

available at www.sciencedirect.comjournal homepage: www.elsevier.com/locate/diabres

International Diabetes Federation

Fatty liver and chronic inflammation in Chinese adults

Pei-Wen Wang^{a,*}, Ching-Jung Hsieh^a, Leung-Chit Psang^b, Yu-Fan Cheng^b,
Chia-Wei Liou^c, Shao-Wen Weng^a, Jung-Fu Chen^a, I-Ya Chen^a,
Rong-Hwai Li^b, Hock-Liew Eng^d

^aDepartment of Internal Medicine, Chang Gung Memorial Hospital-Kaohsiung Medical Center,

Chang Gung University College of Medicine, 123 Ta-Pei Road, Niao-Sung Hsiang, Kaohsiung Hsien 83305, Taiwan

^bDepartment of Radiology, Chang Gung Memorial Hospital-Kaohsiung Medical Center, Chang Gung University College of Medicine, Kaohsiung, Taiwan

^cDepartment of Neurology, Chang Gung Memorial Hospital-Kaohsiung Medical Center, Chang Gung University College of Medicine, Kaohsiung, Taiwan

^dDepartment of Pathology, Chang Gung Memorial Hospital-Kaohsiung Medical Center, Chang Gung University College of Medicine, Kaohsiung, Taiwan

ARTICLE INFO

Article history:

Received 1 November 2007

Accepted 7 April 2008

Published on line 5 June 2008

Keywords:

Fatty liver

Insulin resistance

Metabolic syndrome

C-reactive protein

Adiponectin

ABSTRACT

Objective: To investigate the significance of fatty liver as predictor of insulin resistance (IR) and chronic inflammation.

Research design and methods: This cross-sectional study included 450 adults of Han Chinese origin aged ≥ 35 . Excluded were cases with hepatitis B or C, alcoholic liver disease, or currently using thiazolidinedione. The volunteers were screened for the presence of the components of metabolic syndrome (MtS). IR index was estimated by the homeostasis model assessment. The fatty liver index was evaluated by computed tomography, calculated as the liver/spleen (L/S) ratio arrived at by averaging Hounsfield values obtained for five 3-mm slices. Serum levels of adiponectin, C-reactive protein (CRP), leptin, interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α) were checked in 100 subjects with low-L/S ratio and 100 age- and sex-matched controls.

Results: Fatty liver index correlated with all MtS traits and IR index. The values of L/S ratios in subjects with 0, 1, 2, 3 and ≥ 4 traits of MtS were 1.25 ± 0.13 , 1.18 ± 0.16 , 1.12 ± 0.21 , 1.05 ± 0.25 and 0.92 ± 0.25 , respectively ($p < 0.001$). In our stepwise regression analysis to compare the L/S ratios to the conventional traits of MtS for association with adipokine dysregulation, we found L/S ratio to be independently associated with most of them: adiponectin ($p < 0.001$), CRP ($p < 0.001$), IL-6 ($p = 0.005$) and TNF- α ($p = 0.014$).

Conclusion: In Chinese, fatty liver index correlated well with IR index and can be a better marker of chronic inflammation than the conventional components of MtS.

© 2008 Elsevier Ireland Ltd. All rights reserved.

1. Introduction

All traits of metabolic syndrome (MtS) are risk factors for atherosclerosis [1–4]. Although the original conceptualization

of this syndrome was based on insulin resistance (IR) in the metabolic action of insulin, the pathogenesis remained unclear [1]. Accumulating evidence suggests that deranged adipocyte metabolism and altered body fat distribution are

* Corresponding author. Tel.: +886 7 7317123x8302; fax: +886 7 7322402.

E-mail addresses: wangpw@adm.cgmh.org.tw, c2607c@ms56.hinet.net (P.-W. Wang).

0168-8227/\$ – see front matter © 2008 Elsevier Ireland Ltd. All rights reserved.

doi:10.1016/j.diabres.2008.04.014

important determinants of IR [5–10]. Studies in lipotrophic patients [11,12] and fatless mice [13] have demonstrated that fat accumulation in liver and skeletal muscle is associated with IR. Fat accumulation in an insulin-sensitive tissue is an important determinant of the latter's sensitivity to insulin [5,6,14]. Therefore, fatty liver, or hepatic steatosis, is no longer considered a benign manifestation [15–18].

Recent understanding of the molecular events linking excessive fat to IR has focused on oxidative stress and chronic inflammation in accumulated adipose tissue [19–25]. This study is designed to examine the relationship between fatty liver, MtS and the adipokine regulation in the Chinese population.

2. Research design and methods

2.1. Subjects

Four hundred and fifty volunteers (90 with type 2 diabetes (T2DM), 44 with impair fasting glucose (IFG), and 316 with normal plasma glucose) participated in the study. Glucose dysregulation status was determined according to the 1997 diagnostic criteria of American diabetic association, which was defined as IFG when subjects with fasting plasma glucose (FPG) between 110 and 125 mg/dL (6.1–6.9 mmol/L) and defined as diabetes when subjects with FPG at least 126 mg/dL (≥ 7 mmol/L). The study group included 89 patients from the Meta/Endo clinic and 361 subjects from our health screening center, as well as some medical center employees. All subjects were 35 years old or older and of Han Chinese origin. They were excluded if they had type 1 diabetes, were being treated with thiazolidinedione, had secondary diabetes or hypertension caused by endocrinopathy or drug use, were carriers of hepatitis B or C, and were chronic alcohol drinkers.

