and multiple system atrophy (MSA).<sup>9</sup> To a different extent, NT1,<sup>10,11</sup> AD,<sup>12</sup> PD<sup>13</sup> and DLB<sup>13</sup> have all been associated with

alterations of the autonomic nervous system. The autonomic nervous system is controlled in the context of integrated behavioural responses by the central autonomic network (CAN), an interconnected set of brain structures that are also critical for the control of neuroendocrine, respiratory and sphincter motor neurons.<sup>14</sup> The aim of this narrative review is to discuss and put in a translational context the neuroanatomical and neurophysiological basic research that links the H/O neurons to the CAN.

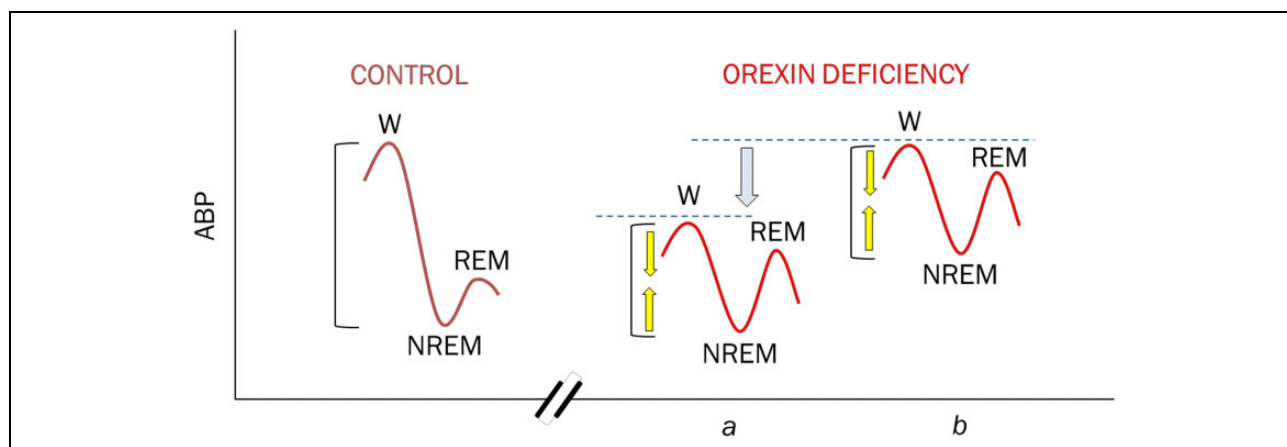
Department of Biomedical and Neuromotor Sciences, University of Bologna, Bologna, Italy

### Corresponding author:

Alessandro Silvani, Department of Biomedical and Neuromotor Sciences, University of Bologna, Piazza di Porta San Donato 2, 40126 Bologna, Italy. Email: [alessandro.silvani3@unibo.it](mailto:alessandro.silvani3@unibo.it)



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<http://www.creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).



**Figure 1.** Diagram showing the alterations of the control of ABP, which have been associated with H/O deficiency. In control subjects (left), ABP physiologically decreases from wakefulness (W) to NREM sleep and rises again during REM sleep. Hypocretin/orexin deficiency (orexin, right) has been found to entail a variable decrease in ABP during W (single arrow) coupled to a blunted decrease in ABP from W to NREM and (particularly) to REM (double arrows). H/O deficiency may thus be associated with ABP values that are lower than normal in W and normal during sleep (a) or normal in W and higher than normal during sleep (b). Reproduced from the study by Berteotti and Silvani<sup>10</sup> with permission. ABP: arterial blood pressure; NREM: non-rapid eye movement; REM: rapid eye movement.

## The H/O neurons

The H/O neurons project to<sup>3</sup> and receive synaptic inputs from<sup>15</sup> a wide array of central nervous system structures.<sup>3</sup> The electrical activity of the H/O neurons is mainly restricted to active wakefulness, although sparse activity bursts also occur during the so-called phasic periods of rapid-eye-movement (REM) sleep that entail bursts of eye movements and muscle twitches.<sup>16</sup> However, direct measurements with brain microdialysis in cats<sup>17</sup> and rats<sup>18</sup> have shown that the interstitial H-1/O-A concentration depends loosely, if at all, on the states of sleep and wakefulness in these species. In humans, H-1/O-A concentration in the cerebrospinal fluid (CSF) obtained by spinal tap is higher during the night than during the day.<sup>19</sup> Thus, for reasons that are still unclear, there seems to be a temporal mismatch between the electrical activity of the H/O neurons and the brain interstitial concentration of H-1/O-A. Corresponding data are lacking for H-2/O-B concentration. Further complexity is added by evidence that H/O neurons co-release dynorphin and glutamate, and potentially also nociceptin/orphanin FQ, galanin, neurotensin and gamma-aminobutyric acid (GABA), together with the H/O neuropeptides.<sup>20</sup> H-1/O-A binds either H/O receptor 1 or H/O receptor 2, whereas H-2/O-B is selective for H/O receptor 2.<sup>2</sup>

## H/O neuron loss and autonomic dysfunction in NT1

The brain of patients with NT1 shows a generalized absence of H/O neuropeptides<sup>4</sup> as well as the loss of other markers of the H/O neurons, such as dynorphin.<sup>21</sup> This indicates the functional loss of the whole H/O neurons, and not just of the H/O neuropeptides, possibly due to an autoimmune reaction.<sup>22</sup> Accordingly, present guidelines for

NT1 diagnosis indicate that a CSF concentration of H-1/O-A lower than one-third that expected in healthy subjects may substitute for findings of cataplexy, short sleep latency and sleep-onset periods of REM sleep.<sup>5</sup>

NT1 has been associated with autonomic anomalies including abnormal pupillary reaction during darkness, impaired nocturnal penile tumescence and alterations in temperature regulation and cardiovascular control.<sup>11</sup> Concerning cardiovascular control, NT1 may entail a blunted fall of arterial pressure from wakefulness to sleep, and particularly to REM sleep, coupled to a variable decrease in arterial pressure during wakefulness (Figure 1), and a tendency towards an increase in heart rate, particularly during sleep.<sup>10</sup> A substantial variability has been noted in the occurrence of these alterations, pointing to the existence of as-yet unknown biological modifiers of the effects of H/O deficiency on cardiovascular control.<sup>10</sup> Nonetheless, recent data indicate that middle-aged and elderly patients with narcolepsy had more health contacts than control patients with respect to several disease domains, such as diabetes, cardiovascular and gastrointestinal disorders,<sup>23</sup> which may result, at least in part, from altered autonomic control. Moreover, higher values of diastolic pressure and heart rate were recently found in patients with NT1 treated with psychostimulants than in untreated patients, suggesting an increased long-term risk of cardiovascular diseases.<sup>24</sup>

## H/O neuron loss and autonomic dysfunction in neurodegenerative diseases

AD is characterized by brain amyloid plaques, tau protein deposits forming neurofibrillary tangles and neurodegeneration.<sup>25</sup> Neurodegeneration in AD may also involve the H/O neurons, whose number was found decreased by 40%

in post-mortem brain analyses.<sup>6</sup> However, this decrease is either inconstant or associated with increased activity of the remaining H/O neurons, as the CSF levels of H-1/O-A in patients with AD have been found only slightly decreased<sup>6</sup> or even increased.<sup>26</sup> This may be of pathophysiological relevance because of preclinical evidence that H/O signalling increases amyloid neuropathology in AD models.<sup>27</sup> The occurrence of autonomic dysfunction in patients with AD has received relatively little attention. Nonetheless, one study reported mild-to-severe impairments in the blood pressure response to the Valsalva manoeuvre and in the heart rate response to isometric contraction in patients with mild-to-moderate AD, suggesting sympathetic dysfunction.<sup>28</sup> Accordingly, the sympathetic response to the stress of standing was also found impaired in patients with AD.<sup>12</sup>

