

Review Article

Sphingolipids in spinal cord injury

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Abstract: Spinal cord injury (SCI) is a debilitating condition that affects millions of individuals worldwide. Despite progress over the last few decades, the molecular mechanisms of secondary SCI that continue to occur days and weeks after the original trauma remain poorly understood. As a result, current therapies for SCI are only marginally effective. Sphingolipids, a diverse class of bioactive lipids, have been shown to regulate SCI repair and key secondary injury processes such as apoptosis, ischemia and inflammation. This review will discuss the numerous roles of sphingolipids and highlight the potential of sphingolipid-targeted therapies for SCI.

Keywords: Spinal cord injury, sphingolipid metabolism, ceramide, S1P, apoptosis, inflammation

Introduction

Spinal cord injury (SCI) is a devastating medical emergency that results from severe physical trauma to the spine. Damage to the spinal cord and surrounding cells begins immediately, and subsequent damage continues to occur days and even weeks later [1]. Accordingly, these two processes can be classified as either primary injury-cell death due to the original trauma or secondary injury-cell death due to inflammation, ischemia, activation of apoptosis pathways or other complex biological responses such as edema, excitotoxicity, free radical production or axon demyelination [2, 3]. An unfortunate consequence of these secondary processes is that they often perpetuate each other in a vicious cycle such that the traumatic injury is compounded and expanded beyond the initial lesion area. Due to the lack of effective therapies, the prognosis for patients with SCI is poor, and these individuals often live with significant physical, emotional, and financial burdens.

Of the secondary SCI mechanisms, inflammation is a major contributor to cell death and loss of neuronal function [4, 5]. The inflammatory response in SCI is marked by the release of inflammatory cytokines in or near the SCI site which then induce the activation and migration

of immune cells toward the lesion area [6]. The role of inflammation in SCI has long been debated, but the general consensus is that there are both harmful and beneficial aspects to inflammatory responses after SCI. Inflammation is a key process in the clearance of cytotoxic cell debris, but sustained activation of inflammatory responses ultimately leads to tissue damage and cell death [2, 3]. While the primary SCI is largely intractable, secondary mediators of injury such as inflammation present several targets that can be exploited for SCI treatment [7-10].

Named after the mythical Sphinx [11], sphingolipids are a class of bioactive lipids made up of long-chain sphingoid bases. The sphingolipids sphingosine, sphingosine-1-phosphate (S1P), ceramide and ceramide-1-phosphate (C1P) were thought to be merely structural components of cellular membranes, although in recent years they have come to be more fully appreciated for their roles in a variety of processes such as signal transduction [12], cell growth [13] and apoptosis [14]. In addition, ceramide is an essential precursor in the synthesis of complex sphingolipids such as sphingomyelin, cerebroside, sulfatides, globosides and gangliosides. The number of bioactive molecules resulting from sphingolipid metabolism is quite staggering, and so is the number of biological processes mediated by these molecules: cell migration