The participants were divided into two groups: those who had MtS ($N = 156$) and those who did not ($N = 294$). Diagnosis of MtS was based on the definition by the National Cholesterol Education Program Adult Treatment Panel III (NCEP ATP III) [26,27]. The waist circumference cut-off point in the ATP III criteria was revised as suggested by the 2000 World Health Organization (WHO) Asia-Pacific Guidelines because the absolute risk of diabetes and cardiovascular disease (CVD) is higher in Asians who are less obese [28–30]. Participants were defined as having MtS if they had three or more of the following five components of metabolic syndrome: (1) waist circumference ≥ 90 cm in men and ≥ 80 cm in women, (2) triglycerides ≥ 150 mg/dL, (3) high-density lipoprotein (HDL) < 40 mg/dL in men and < 50 mg/dL in women, (4) blood pressure $\geq 130/85$ mmHg or if they were taking antihypertension medication and (5) fasting plasma glucose ≥ 110 mg/dL or if they were taking hypoglycemic medication.

2.2. Laboratory method

2.2.1. Measurement of clinical variables of cardiometabolic risks

We screened the fasting glucose, insulin, lipid profile and liver enzyme levels of all subjects. The blood pressure, body height, body weight, waist and hip circumferences were recorded. IR index in non-diabetic subjects was estimated by measuring

their fasting plasma glucose and insulin levels, and evaluating them by homeostasis model assessment [31].

2.2.2. Assessment of liver fat

Liver fat was assessed in all subjects by computed tomography (CT) examination. The examination was performed with a GE Prospeed (GE Medical System, Yokogawa, Japan) using 80 Kvp, 60 mAs, and slice thickness of 3 mm with a scan time of 1 s. Five slices were acquired from each individual on the basis of a scout view from the dome of the liver. The average of the five Hounsfield values obtained for each slice was calculated and reported as the computed tomography density (CTD) of liver for each test subject [32]. The ratio of liver to spleen (L/S ratio) for CT attenuation, as modified from the report of Kelly et al. [46], was determined by calculating the mean Hounsfield units of the liver (2 times of right lobe value + 1 time of left lobe value)/3, and that of the spleen. A L/S ratio < 1 was considered to be indicative of fatty liver.

2.2.3. Measurement of cytokines

One hundred subjects with the lowest L/S ratio and 100 age- and sex-matched controls were examined. None were taking statin, fibrate or thiazolidinedione. Adipokine level measurements were based on plasma that had been collected after overnight fasting and frozen. The specimens were frozen at -70 °C and measured in duplicate by immunoassay kits. The adiponectin kit (Quantikine, Human Adiponectin/Acrp30 Immunoassay), the interleukin-6 (IL-6) kit (Quantikine, Human IL-6 Immunoassay) and the tumor necrosis factor- α (TNF- α) kit (Quantikine, Human TNF- α /TNFSF1A Immunoassay), which were purchased from R&D Systems (Minneapolis, MN, USA) had a minimum detectable dosage of 0.246 ng/mL, 0.70 pg/mL and 1.6 pg/mL, respectively. The high-sensitive C-reactive protein (hs-CRP) was measured by an enzymatically amplified “two-step” sandwich-type immunoassay (US C-Reactive Protein ELISA, Diagnostic Systems Laboratories Inc., Texas, USA), which had a detection limit of 1.6 ng/mL. The leptin level was measured by a commercially available radioimmunoassay kit (Human Leptin RIA Kit, LINCO Research, Missouri, USA) with a detection limit of 0.5 ng/mL.

2.2.4. Statistical analysis

We performed logarithmic transformation of adipokine data because the original values were non-normally distributed. Continuous variables were expressed as means \pm S.D. Comparison of continuous and categorical data was performed using general linear model and Chi-square test, respectively. The relationships between the variables were analyzed by Pearson correlation and stepwise multiple regression analyses. A $p < 0.05$ was considered significant. All statistical operations were performed using the Statistical Package for Social Science Program (SPSS for Windows, Version 11.5; SPSS, Chicago).