PD and DLB are characterized by neurodegeneration and  $\alpha$ -synuclein deposits forming neuronal Lewy bodies and Lewy neurites.<sup>13</sup> Neurodegeneration in PD and DLB was found to entail a massive loss of H/O neurons, which reached a 62% cell loss in Braak stage V of PD,<sup>7</sup> and approximately a 75% cell loss in DLB.<sup>8</sup> The number of H/O neurons was also found reduced to approximately one-third in patients with MSA, a synucleinopathy characterized by neuroglial rather than neuronal cytoplasmic inclusions.<sup>13</sup> Nonetheless, the H/O neurons loss in patients with PD, DLB and MSA is generally insufficient to entail significant decreases in CSF levels of H-1/O-A.<sup>29–31</sup>

There is a lack of information on the involvement of the H/O system in pure autonomic failure, a related neurodegenerative disorder that entails peripheral accumulation of synuclein as neuronal cytoplasmic inclusions.<sup>13</sup> The synucleinopathies entail a spectrum of autonomic dysfunctions including orthostatic hypotension, thermoregulatory dysfunction, gastrointestinal dysmotility and urogenital dysfunction. The contribution of central versus peripheral neurodegeneration to these dysfunctions is thought to be the highest for MSA, intermediate for PD and DLB and the lowest for pure autonomic failure.<sup>13</sup>

## The CAN

The CAN includes the pre-sympathetic neurons (S0N) that project directly to the sympathetic preganglionic neurons (S1N) in the intermediolateral cell group (IML) of the spinal cord (Figure 2). Most of these S0N are located in the rostral ventromedial medulla (RVMM), the caudal raphe nuclei (raphe obscurus and pallidus), the rostral ventrolateral medulla (RVLM), the A5 area of the pons, the hypothalamic PVN and also the lateral hypothalamic area, which includes H/O neurons.<sup>32–34</sup> Comparatively less is known on the pre-parasympathetic neurons (P0N) that send direct projections to the parasympathetic preganglionic neurons (P1N) in the nucleus ambiguus (NAmb) and the dorsal motor nucleus of the vagus nerve (DMNX) of the medulla and to the P1N in the laminae V–VII of the sacral spinal cord (Figure 2). The NAmb and DMNX provide the

bulk of parasympathetic innervation to organs located above and below the diaphragm, respectively, whereas sacral P1N regulate lower gastrointestinal and urogenital functions.<sup>35</sup> Some of the P0N are localized in the PVN, the RVLM and the lateral and ventrolateral parts of the mid-brain periaqueductal grey (PAG) matter.<sup>36–39</sup> As it will be discussed in the following sections, there is evidence that P0N are also localized in the lateral hypothalamic area, which includes the H/O neurons.

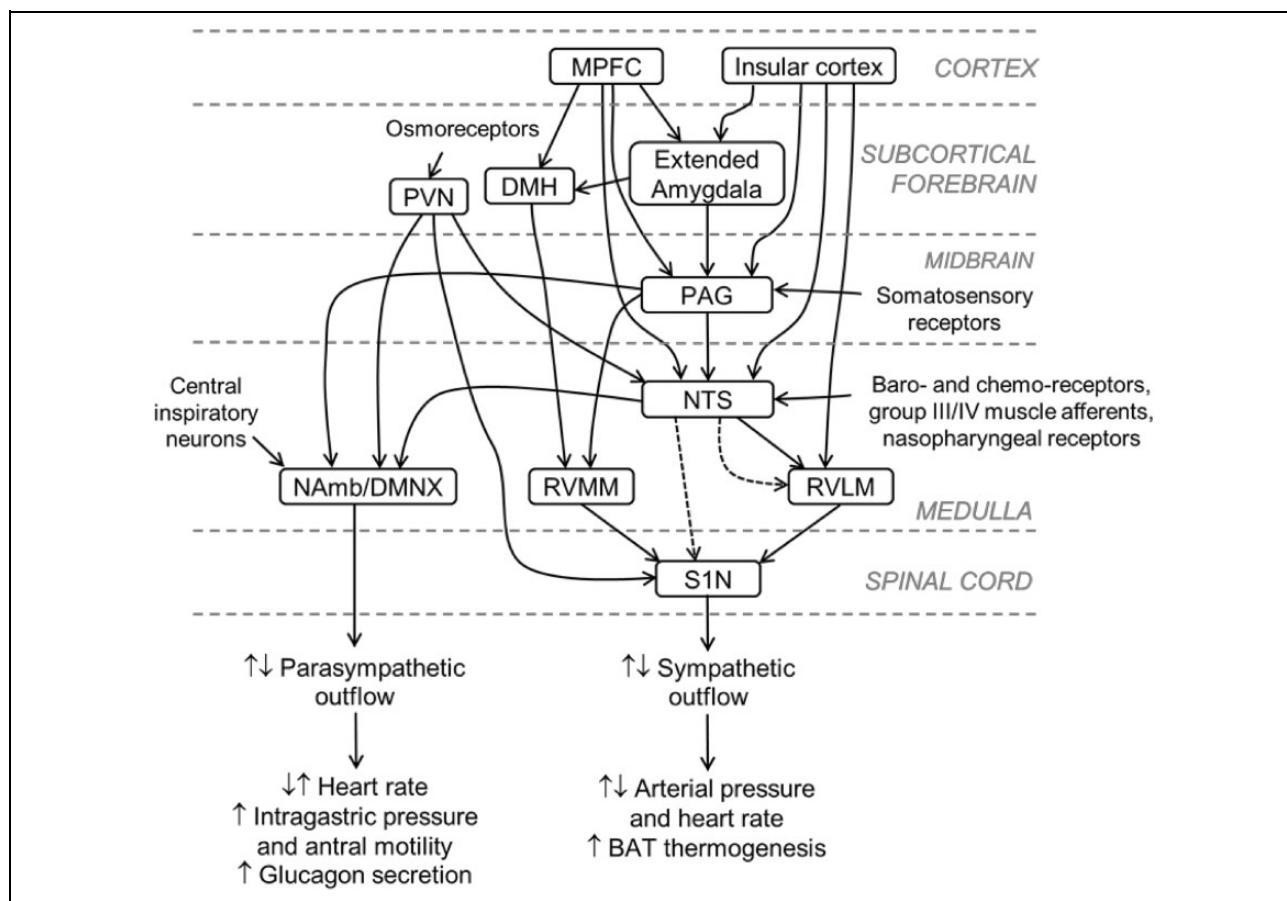
The S0N and P0N are themselves the target of synaptic projections from higher order structures of the CAN, which include the dorsomedial nucleus of the hypothalamus (DMH), the central nucleus of the amygdala, the bed nucleus of the stria terminalis (BNST), the medial prefrontal cortex (MPFC) and the insular cortex (Figure 2).<sup>14,34,40</sup> In addition to the S0N and P0N and their higher order controlling structures, the CAN includes the nucleus of the solitary tract (NTS) of the medulla, which is the main gateway for visceral afferents to the brain, and the parabrachial nucleus (PBN) of the pons, which contributes to relay the information from the NTS to the other structures of the CAN (Figure 2).<sup>14,34</sup>

## The H/O neurons include S0N

The H/O neurons have been reported to send long projections to all levels and all laminae of the spinal cord in mice, rats and human subjects, with a robust innervation of the IML in the thoracic and lumbar spinal cord, where S1N are localized.<sup>41</sup> A later study on rats confirmed H/O projections to the IML of spinal segments T1–L3, but found a progressive rostro-caudal decrease in the content of H-1/O-A and H-2/O-B measured with a radioimmunoassay on spinal cord tissue.<sup>42</sup> A more recent study on rats again confirmed H/O projections to S1N in the IML of the spinal segments T1 and T2, which are mainly involved in the control of the pupil size through the superior cervical ganglion, but reported that H/O projections were rare to segments T4–L2.<sup>43</sup>