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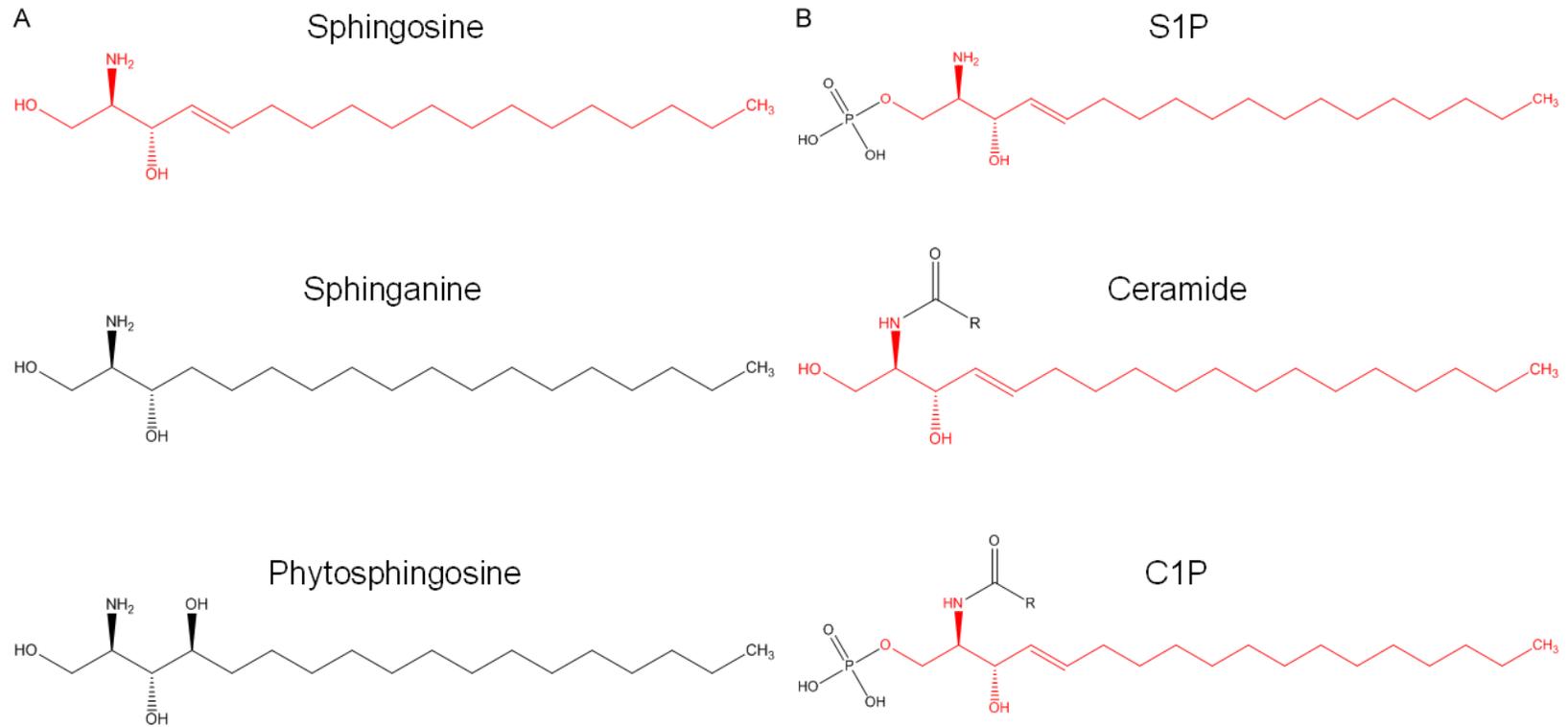


Figure 1. Structure of sphingoid bases and simple sphingolipids. A. The sphingoid bases sphingosine, sphinganine and phytosphingosine are long-chain acyclic aliphatic compounds. B. Sphingosine, shown in red, is the base for the other three simple sphingolipids: S1P, ceramide and C1P. Note the variable chain length of ceramide which adds to the complexity of sphingolipid metabolism.

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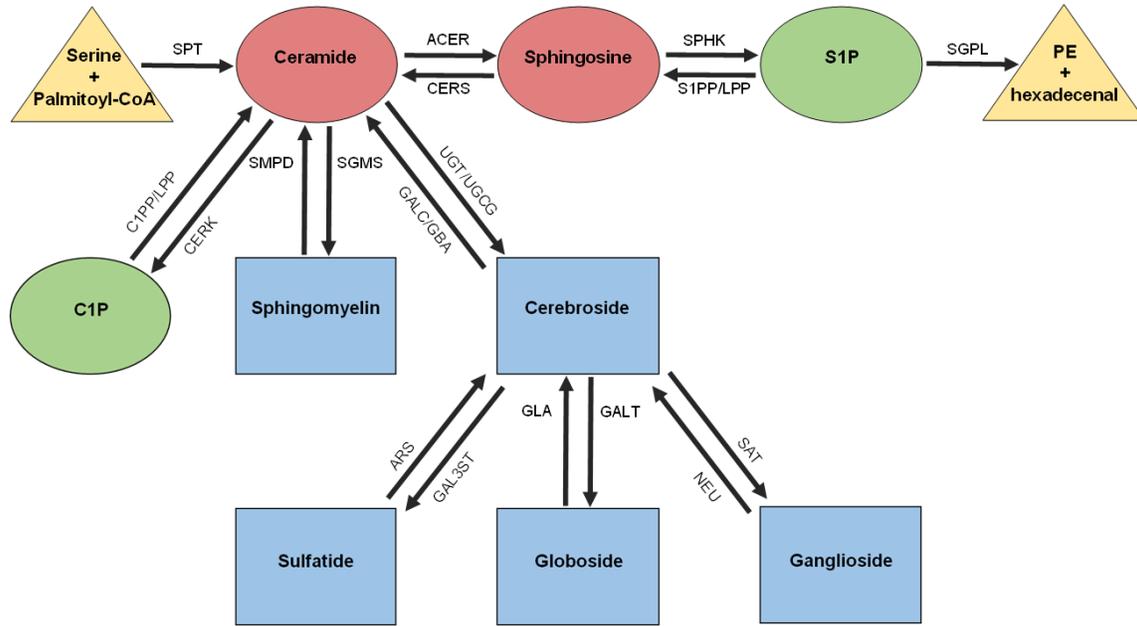