3. Results

3.1. Prevalence of fatty liver

A L/S ratio < 1 was considered to indicate the presence of fatty liver. As such, the overall prevalence of fatty liver was 24.7%

Table 1 – Comparison of clinical characteristics of individuals with fatty liver (L/S < 1.0) and those without (L/S ≥ 1.0)

	L/S < 1.0	L/S ≥ 1.0	p
Age (year)	51.0 ± 9.8	50.4 ± 10.2	0.55
Sex (female/male)	62/49	210/129	0.25
Waist circumference (cm)	92.8 ± 9.9	87.6 ± 9.8	<0.001
BMI (kg/m ²)	27.1 ± 3.6	24.2 ± 3.3	<0.001
Hypertension (%)	55/111 (49.5%)	105/339 (30.9%)	0.001
Hyperglycemia (%)	57/111 (51.4%)	79/339 (23.3%)	<0.001
HDL-cholesterol (mg/dL)	42 ± 9	48 ± 11	<0.001
Triglyceride (mg/dL)	175 ± 108	114 ± 70	<0.001
LDL-cholesterol (mg/dL)	120 ± 33	117 ± 32	0.39
AST (U/L)	32 ± 24	23 ± 13	0.001
ALT (U/L)	42 ± 39	22 ± 17	<0.001

Hypertension subjects were taking antihypertension medication or their blood pressure was at least 130/85 mmHg. Hyperglycemic subjects were taking antidiabetic agents or their fasting plasma glucose was at least 110 mg/dL (7 mmol/L).

(111/450) in our series of individuals aged 35 or older. The clinical characteristics of individuals with or without fatty liver are listed in Table 1. The 156 subjects with MtS had a significantly greater percent (68/156; 43.6%) of fatty liver than the 294 subjects who did not have MtS (43/294; 14.6%) ($p < 0.001$). There was a significant difference in the prevalence of fatty liver in individuals with T2DM (44/90; 48.9%), IFG (12/44; 27.3%) and normal blood glucose (55/316; 17.4%) ($p < 0.001$). In all 450 volunteers, there was a good correlation between L/S ratios and serum levels of alanine transferase (ALT) ($r = -0.38$, $p < 0.001$).

3.2. Relation of fatty liver to the trait of metabolic syndrome

As can be seen in Table 2, the fatty liver index (L/S ratio) correlated with BMI ($r = -0.39$, $p < 0.001$), waist circumference ($r = -0.27$, $p < 0.001$), hypertension status ($r = -0.23$, $p < 0.001$), hyperglycemia status ($r = -0.27$, $p < 0.001$), high-density lipoprotein (HDL) ($r = 0.25$, $p < 0.001$) and triglyceride ($r = -0.31$, $p < 0.001$).

The values of L/S ratios in subjects with 0 ($N = 65$), 1 ($N = 111$), 2 ($N = 118$), 3 ($N = 85$) and ≥ 4 ($N = 71$) traits of MtS were 1.25 ± 0.13 , 1.18 ± 0.16 , 1.12 ± 0.21 , 1.05 ± 0.25 and 0.92 ± 0.25 , respectively. The difference was significant ($p < 0.001$).

The prevalence of fatty liver in subjects with 0, 1, 2, 3 and ≥ 4 traits of MtS were 6.2% (4/65), 9.9% (11/111), 23.7% (28/118),

32.9% (28/85) and 56.3% (40/71), also statistically significant ($p < 0.001$).

In non-diabetic subjects, IR index estimated by the homeostasis model assessment correlated with the L/S ratio very well ($r = -0.21$, $p < 0.001$).

3.3. Physiological significance of fatty liver in subjects with diabetes and metabolic syndrome

In the subgroup with T2DM, those with fatty liver were found to have increased body mass index (BMI) (27.4 ± 3.7 kg/m² vs. 25.3 ± 4.1 kg/m², $p = 0.015$) and elevated levels of plasma triglyceride (188.9 ± 130.6 mg/dL vs. 120.1 ± 61.2 mg/dL, $p = 0.002$) and ALT (46.5 ± 28.5 U/L vs. 27.1 ± 17.2 U/L, $p < 0.001$) compared with T2DM patients without fatty liver.

In the subgroup with MtS, more subjects with fatty liver had a fasting glucose level >110 mg/dL than MtS subjects without fatty liver (44/68 = 64.7% vs. 40/88 = 45.5%, $p = 0.02$).

3.4. The relationship of inflammatory cytokines to metabolic variables

The relationships between adipokines and metabolic variables are shown in Table 3. Leptin correlated with BMI ($r = 0.42$, $p < 0.001$) and waist circumference ($r = 0.20$, $p = 0.04$). Adiponectin correlated with HDL ($r = 0.38$, $p < 0.001$), L/S ratio ($r = 0.35$, $p < 0.001$), BMI ($r = -0.20$, $p = 0.005$) and triglyceride

Table 2 – Relationship of fatty liver to metabolic variables in the 450 cases

	BMI	Waist	Hypertension	Hyperglycemia	HDL	TG	L/S ratio	IR
BMI	1							
Waist	0.68**	1						
Hypertension	0.32**	0.25**	1					
Hyperglycemia	0.25**	0.20**	0.21**	1				
HDL	-0.38**	-0.33**	-0.16**	-0.16**	1			
TG	0.26**	0.23**	0.10*	0.16**	-0.42**	1		
L/S ratio	-0.39**	-0.27**	-0.23**	-0.27**	0.25**	-0.31**	1	
IR	0.36**	0.26**	0.18**	0.37**	-0.25**	0.31**	-0.21**	1

Hypertension subjects were taking antihypertension medication or their blood pressure was at least 130/85 mmHg. Hyperglycemic subjects were taking antidiabetic agents or their fasting plasma glucose was at least 110 mg/dL (7 mmol/L). IR index by the homeostasis model assessment was estimated in non-diabetic subjects only.