From a functional perspective, intrathecal administration of either H-1/O-A or, to a lesser extent, H-2/O-B was found to increase arterial pressure and heart rate by enhancing sympathetic activity on  $\alpha_1$  and  $\beta_1$  adrenergic receptors.<sup>44</sup> Patch clamp recordings on spinal cord slices demonstrated that antidromically identified S1N could be directly activated by H-1/O-A or H-2/O-B.<sup>44</sup> This finding was later confirmed and expanded by the demonstration that the majority of H/O-sensitive S1N in the spinal cord express mRNA for both H/O receptors 1 and 2.<sup>45</sup> However, a more recent study that performed triple immunolabelling on rat spinal cords reported that most S1N express H/O receptor 1, but not H/O receptor 2.<sup>46</sup>

Taken together, these results indicate that H/O neurons include S0N that send direct projections to the S1N in the IML of the spinal cord. The H/O neurons are, therefore, a source of the descending projections to S1N previously recognized to originate from the lateral hypothalamic area (Figure 2).<sup>34</sup> While studies agree that H/O projections



**Figure 2.** Diagram showing the major central structures regulating autonomic outflow that receive projections from H/O neurons. No distinction is made between excitatory and inhibitory connections; dashed lines indicate indirect connections. The A5 area, the parabrachial nucleus of the pons and the sacral parasympathetic preganglionic neurons are omitted for the sake of clarity. Functional studies highlighted the potential for complex bidirectional effects (increases: ↑ or decreases: ↓) of H/O neurons on sympathetic and parasympathetic outflows (cf. the text for details). Only the key functional outcomes highlighted in the text are reported here. Reproduced with modification from the study by Silvani et al.<sup>40</sup> with permission. H/O: hypocretin/orexin; DMH: dorsomedial nucleus of the hypothalamus; DMNX: dorsal motor nucleus of the vagus nerve; MPFC: medial prefrontal cortex; NAmb: nucleus ambiguus; NTS: nucleus of the tractus solitarius (nucleus of the solitary tract); PAG: periaqueductal grey; PVN: hypothalamic paraventricular nucleus; RVLM: rostral ventrolateral medulla; RVMM: rostral ventromedial medulla; S1N: sympathetic preganglionic neurons.

target S1N in the upper thoracic segments of the spinal cord, the extent to which H/O projections target more caudal segments may be lower, and is still debated. The available evidence also indicates that H/O neurons control S1N at least in part by releasing H/O neuropeptides, as opposed to their co-transmitters. The evidence suggests that H-1/O-A binds to H/O receptor 1 in the spinal cord S1N, whereas the functional role of H-2/O-B and the expression of H/O receptor 2 on these S1N are less clear.

## The H/O neurons include P0N

### H/O projections to the NAmb

Fibres immunopositive to H-1/O-A were reported in the NAmb together with the expression of H/O receptor 1 within the neuropil, suggesting H/O modulation of presynaptic axonal terminals rather than of the NAmb neurons

themselves.<sup>47</sup> This may explain the discrepancy with the previous report of no detectable expression of mRNA for H/O receptor 1 in the rat NAmb.<sup>48</sup> Accordingly, microinjection of H-1/O-A in the rat NAmb was found to cause vagally mediated bradycardia and to potentiate the bradycardia driven by the baroreceptor reflex in response to a rise in arterial pressure.<sup>47</sup> These results were later confirmed and expanded by the observation of their enhancement by oestrogen in female rats.<sup>49</sup>

The potential for H/O projections to modulate P1N in the NAmb was also supported by patch-clamp recordings on brainstem slices from rats during early postnatal development. In particular, application of H-1/O-A to the bath of slices from rats at postnatal days 2–7 enhanced GABAergic and glycinergic inhibitory postsynaptic currents on P1N.<sup>50–52</sup> This effect was mediated by H/O receptor 1<sup>51,52</sup> and confirmed on a similar preparation by a different group.<sup>53</sup> Moreover, bath application of

H-1/O-A was found to inhibit glutamatergic excitatory postsynaptic currents on the NAmb P1N.<sup>50</sup> However, focal (as opposed to bath) application of H-1/O-A with a micropipette in the vicinity of NAmb P1N *inhibited* the GABAergic inhibitory postsynaptic currents in slices from rats at postnatal days 20 or 30, but not from rats at postnatal day 5.<sup>52</sup>

Taken together, these results indicate that H-1/O-A may decrease the activity of NAmb P1N by activating GABAergic and glycinergic neurons and/or by inhibiting glutamatergic neurons in other brainstem structures that project to the NAmb. H-1/O-A may also increase the activity of NAmb P1N by inhibiting GABA release from presynaptic terminals. The mechanisms whereby H-1/O-A, which is strongly neuroexcitatory,<sup>54</sup> would inhibit glutamatergic excitatory postsynaptic currents<sup>50</sup> or presynaptic GABA release<sup>52</sup> have not been clarified. Nevertheless, it seems fair to conclude that H-1/O-A has the potential for indirect, bidirectional effects on the P1N of the NAmb (Figure 2).

### *H/O projections to the DMNX*

The DMNX is targeted by a moderate number of H/O projection fibres.<sup>55</sup> Evidence obtained in different laboratories indicates that H-1/O-A and H-2/O-B directly excite P1N in the DMNX,<sup>56–58</sup> likely acting on both H/O receptors 1 and 2.<sup>59</sup> The H/O neurons, therefore, include P0N involved in the control of P1N of the DMNX (Figure 2). However, and somewhat paradoxically, H/O projections also enhance inhibitory inputs to the P1N of the DMNX that control the stomach.<sup>57,58</sup> In this respect, relatively few of the DMNX P1N that project to the antrum/pylorus, duodenum or caecum were reported as H/O responsive.<sup>57</sup> H/O projections may thus exert complex, bidirectional effects on the vagal control of gastric function, particularly at the level of the gastric fundus or corpus. Nonetheless, microinjection of either H-1/O-A or H-2/O-B in the DMNX was reported to elicit only increases in intragastric pressure and antral motility.<sup>59</sup>

Evidence obtained with trans-synaptic tracers indicates that the P1N of the DMNX that innervate the pancreas also receive synaptic projections from H/O neurons.<sup>60</sup> This innervation may be excitatory and functionally relevant, as microinjection of H-1/O-A in the DMNX dramatically increased vagal pancreatic nerve firing, whereas microinjection of an antagonist of H/O receptor 1 in the DMNX or chemical ablation of the lateral hypothalamic area, which includes H/O neurons, inhibited the increase in vagal pancreatic nerve firing elicited by insulin-induced hypoglycaemia.<sup>61</sup> Accordingly, microinjection of an antagonist of H/O receptor 1 in the DMNX prior to acute insulin administration decreased plasma glucose and suppressed glucagon secretion in rats.<sup>62</sup>

Although the DMNX P1N mainly target subdiaphragmatic structures, a subset of DMNX P1N with non-myelinated C-fibre axons target the heart and may regulate coronary blood flow and cardiac contractility to some extent.<sup>40</sup> Recent evidence indicates that these neurons are

also directly targeted by H/O neuron projections, which may be excitatory or inhibitory depending on whether the H/O co-transmitter released is glutamate or GABA.<sup>63</sup>

### *H/O projections to sacral P1N*

H/O projections to the sacral spinal cord have been described by two different reports,<sup>41,55</sup> suggesting a role of H/O peptides also in the control of sacral P1N. Electrophysiological and functional evidence of this role is lacking, however, and would be a relevant target of investigation.