Figure 2. Spingolipid Metabolism. Once ceramide is synthesized *de novo* from serine and palmitoyl-CoA, it can undergo one of several reversible reactions. Ceramide can be phosphorylated by CERK to form C1P. Ceramide is a precursor to the complex sphingolipids including sphingomyelin, cerebroside, sulfatides, globosides and gangliosides. Breakdown of sphingolipids proceeds via hydrolysis of ceramide to sphingosine, phosphorylation by SPHK to form S1P, and lysis to PE and hexadecenal. Per the “sphingolipid rheostat” model, C1P and S1P (green) have pro-survival effects, whereas ceramide and sphingosine (red) have pro-apoptotic effects. Abbreviations: SPT = Serine Palmitoyltransferase, S1P = Sphingosine-1-phosphate, C1P = Ceramide-1-phosphate, ACER = Acid/Alkaline Ceramidase, CERS = Ceramide Synthase, SPHK = Sphingosine Kinase, S1PP = S1P Phosphatase, LPP = Lipid Phosphate Phosphatase, SGPL = S1P Lyase, PE = Phosphoethanolamine, C1PP = C1P Phosphatase, CERK = Ceramide Kinase, SMPD = Sphingomyelin Phosphodiesterase, SGMS = Sphingomyelin Synthase, GALC = Galactosylceramidase, GBA = Glucosidase Beta Acid, UGT = UDP Glycosyltransferase, UGCG = UDP-Glucose Ceramide Glucosyltransferase, ARS = Arylsulfatase, GAL3ST = Galactose-3-O-Sulfotransferase, GLA = Galactosidase, GALT = Galactosyltransferase, NEU = Sialidase, SAT = Sialyltransferase.

and proliferation [15, 16], differentiation [17], stress response [18], neuronal plasticity [19], angiogenesis [20] and immune function [21], among many others. This review is by no means exhaustive but rather endeavors to discuss the role of sphingolipid signaling in SCI processes and underscores the potential of sphingolipid metabolism as a therapeutic target for SCI.

Overview of sphingolipid metabolism

Simple sphingolipids

A discussion of sphingolipid metabolism should naturally begin with *de novo* synthesis of simple sphingolipids and sphingoid bases, the building blocks of sphingolipids (Figure 1). Sphingoid bases are generally described as long-chain acyclic aliphatic compounds and are synthesized *de novo* by serine palmitoyltransferase (SPT) from palmitoyl-CoA and serine or via ceramide catabolism [22]. The most common

sphingoid bases are sphingosine, sphinganine and phytosphingosine. Of these, sphingosine is often regarded as the most biologically relevant sphingoid base in mammals, since sphingosine and its phosphorylated form (S1P) are implicated in a variety of physiological and pathological processes [23]. In addition, sphingosine is reversibly convertible with another highly relevant sphingolipid: ceramide. Sphingolipid metabolism involves a series of such reversible reactions, with anabolic and catabolic processes working in parallel to regulate cellular levels of the various sphingolipids (Figure 2). A particularly important example of this involves the balance between kinases and phosphatases in this pathway. Sphingosine kinase (SPHK) and ceramide kinase (CERK) catalyze the phosphorylation of sphingosine and ceramide, respectively, while S1P phosphatase and C1P phosphatase—in addition to other lipid phosphate phosphatases (LPPs)—catalyze the dephosphorylation of S1P and C1P. Maintaining the appro-

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priate balance of these sphingolipids is vital to cell survival, as an excess of one or the other can have disastrous consequences. This relationship is often referred to as the “sphingolipid rheostat” model. First proposed in 1996 [24], the sphingolipid rheostat model posits that the levels of S1P and ceramide are key determinants of cell fate, with S1P promoting cell survival and ceramide promoting cell death [25].