* p value < 0.05 .

** p value < 0.01 .

Table 3 – Relationship of inflammatory cytokines to metabolic variables

	BMI	Waist	Hypertension	Hyperglycemia	HDL	TG	L/S ratio	IR
Adiponectin	−0.20**	−0.09	−0.13	−0.10	0.38**	−0.15*	0.35**	−0.21**
CRP	0.38**	0.24**	0.15*	0.14	−0.13	0.10	−0.29**	0.22**
Leptin	0.42**	0.20**	−0.09	−0.07	0.07	0.07	−0.12	0.28**
IL-6	0.30**	0.25**	0.17*	0.04	−0.18*	−0.02	−0.23**	0.15
TNF- α	0.13	0.03	0.03	0.07	−0.11	0.09	−0.17*	0.04

Hypertension subjects were taking antihypertension medication or their blood pressure was at least 130/85 mmHg. Hyperglycemic subjects were taking antidiabetic agents or their fasting plasma glucose was at least 110 mg/dL (7 mmol/L). IR index by the homeostasis model assessment was estimated in non-diabetic subjects only.

* p value < 0.05.
 ** p value < 0.01.

($r = -0.15$, $p = 0.03$). CRP correlated with BMI ($r = 0.38$, $p < 0.001$), L/S ratio ($r = -0.29$, $p < 0.001$), waist circumference ($r = 0.24$, $p = 0.001$) and hypertension ($r = 0.15$, $p = 0.04$). IL-6 correlated with BMI ($r = 0.30$, $p < 0.001$), waist circumference ($r = 0.25$, $p < 0.001$), L/S ratio ($r = -0.23$, $p = 0.001$), HDL ($r = -0.18$, $p = 0.01$) and hypertension ($r = 0.17$, $p = 0.02$). TNF- α correlated with L/S ratio ($r = -0.17$, $p = 0.02$).

3.5. Independent predictor of adipokines

We used stepwise regression analysis to compare the fatty liver index to the five known components of MtS by ATP III definition and clarify their association with adipokine dysregulation. HDL ($p < 0.001$) and L/S ratio ($p < 0.001$) were independently related to serum level of adiponectin (for the entire model, $R^2 = 0.21$). L/S ratio ($p < 0.001$) and waist circumference ($p = 0.018$) were independent predictors of serum level of hs-CRP (for the entire model, $R^2 = 0.11$). L/S ratio ($p = 0.005$) and waist circumference ($p = 0.007$) were independent predictors of serum level of IL-6 (for the entire model, $R^2 = 0.095$). Waist circumference ($R^2 = 0.042$, $p = 0.004$) was independent predictor of serum level of leptin. L/S ratio ($R^2 = 0.03$, $p = 0.014$) was independent predictor of serum level of TNF- α . Among the five cytokines assayed, four, three and one were independently predicted by L/S ratio, waist circumference and HDL, respectively. The L/S ratio could predict more number of adipokines as well as the most informative adipokines (adiponectin and hsCRP) in this study. It was a better predictor of adipokines dysregulation than the five known components of MtS defined by ATP III.

4. Discussion

Despite the ongoing debate regarding the clinical use of MtS [33–35], its constituent risk factors are well recognized as predictors of cardiovascular disease and T2DM [36–39]. As cardiometabolic risk factors tend to cluster, patients often have additional subclinical features that can be discovered through comprehensive evaluation. This study provides data regarding fatty liver in a Chinese population in Taiwan. Like a previous study of Chinese population in Shanghai [40], it found a strong association between fatty liver and the risk factors characteristic of MtS.

Based on our studies using CT imaging, there is a much higher prevalence of fatty liver in patients with type 2 diabetes

(48.9%) and MtS (43.6%) than in those without these metabolic defects (14.6–17.4%) in the people of Han Chinese origin in Taiwan. The prevalence was similar to that observed by a study using ultrasonography in a Chinese population in Shanghai (type 2 diabetes, 41.2%; MtS, 48.2%) [40], though our results were more precisely quantitated using L/S ratio. We also found a significant association between fatty liver index and individual traits of MtS, including waist circumference, hypertension status, a fasting glucose level >110 mg/dL, plasma HDL and triglyceride levels. Furthermore, the more components of MtS present in an individual, the lower the fatty liver index, suggesting that the fatty liver index may be able to reflect the severity of this metabolic disorder quantitatively. This quantitative index can be well estimated by our routine liver function test of ALT. Our findings support the data of Vozarova et al. [41], in which elevated ALT levels in cases of nonalcoholic fatty liver disease (NAFLD) were associated with decreased hepatic insulin sensitivity and predictive of future development of diabetes. These observations are enforced by a recent publication of Lee et al. [42] who pointed out that serum gamma-glutamyl transferase (GGT) predicts CVD morbidity and mortality, suggesting that GGT can be used as a marker of metabolic and cardiovascular risk. Haffner further proposed that elevated liver enzymes, an indicator of non-alcoholic fatty liver disease (NAFLD), may comprise an additional component of the MtS [43].