## **The H/O neurons project to multiple CAN structures that include S0N and P0N**

### *H/O projections to the RVMM and the caudal raphe nuclei*

The RVMM and the caudal raphe nuclei include more S0N than any other brain region (Figure 2). These S0N mostly contain serotonin and are activated during arousal and stress, whereas they do not generate cardiac responses to baroreceptor or chemoreceptor inputs.<sup>33,40</sup> The S0N in the raphe pallidus are involved in the control of heat loss and brown adipose tissue thermogenesis.<sup>64</sup> The S0N in the raphe obscurus may play a role in the differential changes in regional sympathetic nerve activity (i.e. increases to skeletal muscles and decreases to kidneys) that occur during REM sleep.<sup>65,66</sup> These diverse functions may be modulated by H/O neurons through their projections to the RVMM,<sup>3,67,68</sup> the raphe pallidus<sup>3,68,69</sup> and the raphe obscurus,<sup>3,68</sup> often with direct contacts with the S0N.<sup>68,69</sup> There is evidence for expression of H/O receptor 1 in the RVMM,<sup>67</sup> and at least of its mRNA in the raphe obscurus.<sup>48</sup> Accordingly, H-1/O-A microinjection in the RVMM of rats was found to increase heart rate by increasing sympathetic activity and, at the same time, decreasing parasympathetic activity to the heart.<sup>67</sup> Nano-injection of H-1/O-A in the rostral raphe pallidus was reported to increase sympathetic activity to and thermogenesis by the brown adipose tissue.<sup>69</sup> This evidence indicates that the H/O projections to the RVMM and raphe pallidus may be functionally relevant. Corresponding functional evidence is lacking for H/O action on the raphe obscurus and would be an interesting topic of research, also considering the alterations in cardiovascular control often reported during REM sleep in patients with NT1.<sup>10</sup>

### *H/O projections to the RVLM*

The S0N in the RVLM are mainly glutamatergic and adrenergic neurons of the C1 cell group (Figure 2). These S0N play a key role in the expression of cardiovascular reflex responses to baroreceptor, chemoreceptor and other inputs.<sup>40</sup> Recent evidence indicates that C1 cells of the RVLM also play important roles in non-homeostatic,

adaptive responses to stress.<sup>70</sup> In addition, some C1 cells of the RVLM are P0N that control PIN in the DMNX.<sup>38</sup>

H/O neurons project to the RVLM.<sup>3,71</sup> Many putative C1 neurons of the RVLM express H/O receptors 1 and 2<sup>72</sup> and are in close proximity to H/O synaptic terminals.<sup>71,72</sup> On the other hand, C1 cells of the RVLM may also synapse with, and presumably excite, H/O neurons.<sup>73</sup>

Several lines of evidence suggest the functional relevance of these connections between H/O neurons and the RVLM. Microinjection of H-1/O-A in the rat RVLM was reported to increase arterial pressure, heart rate and splanchnic sympathetic nerve activity.<sup>71,72,74</sup> Microinjection of an antagonist of H/O receptor 1 in the RVLM decreased these responses to H-1/O-A partially, indicating that they were mediated by both H/O receptor types.<sup>72</sup> A similar conclusion was reached based on in-vitro recordings of RVLM neurons, suggesting, however, that H/O receptor 2 played the greater role.<sup>75</sup> Accordingly, H/O receptor 2 in the RVLM was reported to mediate the activation of adrenal nerve activity caused by H/O release in response to neuroglucoprivation.<sup>76,77</sup> Moreover, there is evidence that in the RVLM, signalling mediated by H/O receptor 2 contributes to hypertension in spontaneously hypertensive rats, a genetic model of essential hypertension.<sup>78</sup>

Experiments on brainstem slices indicated that H-1/O-A increased the frequency of spontaneous excitatory and inhibitory postsynaptic potentials in RVLM neurons, suggesting that H-1/O-A may also modulate transmitter release from presynaptic terminals in the RVLM.<sup>79</sup> This may contribute to explain why microinjection of H-1/O-A in the RVLM could exert bidirectional effects on cardiovascular reflex responses in anesthetized rats, increasing baroreflex sensitivity and sympathetic activity in conditions of hypoxia and decreasing the somato-sympathetic reflex and sympathetic activity in conditions of hypercapnia.<sup>72</sup>

### *H/O projections to the A5 noradrenergic cell group of the pons*

The S0N in the A5 area synthesize noradrenaline, give rise to one of the most extensive projections to the spinal cord IML and may contribute to cardiovascular and cardiorespiratory integration, although their precise functional role is still unclear.<sup>34</sup>

The A5 area receives dense descending H/O projections,<sup>3</sup> which, however, may be largely fibres of passage.<sup>80</sup> Nonetheless, at least H-1/O-A has the potential to modulate A5 neurons, as expression of H/O receptor 1 was documented in the A5 cell group at the mRNA<sup>48</sup> and protein levels.<sup>81</sup> Functional studies are needed to understand whether H/O neuropeptides actually play a role in modulating A5 cells.

### *H/O projections to the PVN*

The PVN is a master controller of the autonomic nervous system, which integrates neuroendocrine, homeostatic and

stress responses. The S1N in the PVN, many of which contain oxytocin, corticotropin-releasing factor, vasopressin or angiotensin, may be essential for generating cardiac sympathetic responses to changes in blood volume or plasma osmolality.<sup>40</sup>

A high H/O fibre density was reported in the PVN with immunohistochemistry,<sup>3,55</sup> and H/O content in the PVN was confirmed by radioimmunoassay.<sup>82</sup> The PVN was found to express H/O receptor 2 at mRNA<sup>48</sup> and protein<sup>83</sup> levels. Accordingly, electrophysiological evidence indicates that H-2/O-B, which is selective for H/O receptor 2,<sup>2</sup> depolarizes PVN neurons.<sup>84</sup> However, the expression of H/O receptor 1 was also reported in the PVN,<sup>85</sup> including on spinally-projecting PVN neurons.<sup>86</sup> Overexpression of H/O receptor 1 in the PVN of a rat model of obesity was inferred to play a causal role in increasing renal sympathetic nerve activity in that model.<sup>86</sup> Microinjection of H-1/O-A into the rat PVN was recently found to increase splanchnic and renal sympathetic nerve activity acting, at least in part, on H/O receptor 1.<sup>87</sup> On the other hand, optogenetic experiments indicate that H/O neurons target spinally projecting PVN neurons by co-releasing glutamate or GABA with H/O neuropeptides, which facilitate glutamatergic transmission at this level.<sup>88</sup> Thus, functional evidence supports the view that H/O neurons may exert a complex modulation of PVN S0N, at least in part through H/O receptor 1. Conversely, the role of H/O projections, if any, in modulating the P0N of the PVN that target the DMNX<sup>36,39</sup> is unknown and warrants further investigation (Figure 2).

### *H/O projections to the PAG*

The lateral and ventrolateral PAG projects to the DMNX and Namb.<sup>40</sup> A high density of H/O fibres,<sup>3</sup> high H/O neuropeptide content<sup>82</sup> and moderate expression of mRNA for both H/O receptors 1 and 2<sup>48</sup> were reported in the PAG. The ventrolateral and, to a much lower extent, the lateral PAG also send projections back to H/O neurons.<sup>15</sup> Non-ingestive effects of H/O neuropeptides resemble in many respects the effects mediated via the lateral and dorsolateral columns of the PAG, which consist in somatomotor activation, vigilance, tachycardia, hypertension and non-opioid analgesia.<sup>89</sup> In light of the multiple connections of the PAG with other CAN structures, it is therefore possible, but unproven, that the PAG is involved in H/O neuron modulation of parasympathetic as well as sympathetic activity (Figure 2).