Complex sphingolipids

Complex sphingolipids such as phosphosphingolipids and glycosphingolipids (GSLs) are predominantly structural components of plasma membranes [22], and their synthesis requires ceramide (**Figure 3**). Of note, mammals have six genes dedicated to the synthesis of ceramide and are appropriately named ceramide synthases (CerS). Ceramide is unique among sphingolipids in the sheer number of genes dedicated to its synthesis, suggesting that ceramides and the CerS serve vital functions. Through the action of sphingomyelin synthase (SMS), phosphocholine is added to ceramide to form sphingomyelin. Conversely, sphingomyelinases (SMases) catalyze the reverse reaction, generating ceramide. Sphingomyelin is a complex phosphosphingolipid and a major component of both myelin sheath and cell plasma membrane. In humans, the sphingomyelin content of CNS and PNS myelin is 7.9% and 17.7%, respectively (**Table 1**). Plasma membrane sphingomyelin content normally falls between 10-20% in humans and is highly variable by cell type, with Schwann cells, the PNS myelin-producing cells, reaching as high as 30% [27].

GSLs are formed by the addition of varying carbohydrate groups to ceramide. Cerebrosides, sulfatides, globosides and gangliosides constitute the four main classes of GSLs, and they have both overlapping and non-overlapping functions within cells [28]. Cerebrosides, as their name suggests, were first isolated from the brain [11], and are the most abundant class of GSLs found in nervous tissue. Cerebrosides consist of ceramide with an added glucose or galactose, yielding glucocerebroside (also known as glucosylceramide) and galactocerebroside (also known as galactosylceramide), respectively. The reverse reaction generates ceramide via the action of cerebrosidases. The diversity of ceramides coupled with the diversity of glycan modifications yields a remarkable

number of permutations for this class of lipids [28]. Cerebrosides can be sulfated (sulfatides), glycosylated (globosides) or sialyated (gangliosides) to generate bioactive GSLs with roles in numerous biological processes. The plasma membrane concentration of GSLs is relatively low and ranges by cell type under 10%, while the GSL content of CNS and PNS myelin in humans is much higher, at 27.5% and 22.1%, respectively (**Table 1**).

Despite the vast complexity of sphingolipid metabolism, all sphingolipids share a common synthesis and breakdown pathway through ceramide (**Figure 2**). Ceramide can be irreversibly synthesized *de novo* from serine and palmitoyl-CoA, or it can be generated by SMases, cerebrosidases, LPPs or CerS in the ceramide salvage pathway. Likewise, the common sphingolipid breakdown pathway involves catabolism to ceramide, conversion to sphingosine, phosphorylation to S1P and irreversible degradation by S1P lyase to form phosphoethanolamine and hexadecenal.

Sphingolipids and SCI

While the biochemical changes involved in SCI are not completely understood, recent studies suggest that sphingolipids may play a prominent role [29-31]. The simple sphingolipids ceramide, C1P, sphingosine and S1P have been shown to mediate several aspects of SCI pathogenesis. Nearly three decades ago, researchers demonstrated that exogenous ceramide promotes survival or death of spinal motor neurons by regulating apoptosis in a dose-dependent manner [32]. This key role of ceramide was further elucidated in subsequent work which showed that inhibition of ceramide biosynthesis via CerS and SMase inhibitors significantly improved motor function and reduced the amount of tissue injury, neutrophil infiltration, apoptosis and cytokine production in a mouse model of SCI [33]. C1P has also been implicated in spinal neuronal death via the activation of cytosolic phospholipase A2 (cPLA2)-a key enzyme in the production of various inflammatory lipid mediators. Further, genetic deletion of cPLA2 or pharmacological inhibition at just 30 minutes post-injury substantially reversed these effects in mice, improving motor function and reducing tissue damage after SCI [30, 34]. Several studies in rodent models have shown that administration of FTY720, a sphin-