The ATP III criteria have been evaluated by Cheal et al. for their ability to identify IR, and they found that the individual components varied in their relationship to IR, with the obesity and lipid criteria being most useful [44]. In our series, individuals with T2DM and fatty liver (L/S ratio <1.0) had a significantly greater increase in BMI and plasma triglyceride levels than such patients without fatty liver. We also found that a greater percentage of individuals with MtS and fatty liver had a fasting glucose >110 mg/dL than MtS individuals without it. These data suggest that fatty liver index may provide additional information about IR for those with diabetes or MtS when diagnosed using the ATP III criteria. Toledo et al. in a study using CT imaging has recently reported that hepatic steatosis influenced the severity and composition of dyslipidemia in T2DM patients by increasing triglycerides, reducing HDL levels and increasing small, dense LDL [45]. Using the more sophisticated euglycemic insulin infusion for insulin sensitivity measurement and ($^2\text{H}_2$) glucose infusion to assess endogenous glucose production, Kelly et al. [46] have also demonstrated that individuals with T2DM and fatty liver

were more insulin resistant and had more severe dyslipidemia than those who had diabetes but no fatty liver.

Taking an adipocentric point of view, we found that fatty liver index, waist circumference and serum HDL level were better predictors of the proinflammatory adipokines than hyperglycemia and hypertension. With regard to fatty liver index and waist circumference, fatty liver index could independently predict adiponectin (but not leptin), while waist circumference could independently predict leptin (but not adiponectin). The relationship between adiponectin and fatty liver in our study has been supported by several previous clinical studies [47–50]. The role of leptin in liver injury is less frequently mentioned [51], though it has been suggested that the adiponectin–leptin ratio is a more efficacious measure of IR [52].

Leptin is secreted from adipocytes in proportion to adipose tissue mass [53]. What is known about its relationship with IR comes from studies of rare leptin-deficiency patients with lipodystrophy [11,12]; most obese insulin resistant individuals are not in the same condition. Circulating adiponectin levels, on the contrary, correlate more with hyperinsulinemia and IR than obesity or body fat [54]. Although IR and obesity are both associated with lower plasma adiponectin concentrations, Abbasi et al. have demonstrated that adiponectin levels were more closely related to differences in insulin-mediated glucose disposal than BMI [55]. Furthermore, low-plasma adiponectin has been found to be an independent risk factor for future development of T2DM [56,57], not for obesity [58]. The beneficial effect of thiazolidinedione on insulin sensitivity and reversing the abnormality in hepatic fat mobilization is proved to be related to its ability to increase plasma adiponectin concentrations [59]. In addition to being able to predict adiponectin, we found the fatty liver index independently predict serum hs-CRP levels. It has been reported that mild chronic elevations of CRP concentrations are independently predictive of future CVD events both in women and men [60,61]. Recently, in the Framingham Offspring Study of 3037 subjects, CRP has also been proved to predict new CVD events over a 7-year follow-up [62]. Taken together, these findings support the notion that fatty liver plays an important role in the chronic inflammation triggering of CVD.

In conclusion, in the people of Han Chinese origin in Taiwan, the fatty liver index can be a quantitative index for MtS and may be a good marker of IR related to chronic inflammation than the currently recognized components of MtS.

Acknowledgements

This work was supported by research grant (CMRPG8065) from Chang Gung Memorial Hospital-Kaohsiung Medical Center, Chang Gung University College of Medicine, Taiwan.

Conflict of interest

There are no conflicts of interest.