## **The H/O neurons target CAN structures that receive and relay visceral afferent information**

### *H/O projections to the NTS*

The NTS relays visceral afferent information to the CAN through two parallel pathways: one targets the PBN and a

number of other circumscribed brain regions and the other starts from the PBN, and has projections that mirror those from the NTS.<sup>34</sup>

A high density of H/O fibres was found in the NTS,<sup>3,55,90,91</sup> which represent, in part, collaterals of projections to the NAmb.<sup>92</sup> The NTS also showed moderate H/O neuropeptide content,<sup>82</sup> moderate expression of mRNA for H/O receptor 1 and low expression of mRNA for H/O receptor 2.<sup>48</sup> Accordingly, either H-1/O-A<sup>93,94</sup> or H-2/O-B<sup>91</sup> was found to directly depolarize NTS neurons,<sup>93,94</sup> and H-2/O-B was also found to enhance excitatory synaptic activity on NTS neurons.<sup>91</sup> However, H-1/O-A microinjection in different NTS regions caused either pressor/tachycardic or depressor/bradycardic responses, the latter partially due to increased cardiac vagal activity, and potentiated the bradycardia to activation of arterial baroreceptors.<sup>90</sup> A potential explanation for these bidirectional effects is that H/O projections to the NTS may co-release H-1/O-A and leucine-enkephalin, which would enhance the depressor and bradycardic responses elicited by H-1/O-A at this level.<sup>95</sup> Another explanation points to a link with different H/O doses and mechanisms of action.<sup>96</sup> In particular, lower H/O doses in the NTS decreased heart rate and arterial pressure due to the activation of the soluble guanylate cyclase signalling pathway by the nitric oxide produced by neuronal nitric oxide synthase. Higher H/O doses in the NTS increased heart rate and arterial pressure by activating GABAergic or enhancing nitroergic neurotransmission.<sup>96</sup> These complexities notwithstanding, the available evidence strongly supports a modulatory role of H/O projection on the NTS (Figure 2).

### *H/O projections to the PBN*

Descending H/O fibres have been reported to the PBN,<sup>3</sup> including the Kolliker-Fuse subnucleus,<sup>97</sup> and the lateral PBN projects back to the H/O neurons.<sup>15</sup> However, no significant expression of mRNA for H/O receptor 1 and just a slight expression of mRNA for H/O receptor 2 above background were reported in the PBN.<sup>48</sup> Functional evidence on the role of H/O projections to the PBN is still lacking.

## **H/O projections to higher order structures of the CAN**

### *H/O projections to the DMH*

The DMH receives input from the amygdala and the insular cortex, projects to the caudal midline raphe and the RVMM and is critical for the expression of the cardiovascular response to stress (Figure 2).<sup>40</sup> The DMH receives abundant H/O projections<sup>3</sup> and includes neurons that project back to the H/O neurons, relaying information from the suprachiasmatic nucleus.<sup>55</sup> The DMH also includes some H/O neurons,<sup>3</sup> which contribute projections to the NTS/

DMNX region of the dorsal medulla.<sup>98</sup> The DMH has a moderate expression of mRNA for H/O receptor 2 and a low-density expression of mRNA for H/O receptor 1.<sup>48</sup> Accordingly, immunohistochemical studies reported expression of both H/O receptor 1<sup>85</sup> and H/O receptor 2<sup>83</sup> in the DMH, and a recent functional study indicated that injection of either H-1/O-A or H-2/O-B in the DMH increased arterial pressure and heart rate.<sup>99</sup>

### *H/O projections to the MPFC*

The MPFC comprises the anterior cingulate cortex and the prelimbic and infralimbic areas and is involved in both cognitive and visceromotor functions. The MPFC may increase parasympathetic activity and decrease vasomotor sympathetic activity by indirect pathways that involve the amygdala, PAG and NTS (Figure 2).<sup>40</sup> H/O projections target the MPFC,<sup>55,100</sup> which shows a mild expression of H/O receptor 1 at the mRNA<sup>48</sup> and protein<sup>85</sup> levels. H-2/O-B released by H/O projections to the thalamus also excites neurons of the thalamic paraventricular nucleus which send excitatory projections to the MPFC.<sup>101</sup> H/O thus have the potential to modulate the activity of the MPFC both directly and indirectly, but the functional consequences of this modulation still await investigation.

### *H/O projections to the insular cortex*

The insula is a viscerosensory and visceromotor region, which can increase or decrease heart rate depending on the stimulated site. It is not completely clear whether these changes in heart rate are due to sympathetic or parasympathetic effects, which may be exerted through indirect pathways, involving the amygdala, PAG and NTS, as well as through direct projections to the RVLM (Figure 2).<sup>40</sup> H/O neurons project to the insular cortex,<sup>55</sup> but the functional significance of these projections remains unclear. The insular cortex does not express mRNA for H/O receptor 2 and shows only a light expression of mRNA for H/O receptor 1,<sup>48</sup> which is consistent with other results on receptor protein expression.<sup>85</sup>

### *H/O projections to the extended amygdala*

The central nucleus of the amygdala and the BNST constitute an anatomo-functional unit called the extended amygdala.<sup>14</sup> The insular cortex and the extended amygdala constitute a corticostriatal-pallidal circuit that processes emotional information with autonomic responses (Figure 2) and projects to the hypothalamic behaviour control column.<sup>40</sup>

H/O neurons target the amygdala, including its central nucleus, as well as the BNST.<sup>3,55</sup> The central nucleus of the amygdala and the BNST project back to the H/O neurons.<sup>15</sup> Experiments on brain slices indicate that H/O neuropeptides excite neurons in the central medial nucleus of the



amygdala through H/O receptor 2.<sup>102</sup> Although an early report did not show any expression of mRNA for H/O receptor 2 in the central nucleus of the amygdala,<sup>48</sup> expression of this receptor at protein level was reported by a later study.<sup>83</sup> The involvement of H/O neuropeptides in mediating the cardiovascular responses elicited by the amygdala and the BNST is supported by the finding that stimulation of these structures induces long-lasting cardiorespiratory excitation in wild-type mice, but not in H/O-neuron ablated mice.<sup>103</sup> However, the results of a more recent study<sup>104</sup> highlighted the complexity of autonomic cardiovascular control by the BNST. In particular, chemical stimulation of the BNST was found to decrease arterial pressure and heart rate, and this effect was attenuated by H-1/O-A. Pharmacological experiments indicated that H-1/O-A facilitated noradrenaline release in the BNST. Noradrenaline in the BNST would activate local GABAergic neurons through  $\alpha$ -2 adrenergic receptors, and GABA released by these neurons would, in turn, inhibit a sympathoinhibitory pathway starting from the BNST by activating GABA<sub>A</sub> receptors.<sup>104</sup>

## Conclusions

The evidence discussed in the previous sections indicates that H/O neurons include S0N and P0N, as they send direct projections to S1N in the spinal cord and to P1N in the DMNX. The H/O neurons also project to other CAN structures that include S0N and P0N, such as the RVMM and caudal raphe nuclei, the RVLM and the hypothalamic PVN. While it is not always clear whether the H/O neurons directly target S0N and P0N in these structures or, rather, target neurons in their vicinity, the latter may nonetheless modulate S0N and P0N through local circuits. Therefore, the H/O neurons have the potential to control S1N and P1N both directly and indirectly, through other pre-autonomic neurons of the CAN. There is also abundant evidence that H/O neurons target the NTS, which is a key CAN structure involved in relaying visceral afferent input, and higher order CAN structures including the DMH and the extended amygdala. Significant gaps in our knowledge on the anatomic and/or functional links between H/O neurons and structures of the CAN persist, particularly regarding lower thoracic and lumbar S1N, sacral P1N, the medullary raphe obscurus, the pontine A5 group and PBN, the midbrain PAG, the MPFC and the insular cortex. Nonetheless, the available evidence appears sufficient to regard the H/O neurons as a key component of the CAN (Figure 2).