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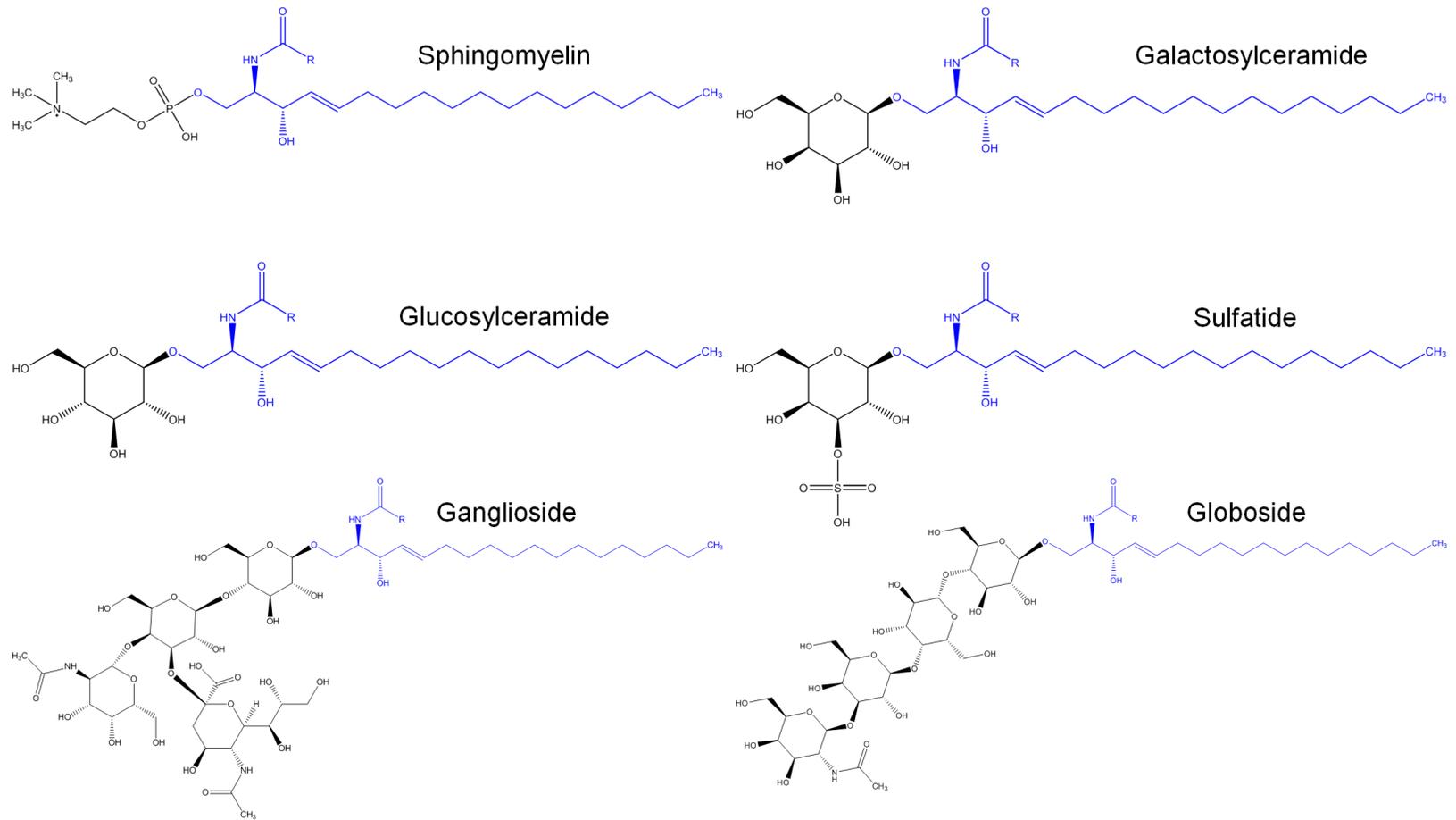


Figure 3. Structure of complex sphingolipids. Phosphosphingolipids and glycosphingolipids are synthesized via modifications to ceramide, shown in blue. Addition of phosphocholine to ceramide yields sphingomyelin. Addition of glucose or galactose to ceramide yields the cerebroside glucosylceramide and galactosylceramide, respectively. These cerebroside can be further glycosylated (globoside), sulfated (sulfatide) or sialyated (ganglioside).