REFERENCES

- [1] G.M. Reaven, Role of insulin resistance in human disease, *Diabetes* 37 (1988) 1595–1607.
- [2] S.M. Haffner, R.A. Valdez, H.P. Hazuda, B.D. Mitchell, P.A. Morales, M.P. Stern, Prospective analysis of the insulin-resistance syndrome (syndrome X), *Diabetes* 41 (1992) 715–722.
- [3] B. Isomaa, P. Almgren, T. Tuomi, B. Forsen, K. Lahti, M. Nissen, et al., Cardiovascular morbidity and mortality associated with the metabolic syndrome, *Diab. Care* 24 (2001) 683–689.
- [4] M. Trevisan, J. Liu, F.B. Bahsas, A. Menotti, Syndrome X and mortality: a population-based study, *Am. J. Epidemiol.* 148 (1998) 958–966.
- [5] D.E. Kelly, B.H. Goodpaster, Skeletal muscle triglyceride, *Diab. Care* 24 (2001) 933–941.
- [6] A. Seppälä-Lindroos, S. Vehkavaara, A.M. Häkkinen, T. Goto, J. Westerbacka, A. Sovijärvi, et al., Fat accumulation in the liver is associated with defects in insulin suppression of glucose production and serum free fatty acids independent of obesity in normal men, *J. Clin. Endocrinol. Metab.* 87 (2002) 3023–3028.
- [7] E. Ravussin, S.R. Smith, Increased fat intake, impaired fat oxidation, and failure of fat cell proliferation result in ectopic fat storage, insulin resistance, and type 2 diabetes mellitus, *Ann. NY Acad. Sci.* 967 (2002) 363–378.
- [8] H. Bays, L. Mandarino, R.A. DeFronzo, Role of adipocyte, free fatty acid, and ectopic fat in pathogenesis of type 2 diabetes mellitus: peroxisomal proliferator-activated receptor agonists provide a rational therapeutic approach, *J. Clin. Endocrinol. Metab.* 89 (2004) 463–478.
- [9] B.H. Goodpaster, J. He, S. Watkins, D.E. Kelly, Skeletal muscle lipid content and insulin resistance: evidence for a paradox in endurance-trained athletes, *J. Clin. Endocrinol. Metab.* 86 (2001) 5755–5761.
- [10] J.J. Puder, S. Varga, M. Kraenzlin, C. De Geyter, U. Keller, B. Muller, Central fat excess in polycystic ovary syndrome: relation to low-grade inflammation and insulin resistance, *J. Clin. Endocrinol. Metab.* 90 (2005) 6014–6021.
- [11] K.F. Petersen, E.A. Oral, S. Dufour, D. Befroy, C. Ariyan, C. Yu, et al., Leptin reverses insulin resistance and hepatic steatosis in patients with severe lipodystrophy, *J. Clin. Invest.* 109 (2002) 1345–1350.
- [12] J.H. Lee, J.L. Chan, E. Sourlas, V. Raptopoulos, C.S. Mantzoros, Recombinant methionyl human leptin therapy in replacement doses improves insulin resistance and metabolic profile in patients with lipoatrophy and metabolic syndrome induced by the highly active antiretroviral therapy, *J. Clin. Endocrinol. Metab.* 91 (2006) 2605–2611.
- [13] J.K. Kim, O. Gavrilova, Y. Chen, M.L. Reitman, G.I. Shulman, Mechanism of insulin resistance in A-ZIP/F-1 fatless mice, *J. Biol. Chem.* 275 (2000) 8456–8460.
- [14] K.F. Petersen, S. Dufour, D. Befroy, R. Garcia, G.I. Shulman, Impaired mitochondrial activity in the insulin-resistant offspring of patients with type 2 diabetes, *N. Engl. J. Med.* 350 (2004) 664–671.
- [15] A.J. Sanyal, C. Campbell-Sargent, F. Mirshahi, W.B. Rizzo, M.J. Contos, R.K. Sterling, et al., Nonalcoholic steatohepatitis: association of insulin resistance and mitochondrial abnormalities, *Gastroenterology* 120 (2001) 1183–1192.
- [16] G. Marchesini, M. Brizi, G. Bianchi, S. Tomassetti, E. Bugianesi, M. Lenzi, et al., Nonalcoholic fatty liver disease: a feature of the metabolic syndrome, *Diabetes* 50 (2001) 1844–1850.