While a loss of H/O neurons has been reported in the brain of patients with different neurodegenerative diseases, such a loss is far from complete and may even be accompanied by hyperactivity of residual H/O neurons in patients with AD. Moreover, neurodegeneration in PD, DLB and MSA is known to involve the CAN and/or the peripheral autonomic nervous system, thus providing a parsimonious explanation of autonomic dysfunction. Nonetheless, given

the substantial role of H/O neurons in the CAN, alterations of H/O signalling have the potential to contribute, at least in part, to autonomic dysfunction in patients with neurodegenerative diseases. On the other hand, it appears biologically plausible that functional alterations of the CAN due to the dramatic H/O neuron deficiency in patients with NT1 underlie the autonomic anomalies reported in these patients, such as alterations in autonomic cardiovascular control during sleep (Figure 1).


## Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

## ORCID iD

Alessandro Silvani  <http://orcid.org/0000-0003-3992-3892>

## References

1. de Lecea L, Kilduff TS, Peyron C, et al. The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. *Proc Natl Acad Sci USA* 1998; 95: 322–327.
2. Sakurai T, Amemiya A, Ishii M, et al. Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* 1998; 92: 573–585.
3. Peyron C, Tighe DK, van den Pol AN, et al. Neurons containing hypocretin (orexin) project to multiple neuronal systems. *J Neurosci* 1998; 18: 9996–10015.
4. Peyron C, Faraco J, Rogers W, et al. A mutation in a case of early onset narcolepsy and a generalized absence of hypocretin peptides in human narcoleptic brains. *Nature Med* 2000; 6: 991–997.
5. Sateia MJ. *International classification of sleep disorders*. 3rd ed. Darien: American Academy of Sleep Medicine, Chest. 2014; 146(5): 1387–1394. doi: 10.1378/chest.14-0970
6. Fronczek R, van Geest S, Frolich M, et al. Hypocretin (orexin) loss in Alzheimer's disease. *Neurobiol Aging* 2012; 33: 1642–1650.
7. Thannickal TC, Lai YY and Siegel JM. Hypocretin (orexin) cell loss in Parkinson's disease. *Brain* 2007; 130: 1586–1595.
8. Kasanuki K, Iseki E, Kondo D, et al. Neuropathological investigation of hypocretin expression in brains of dementia with Lewy bodies. *Neurosci Lett* 2014; 569: 68–73.
9. Benarroch EE, Schmeichel AM, Sandroni P, et al. Involvement of hypocretin neurons in multiple system atrophy. *Acta Neuropathol* 2007; 113: 75–80.
10. Berteotti C and Silvani A. The link between narcolepsy and autonomic cardiovascular dysfunction: a translational perspective. *Clin Auton Res* 2017, in press. DOI: 10.1007/s10286-017-0473-z.



11. Grimaldi D, Silvani A, Benarroch EE, et al. Orexin/hypocretin system and autonomic control: new insights and clinical correlations. *Neurology* 2014; 82: 271–278.
12. Vitiello B, Veith RC, Molchan SE, et al. Autonomic dysfunction in patients with dementia of the Alzheimer type. *Biol Psychiatry* 1993; 34: 428–433.
13. Coon EA, Cutsforth-Gregory JK and Benarroch EE. Neuropathology of autonomic dysfunction in synucleinopathies. *Mov Disord* 2018; 33: 349–358.
14. Benarroch EE. The central autonomic network: functional organization, dysfunction, and perspective. *Mayo Clin Proc* 1993; 68: 988–1001.
15. Yoshida K, McCormack S, Espana RA, et al. Afferents to the orexin neurons of the rat brain. *J Comp Neurol* 2006; 494: 845–861.
16. Mileykovskiy BY, Kiyashchenko LI and Siegel JM. Behavioral correlates of activity in identified hypocretin/orexin neurons. *Neuron* 2005; 46: 787–798.
17. Kiyashchenko LI, Mileykovskiy BY, Maidment N, et al. Release of hypocretin (orexin) during waking and sleep states. *J Neurosci* 2002; 22: 5282–5286.
18. Yoshida Y, Fujiki N, Nakajima T, et al. Fluctuation of extracellular hypocretin-1 (orexin A) levels in the rat in relation to the light-dark cycle and sleep-wake activities. *Eur J Neurosci* 2001; 14: 1075–1081.
19. Grady SP, Nishino S, Czeisler CA, et al. Diurnal variation in CSF orexin-A in healthy male subjects. *Sleep* 2006; 29: 295–297.
20. Bonnavion P, Mickelsen LE, Fujita A, et al. Hubs and spokes of the lateral hypothalamus: cell types, circuits and behaviour. *J Physiol* 2016; 594: 6443–6462.
21. Crocker A, Espana RA, Papadopoulou M, et al. Concomitant loss of dynorphin, NARP, and orexin in narcolepsy. *Neurology* 2005; 65: 1184–1188.
22. Barateau L, Liblau R, Peyron C, et al. Narcolepsy type 1 as an autoimmune disorder: evidence, and implications for pharmacological treatment. *CNS Drugs* 2017; 31: 821–834.
23. Jennum P, Thorstensen EW, Pickering L, et al. Morbidity and mortality of middle-aged and elderly narcoleptics. *Sleep Med* 2017; 36: 23–28.
24. Bosco A, Lopez R, Barateau L, et al. Effect of psychostimulants on blood pressure profile and endothelial function in narcolepsy. *Neurology* 2018; 90: e479–e491.
25. Knopman DS, Haeblerlein SB, Carrillo MC, et al. The national institute on aging and the Alzheimer's association research framework for Alzheimer's disease: perspectives from the research roundtable. *Alzheimers Dement* 2018; 14: 563–575.
26. Gabelle A, Jaussent I, Hirtz C, et al. Cerebrospinal fluid levels of orexin-A and histamine, and sleep profile within the Alzheimer process. *Neurobiol Aging* 2017; 53: 59–66.
27. Roh JH, Jiang H, Finn MB, et al. Potential role of orexin and sleep modulation in the pathogenesis of Alzheimer's disease. *J Exp Med* 2014; 211: 2487–2496.
28. Jensen-Dahm C, Waldemar G, Staehelin Jensen T, et al. Autonomic dysfunction in patients with mild to moderate Alzheimer's disease. *J Alzheimers Dis* 2015; 47: 681–689.
29. Wienecke M, Werth E, Poryazova R, et al. Progressive dopamine and hypocretin deficiencies in Parkinson's disease: is there an impact on sleep and wakefulness? *J Sleep Res* 2012; 21: 710–717.
30. Baumann CR, Dauvilliers Y, Mignot E, et al. Normal CSF hypocretin-1 (orexin A) levels in dementia with Lewy bodies associated with excessive daytime sleepiness. *Eur Neurol* 2004; 52: 73–76.
31. Abdo WF, Bloem BR, Kremer HP, et al. CSF hypocretin-1 levels are normal in multiple-system atrophy. *Parkinsonism Relat Disord* 2008; 14: 342–344.
32. Strack AM, Sawyer WB, Hughes JH, et al. A general pattern of CNS innervation of the sympathetic outflow demonstrated by transneuronal pseudorabies viral infections. *Brain Res* 1989; 491: 156–162.
33. Dampney RA. Functional organization of central pathways regulating the cardiovascular system. *Physiol Rev* 1994; 74: 323–364.
34. Card JP and Sved AF. Central autonomic pathways. In: Llewellyn Smith IJ and Verberne AJM (eds) *Central regulation of autonomic functions*. 2nd ed. New York: Oxford University Press, 2011, pp. 3–22.
35. Jordan D. Parasympathetic preganglionic neurons. In: Llewellyn Smith IJ and Verberne AJM (eds) *Central regulation of autonomic functions*. 2nd ed. New York: Oxford University Press, 2011, pp. 120–139.
36. Portillo F, Carrasco M and Vallo JJ. Hypothalamic neuron projection to autonomic preganglionic levels related with glucose metabolism: a fluorescent labelling study in the rat. *Neurosci Lett* 1996; 210: 197–200.
37. Farkas E, Jansen AS and Loewy AD. Periaqueductal gray matter projection to vagal preganglionic neurons and the nucleus tractus solitarius. *Brain Res* 1997; 764: 257–261.
38. DePuy SD, Stornetta RL, Bochorishvili G, et al. Glutamatergic neurotransmission between the C1 neurons and the parasympathetic preganglionic neurons of the dorsal motor nucleus of the vagus. *J Neurosci* 2013; 33: 1486–1497.
39. Buijs RM, la Fleur SE, Wortel J, et al. The suprachiasmatic nucleus balances sympathetic and parasympathetic output to peripheral organs through separate preautonomic neurons. *J Comp Neurol* 2003; 464: 36–48.
40. Silvani A, Calandra-Buonaura G, Dampney RA, et al. Brain-heart interactions: physiology and clinical implications. *Philos Trans A Math Phys Eng Sci* 2016; 374: 20150181.
41. van den Pol AN. Hypothalamic hypocretin (orexin): robust innervation of the spinal cord. *J Neurosci* 1999; 19: 3171–3182.
42. Date Y, Mondal MS, Matsukura S, et al. Distribution of orexin-A and orexin-B (hypocretins) in the rat spinal cord. *Neurosci Lett* 2000; 288: 87–90.
43. Llewellyn-Smith IJ, Martin CL, Marcus JN, et al. Orexin-immunoreactive inputs to rat sympathetic preganglionic neurons. *Neurosci Lett* 2003; 351: 115–119.

44. Antunes VR, Brailoiu GC, Kwok EH, et al. Orexins/hypocretins excite rat sympathetic preganglionic neurons in vivo and in vitro. *Am J Physiol Regul Integr Comp Physiol* 2001; 281: R1801–R1807.
45. van den Top M, Nolan MF, Lee K, et al. Orexins induce increased excitability and synchronisation of rat sympathetic preganglionic neurones. *J Physiol* 2003; 549: 809–821.
46. Beig MI, Dampney BW and Carrive P. Both Ox1r and Ox2r orexin receptors contribute to the cardiovascular and locomotor components of the novelty stress response in the rat. *Neuropharmacology* 2015; 89: 146–156.
47. Ciriello J and de Oliveira CV. Cardiac effects of hypocretin-1 in nucleus ambiguus. *Am J Physiol Regul Integr Comp Physiol* 2003; 284: R1611–R1620.
48. Marcus JN, Aschkenasi CJ, Lee CE, et al. Differential expression of orexin receptors 1 and 2 in the rat brain. *J Comp Neurol* 2001; 435: 6–25.
49. de Oliveira CV and Ciriello J. Cardiovascular responses to hypocretin-1 in nucleus ambiguus of the ovariectomized female rat. *Brain Res* 2003; 986: 148–156.
50. Dergacheva O, Wang X, Huang ZG, et al. Hypocretin-1 (orexin-A) facilitates inhibitory and diminishes excitatory synaptic pathways to cardiac vagal neurons in the nucleus ambiguus. *J Pharmacol Exp Ther* 2005; 314: 1322–1327.
51. Dergacheva O, Philbin K, Bateman R, et al. Hypocretin-1 (orexin A) prevents the effects of hypoxia/hypercapnia and enhances the GABAergic pathway from the lateral paragigantocellular nucleus to cardiac vagal neurons in the nucleus ambiguus. *Neuroscience* 2011; 175: 18–23.
52. Dergacheva O, Bateman R, Byrne P, et al. Orexinergic modulation of GABAergic neurotransmission to cardiac vagal neurons in the brain stem nucleus ambiguus changes during development. *Neuroscience* 2012; 209: 12–20.
53. Wang JJ, Chen YH, Li KY, et al. Differential sensitivity of GABAergic and glycinergic inputs to orexin-A in preganglionic cardiac vagal neurons of newborn rats. *Acta Pharmacol Sin* 2005; 26: 1442–1447.
54. Kukkonen JP. Physiology of the orexinergic/hypocretinergic system: a revisit in 2012. *Am J Physiol Cell Physiol* 2013; 304: C2–32.
55. Date Y, Ueta Y, Yamashita H, et al. Orexins, orexigenic hypothalamic peptides, interact with autonomic, neuroendocrine and neuroregulatory systems. *Proc Natl Acad Sci USA* 1999; 96: 748–753.
56. Hwang LL, Chen CT and Dun NJ. Mechanisms of orexin-induced depolarizations in rat dorsal motor nucleus of vagus neurones in vitro. *J Physiol* 2001; 537: 511–520.
57. Grabauskas G and Moises HC. Gastrointestinal-projecting neurones in the dorsal motor nucleus of the vagus exhibit direct and viscerotopically organized sensitivity to orexin. *J Physiol* 2003; 549: 37–56.
58. Davis SF, Williams KW, Xu W, et al. Selective enhancement of synaptic inhibition by hypocretin (orexin) in rat vagal motor neurons: implications for autonomic regulation. *J Neurosci* 2003; 23: 3844–3854.
59. Krowicki ZK, Burmeister MA, Berthoud HR, et al. Orexins in rat dorsal motor nucleus of the vagus potentially stimulate gastric motor function. *Am J Physiol Gastr Liver Physiol* 2002; 283: G465–G472.
60. Buijs RM, Chun SJ, Nijijima A, et al. Parasympathetic and sympathetic control of the pancreas: a role for the suprachiasmatic nucleus and other hypothalamic centers that are involved in the regulation of food intake. *J Comp Neurol* 2001; 431: 405–423.
61. Wu X, Gao J, Yan J, et al. Hypothalamus-brain stem circuitry responsible for vagal efferent signaling to the pancreas evoked by hypoglycemia in rat. *J Neurophysiol* 2004; 91: 1734–1747.
62. Paranjape S, Vavaiya K, Kale A, et al. Role of dorsal vagal motor nucleus orexin-receptor-1 in glycemic responses to acute versus repeated insulin administration. *Neuropeptides* 2007; 41: 111–116.
63. Dergacheva O, Yamanaka A, Schwartz AR, et al. Direct projections from hypothalamic orexin neurons to brainstem cardiac vagal neurons. *Neuroscience* 2016; 339: 47–53.
64. Morrison SF. Central neural control of thermoregulation and brown adipose tissue. *Auton Neurosci* 2016; 196: 14–24.
65. Futuro Neto HA and Coote JH. Desynchronized sleep-like pattern of sympathetic activity elicited by electrical stimulation of sites in the brainstem. *Brain Res* 1982; 252: 269–276.
66. Silvani A and Dampney RA. Central control of cardiovascular function during sleep. *Am J Physiol Heart Circ Physiol* 2013; 305: H1683–H1692.
67. Ciriello J, Li Z and de Oliveira CV. Cardioacceleratory responses to hypocretin-1 injections into rostral ventromedial medulla. *Brain Res* 2003; 991: 84–95.
68. Berthoud HR, Patterson LM, Sutton GM, et al. Orexin inputs to caudal raphe neurons involved in thermal, cardiovascular, and gastrointestinal regulation. *Hist Cell Biol* 2005; 123: 147–156.
69. Tupone D, Madden CJ, Cano G, et al. An orexinergic projection from perifornical hypothalamus to raphe pallidus increases rat brown adipose tissue thermogenesis. *J Neurosci* 2011; 31: 15944–15955.
70. Stornetta RL and Guyenet PG. C1 neurons: a nodal point for stress? *Exp Physiol* 2018; 103: 332–336.
71. Machado BH, Bonagamba LG, Dun SL, et al. Pressor response to microinjection of orexin/hypocretin into rostral ventrolateral medulla of awake rats. *Regul Pept* 2002; 104: 75–81.
72. Shahid IZ, Rahman AA and Pilowsky PM. Orexin A in rat rostral ventrolateral medulla is pressor, sympatho-excitatory, increases barosensitivity and attenuates the somato-sympathetic reflex. *Br J Pharmacol* 2012; 165: 2292–2303.
73. Bochorishvili G, Nguyen T, Coates MB, et al. The orexinergic neurons receive synaptic input from C1 cells in rats. *J Comp Neurol* 2014; 522: 3834–3846.
74. Chen CT, Hwang LL, Chang JK, et al. Pressor effects of orexins injected intracisternally and to rostral ventrolateral medulla of anesthetized rats. *Am J Physiol Regul Integr Comp Physiol* 2000; 278: R692–R697.