- [17] S. Chitturi, S. Abeygunasekera, G.C. Farrell, J. Holmes-Walker, J.M. Hui, C. Fung, et al., NASH and insulin resistance: insulin hypersecretion and specific association with insulin resistance syndrome, *Hepatology* 35 (2002) 373–379.
- [18] G. Pagano, G. Pacini, G. Musso, R. Gambino, F. Mecca, N. Depetris, et al., Nonalcoholic steatohepatitis, insulin resistance, and metabolic syndrome: further evidence for an etiologic association, *Hepatology* 35 (2002) 367–372.
- [19] A. Festa, R. D'Agostino, G. Howard, L. Mykkanen, R.P. Tracy, S.M. Haffner, Chronic subclinical inflammation as part of the insulin resistance syndrome. The Insulin Resistance Atherosclerosis Study (IRAS), *Circulation* 102 (2000) 42–47.
- [20] A.G. Pittas, N.A. Joseph, A.S. Greenberg, Adipocytokines and insulin resistance I, *Clin. Endocrinol. Metab.* 89 (2004) 447–452.
- [21] K.E. Wellen, G.S. Hotamisligil, Inflammation, stress, and diabetes, *J. Clin. Invest.* 115 (2005) 1111–1119.
- [22] S. Furukawa, T. Fujita, M. Shimabukuro, M. Iwaki, Y. Yamada, Y. Nakajima, et al., Increased oxidative stress in obesity and its impact on metabolic syndrome, *J. Clin. Invest.* 114 (2004) 1752–1761.
- [23] S.E. Shoelson, J. Lee, A.B. Goldfine, Inflammation and insulin resistance, *J. Clin. Invest.* 116 (2006) 1793–1801.
- [24] B.B. Kahn, J.S. Flier, Obesity and insulin resistance, *J. Clin. Invest.* 106 (2000) 473–481.
- [25] P. Dandona, A. Aljada, A. Chaudhuri, P. Mohanty, R. Garg, Metabolic syndrome. A comprehensive perspective based on interaction between obesity, diabetes, and inflammation, *Circulation* 111 (2005) 1448–1454.
- [26] Executive Summary of The Third Report of The National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III), *JAMA* 285 (2001) 2486–2497.
- [27] S.M. Grundy, H.B. Brewer, J.I. Cleeman, S.C. Smith, C. Lenfant, Definition of metabolic syndrome: report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition, *Circulation* 109 (2004) 433–438.
- [28] T.P. Gill, Cardiovascular risk in the Asia-Pacific region from a nutrition and metabolic point of view: abdominal obesity, *Asia Pac. J. Clin. Nutr.* 10 (2001) 85–89.
- [29] World Health Organization, The Asia-Pacific Perspective: Redefining Obesity and its Treatment, WHO, Geneva, 2000.
- [30] C.E. Tan, S. Ma, D. Wai, S.K. Chew, E.S. Tai, Can we apply the National Cholesterol Education Program Adult Treatment Panel definition of the metabolic syndrome to Asians? *Diab. Care* 27 (2004) 1182–1186.
- [31] D.R. Matthews, J.P. Hosker, A.S. Rudenski, B.A. Naylor, D.F. Treacher, R.C. Turner, Homeostasis model assessment: insulin resistance and β -cell function from fasting plasma glucose and insulin concentration in man, *Diabetologia* 28 (1985) 412–419.
- [32] Y.F. Cheng, C.L. Chen, C.Y. Lai, T.Y. Chen, T.L. Huang, T.Y. Lee, et al., Assessment of donor fatty livers for liver transplantation, *Transplantation* 71 (2001) 1221–1225.
- [33] J.B. Meigs, The metabolic syndrome, *BMJ* 327 (2003) 61–62.
- [34] R. Kahn, J. Buse, E. Ferrannini, M. Stern, The metabolic syndrome: time for a critical appraisal: joint statement from the American Diabetes Association and the European Association for the Study of Diabetes, *Diab. Care* 28 (2005) 2289–2304.
- [35] S.H. Kim, G.M. Reaven, The metabolic syndrome: one step forward, two steps back, *Diab. Vasc. Dis. Res.* 1 (2004) 68–75.
- [36] M.P. Reilly, D.J. Rader, The metabolic syndrome: more than the sum of its parts? *Circulation* 108 (2003) 1546–1551.
- [37] S. Malik, N.D. Wong, S.S. Franklin, T.V. Kamath, G.J. L'Italian, J.R. Pio, et al., Impact of the metabolic syndrome on mortality from coronary heart disease, cardiovascular disease, and all causes in United States adults, *Circulation* 110 (2004) 1245–1250.
- [38] E.S. Ford, Risks for all-cause mortality, cardiovascular disease, and diabetes associated with the metabolic syndrome: a summary of the evidence, *Diab. Care* 28 (2005) 1769–1778.
- [39] N. Sattar, A. Gaw, O. Scherbakova, I. Ford, D.S. O'Reilly, S.M. Haffner, et al., Metabolic syndrome with and without C-reactive protein as a predictor of coronary heart disease and diabetes in the west of Scotland Coronary Prevention Study, *Circulation* 108 (2003) 141–149.
- [40] J.G. Fan, J. Zhu, X.J. Li, L. Chen, Y.S. Lu, L. Li, et al., Fatty liver and the metabolic syndrome among Shanghai adults, *J. Gastroenterol. Hepatol.* 20 (2005) 1825–1832.
- [41] B. Vozarova, N. Stefan, R.S. Lindsay, A. Saremi, R.E. Pratley, C. Bogardus, et al., High alanine aminotransferase is associated with decreased hepatic insulin sensitivity and predicts the development of type 2 diabetes, *Diabetes* 51 (2002) 1889–1895.
- [42] D.S. Lee, J.C. Evans, S.J. Robins, P.W. Wilson, I. Albano, C.S. Fox, et al., Gamma glutamyl transferase and metabolic syndrome, cardiovascular disease, and mortality risk: the Framingham Heart Study, *Arterioscler. Thromb. Vasc. Biol.* 27 (2007) 127–133.
- [43] S.M. Haffner, Relationship of metabolic risk factors and development of cardiovascular disease and diabetes, *Obesity* 14 (Suppl. 3) (2006) 121S–127S.
- [44] K.L. Cheal, F. Abbasi, C. Lamendola, T. McLaughlin, G.M. Reaven, E.S. Ford, Relationship to insulin resistance of the adult treatment panel III diagnostic criteria for identification of the metabolic syndrome, *Diabetes* 53 (2004) 1195–1200.
- [45] ToledoFG, A.D. Sniderman, D.E. Kelly, Influence of hepatic steatosis (fatty liver) on severity and composition of dyslipidemia in type 2 diabetes, *Diab. Care* 29 (2006) 1845–1850.
- [46] D.E. Kelly, T.M. McKolanis, R.A. Hegazi, L.H. Kuller, S.C. Kalhan, Fatty liver in type 2 diabetes mellitus: relation to regional adiposity, fatty acids and insulin resistance, *Am. J. Physiol. Endocrinol. Metab.* 285 (2003) E906–E916.
- [47] E. Bugianesi, U. Pagotto, R. Manini, E. Vanni, A. Gastaldelli, R. Iasio, et al., Plasma adiponectin in nonalcoholic fatty liver is related to hepatic insulin resistance and hepatic fat content, not to liver disease severity, *J. Clin. Endocrinol. Metab.* 90 (2005) 3498–3504.
- [48] M. Yoneda, T. Iwasaki, K. Fujita, H. Kirikoshi, M. Inamori, Y. Nozaki, et al., Hypoadiponectinemia plays a crucial role in the development of nonalcoholic fatty liver disease in patients with type 2 diabetes mellitus independent of visceral adipose tissue, *Alcohol. Clin. Exp. Res.* 31 (2007) 15S–21S.
- [49] V.W.-S. Wong, A.Y. Hui, S.W.-C. Tsang, J.L.-Y. Chan, A.M.-L. Tse, K.F. Chan, et al., Metabolic and adipokine profile of Chinese patients with nonalcoholic fatty liver disease, *Clin. Gastroenterol. Hepatol.* 4 (2006) 1154–1161.
- [50] N. Mendez-Sanchez, N.C. Chavez-Tapia, R. Medina-Santillan, A.R. Villa, K. Sanchez-Lara, G. Ponciano-Rodriguez, et al., The efficacy of adipokines and indices of metabolic syndrome as predictors of severe obesity-related hepatic steatosis, *Dig. Dis. Sci.* 51 (2006) 1716–1722.
- [51] V. Nobili, M. Manco, P. Ciampalini, V. Diciommo, R. Devito, F. Piemonte, et al., Leptin, free leptin index, insulin resistance and liver fibrosis in children with non-alcoholic fatty liver disease, *Eur. J. Endocrinol.* 155 (735) (2006) 743.
- [52] M. Inoue, M. Yano, M. Yamakado, E. Maehata, S. Suzuki, Relationship between the adiponectin-leptin ratio and parameters of insulin resistance in subjects without hyperglycemia, *Metabolism* 55 (2006) 124–1254.