75. Huang SC, Dai YW, Lee YH, et al. Orexins depolarize rostral ventrolateral medulla neurons and increase arterial pressure and heart rate in rats mainly via orexin 2 receptors. *J Pharmacol Exp Ther* 2010; 334: 522–529.
76. Korim WS, Bou Farah L, McMullan S, et al. Orexinergic activation of medullary premotor neurons modulates the adrenal sympathoexcitation to hypothalamic glucoprivation. *Diabetes* 2014; 63: 1895–1906.
77. Korim WS, Llewellyn-Smith IJ and Verberne AJ. Activation of medulla-projecting perifornical neurons modulates the adrenal sympathetic response to hypoglycemia: involvement of orexin type 2 (OX2-R) receptors. *Endocrinology* 2016; 157: 810–819.
78. Lee YH, Dai YW, Huang SC, et al. Blockade of central orexin 2 receptors reduces arterial pressure in spontaneously hypertensive rats. *Exp Physiol* 2013; 98: 1145–1155.
79. Dun NJ, Le Dun S, Chen CT, et al. Orexins: a role in medullary sympathetic outflow. *Regul Pept* 2000; 96: 65–70.
80. Puskas N, Papp RS, Gallatz K, et al. Interactions between orexin-immunoreactive fibers and adrenaline or noradrenaline-expressing neurons of the lower brainstem in rats and mice. *Peptides* 2010; 31: 1589–1597.
81. Darwinkel A, Stanic D, Booth LC, et al. Distribution of orexin-1 receptor-green fluorescent protein-(OX1-GFP) expressing neurons in the mouse brain stem and pons: colocalization with tyrosine hydroxylase and neuronal nitric oxide synthase. *Neuroscience* 2014; 278: 253–264.
82. Mondal MS, Nakazato M, Date Y, et al. Characterization of orexin-A and orexin-B in the microdissected rat brain nuclei and their contents in two obese rat models. *Neurosci Lett* 1999; 273: 45–48.
83. Cluderay JE, Harrison DC and Hervieu GJ. Protein distribution of the orexin-2 receptor in the rat central nervous system. *Regul Pept* 2002; 104: 131–144.
84. Shirasaka T, Miyahara S, Kunitake T, et al. Orexin depolarizes rat hypothalamic paraventricular nucleus neurons. *Am J Physiol Regul Integr Comp Physiol* 2001; 281: R1114–R1118.
85. Hervieu GJ, Cluderay JE, Harrison DC, et al. Gene expression and protein distribution of the orexin-1 receptor in the rat brain and spinal cord. *Neuroscience* 2001; 103: 777–797.
86. Zhou JJ, Yuan F, Zhang Y, et al. Upregulation of orexin receptor in paraventricular nucleus promotes sympathetic outflow in obese Zucker rats. *Neuropharmacology* 2015; 99: 481–490.
87. Fan Y, Jiang E, Hahka T, et al. Orexin A increases sympathetic nerve activity through promoting expression of proinflammatory cytokines in Sprague Dawley rats. *Acta Physiol (Oxf)* 2018; 222: e12963. DOI: 10.1111/apha.12963.
88. Dergacheva O, Yamanaka A, Schwartz AR, et al. Optogenetic identification of hypothalamic orexin neuron projections to paraventricular spinally projecting neurons. *Am J Physiol Heart Circ Physiol* 2017; 312: H808–H817.
89. Rodgers RJ, Ishii Y, Halford JC, et al. Orexins and appetite regulation. *Neuropeptides* 2002; 36: 303–325.
90. de Oliveira CV, Rosas-Arellano MP, Solano-Flores LP, et al. Cardiovascular effects of hypocretin-1 in nucleus of the solitary tract. *Am J Physiol Heart Circ Physiol* 2003; 284: H1369–H1377.
91. Smith BN, Davis SF, Van Den Pol AN, et al. Selective enhancement of excitatory synaptic activity in the rat nucleus tractus solitarius by hypocretin 2. *Neuroscience* 2002; 115: 707–714.
92. Ciriello J, McMurray JC, Babic T, et al. Collateral axonal projections from hypothalamic hypocretin neurons to cardiovascular sites in nucleus ambiguus and nucleus tractus solitarius. *Brain Res* 2003; 991: 133–141.
93. Yang B and Ferguson AV. Orexin-A depolarizes nucleus tractus solitarius neurons through effects on nonselective cationic and  $K^+$  conductances. *J Neurophysiol* 2003; 89: 2167–2175.
94. Yang B, Samson WK and Ferguson AV. Excitatory effects of orexin-A on nucleus tractus solitarius neurons are mediated by phospholipase C and protein kinase C. *J Neurosci* 2003; 23: 6215–6222.
95. Ciriello J, Caverson MM, McMurray JC, et al. Colocalization of hypocretin-1 and leucine-enkephalin in hypothalamic neurons projecting to the nucleus of the solitary tract and their effect on arterial pressure. *Neuroscience* 2013; 250: 599–613.
96. Shih CD and Chuang YC. Nitric oxide and GABA mediate bi-directional cardiovascular effects of orexin in the nucleus tractus solitarius of rats. *Neuroscience* 2007; 149: 625–635.
97. Lavezzi AM, Ferrero S, Roncati L, et al. Impaired orexin receptor expression in the Kolliker-Fuse nucleus in sudden infant death syndrome: possible involvement of this nucleus in arousal pathophysiology. *Neurol Res* 2016; 38: 706–716.
98. Harrison TA, Chen CT, Dun NJ, et al. Hypothalamic orexin A-immunoreactive neurons project to the rat dorsal medulla. *Neurosci Lett* 1999; 273: 17–20.
99. Li TL, Chen JYS, Huang SC, et al. Cardiovascular pressor effects of orexins in the dorsomedial hypothalamus. *Eur J Pharmacol* 2018; 818: 343–350.
100. Fadel J and Deutch AY. Anatomical substrates of orexin–dopamine interactions: lateral hypothalamic projections to the ventral tegmental area. *Neuroscience* 2002; 111: 379–387.
101. Huang H, Ghosh P and van den Pol AN. Prefrontal cortex-projecting glutamatergic thalamic paraventricular nucleus-excited by hypocretin: a feedforward circuit that may enhance cognitive arousal. *J Neurophysiol* 2006; 95: 1656–1668.
102. Bisetti A, Cvetkovic V, Serafin M, et al. Excitatory action of hypocretin/orexin on neurons of the central medial amygdala. *Neuroscience* 2006; 142: 999–1004.
103. Kuwaki T. Orexin links emotional stress to autonomic functions. *Auton Neurosci* 2011; 161: 20–27.
104. Ciriello J, Caverson MM and Li Z. Effects of hypocretin and norepinephrine interaction in bed nucleus of the stria terminalis on arterial pressure. *Neuroscience* 2013; 255: 278–291.