- [53] R.V. Considine, M.K. Sinha, M.L. Heiman, A. Kriauciunas, T.W. Stephens, M.R. Nyce, et al., Serum immunoreactive-leptin concentrations in normal-weight and obese humans, *N. Engl. J. Med.* 334 (1996) 292–295.
- [54] C. Weyer, T. Funahashi, S. Tanaka, K. Hotta, Y. Matsuzawa, R.E. Pratley, et al., Hypoadiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia, *J. Clin. Endocrinol. Metab.* 86 (2001) 1930–1935.
- [55] F. Abbasi, J.W. Chu, C. Lamendola, T. McLaufhlin, J. Hayden, G.M. Reaven, et al., Discrimination between obesity and insulin resistance in the relationship with adiponectin, *Diabetes* 53 (2004) 585–590.
- [56] R.S. Lindsay, T. Funahashi, R.I. Hanson, Y. Matsuzawa, S. Tanaka, P.A. Tataranni, et al., Adiponectin and development of type 2 diabetes in the Pima Indian population, *Lancet* 360 (2002) 57–58.
- [57] J. Spranger, A. Kroke, M. Mohlig, M.M. Bergmann, M. Ristow, H. Boeing, et al., Adiponectin and protection against type 2 diabetes mellitus, *Lancet* 361 (2003) 226–228.
- [58] B. Vozarova, N. Stefan, R.S. Lindsay, J. Krakoff, W.C. Knowler, T. Funahashi, et al., Low plasma adiponectin concentrations do not predict weight gain in humans, *Diabetes* 51 (2002) 2964–2967.
- [59] M. Bajaj, S. Suraamornkul, P. Piper, L.J. Hardie, L. Glass, E. Cersosimo, et al., Decreased plasma adiponectin concentration are closely related to hepatic fat content and hepatic insulin resistance in pioglitazone-treated type 2 diabetes patients, *J. Clin. Endocrinol. Metab.* 89 (2004) 200–206.
- [60] P.M. Ridker, C.H. Hennekens, J.E. Buring, N. Rifai, C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women, *N. Engl. J. Med.* 342 (2000) 836–843.
- [61] W. Koenig, M. Sund, M. Frohlich, H.G. Fisher, H. Lowel, A. Doring, et al., C-reactive protein, a sensitive marker of inflammation, predicts future risk of coronary heart disease in initially healthy middle-aged men: results from the MONICA (Monitoring Trends and Determinants in Cardiovascular Disease) Augsburg Cohort Study, 1984 to 1992, *Circulation* 99 (1999) 237–242.
- [62] M.K. Rutter, J.B. Meigs, L.M. Sullivan, R.B. D'Agostino, P.W. Wilson, C-reactive protein, the metabolic syndrome, and prediction of cardiovascular events in the Framingham Offspring Study, *Circulation* 110 (2004) 380–385.