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Table 1. Myelin composition [26]

Component	Human	Rat CNS	Human	Rat PNS
	CNS Myelin	Myelin	PNS Myelin	Myelin
Protein	30.0	29.5	28.7	-
Lipid	70.0	70.5	71.3	-
Cholesterol	27.7	27.3	23.0	27.2
Total Galactolipid	27.5	31.5	22.1	21.5
Cerebroside	22.7	23.7	-	15.8
Sulfatide	3.8	7.1	-	5.7
Sphingomyelin	7.9	3.2	17.7	7.0

gossine analog, promotes functional recovery after SCI [35-37] and reduces trauma-induced neuropathic pain via spinal S1P receptors (S1PRs) [38]. S1P was found to be elevated at the SCI site and enhanced the viability, migration and differentiation of neural progenitor cells [39]. This increased S1P at the injury site is posited to act as a chemoattractant for microglia and macrophages that is intended to be protective but inevitably becomes destructive.

The complex sphingolipids sphingomyelin and GM1 ganglioside are linked to SCI as well. A recent study used shiverer (myelin deficient) mice to assess axon regeneration following SCI. While *in vitro* shiverer neurons displayed neurite outgrowth comparable to wildtype neurons, *in vivo* shiverer fibers had an increased regenerative capacity. In this SCI model, myelin lipids—specifically cholesterol and sphingomyelin—were highly inhibitory for neurite outgrowth, and treatment with 2-hydroxypropyl- β -cyclodextrin, a drug that reduces the levels of these lipids, increased regeneration of wildtype axons following SCI [31]. GM1 ganglioside has been studied for decades (albeit with some debate [40]) as a therapeutic for SCI and is reported to have anti-neurotoxic, anti-inflammatory and neuroprotective effects that result in limited neurological improvement [41-44]. Nevertheless, this drug is not available for widespread clinical use.

In addition to these direct effects, simple and complex sphingolipids are well known mediators of secondary SCI mechanisms, namely apoptosis, ischemia and inflammation. What follows is a discussion of sphingolipids in each of these processes.

Apoptosis and cell survival

Apoptosis of neurons and oligodendrocytes (the CNS myelin-producing cells) in the injured

spinal cord can be observed within a few hours of the traumatic event [45, 46]. As time goes on, expansion of the lesion area and Wallerian degeneration take effect, exacerbating the deleterious effects of the initial injury [47, 48]. According to the sphingolipid rheostat model, the dynamic balance between ceramide and S1P largely determines cellular fates [24]. More broadly, a mass of evidence suggests that sphingosine and C1P can be included in this model as promoters of apoptosis and cell survival, respectively. Whether directly or indirectly, a variety of cellular events alter the levels of ceramide and sphingosine to promote apoptosis, just as a variety of events alter the levels of S1P and C1P to promote cell survival [49, 50]. Complex sphingolipids such as sphingomyelin [51] and gangliosides [52, 53] have also been linked to apoptosis in diverse cell types.

Ceramide and sphingosine: Sphingosine and the FTY720 analog have been shown to induce apoptosis in a variety of cell types [54-57], and ceramide-induced apoptosis has been an intensely studied phenomena since its discovery in the early 1990s [14, 58, 59]. Numerous apoptotic stimuli can activate acid SMase and neutral SMase to generate ceramide [60, 61], while SMS can suppress ceramide-induced apoptosis [62]. CerS and SPT are also activated during apoptosis in response to various stimuli [63-65]. Of note, recent work has shown that ceramides are capable of forming protein-permeable mitochondrial outer membrane channels, and that this process is inhibited *in vitro* and *in vivo* by B-cell lymphoma (Bcl) extra-large [66, 67]. This finding has meaningful implications for our understanding of the regulation of apoptosis by ceramide and provides intriguing new insights into the process of apoptosis.

S1P and C1P: S1P promotes cell survival and proliferation in a myriad of ways, either via the action of S1P transporters [68], S1PRs [69] or LPPs [70]. S1P stimulates a number of secondary messengers including nitric oxide synthase, phosphoinositide 3-kinase (PI3K), mitogen-activated protein kinase (MAPK) and protein kinase B and suppresses c-Jun N-terminal kinase and Bcl-2 associated X protein to enhance survival [71-73]. Likewise, C1P functions through PI3K, protein kinase B, protein kinase C, c-Jun N-terminal kinase, MAPK, nitric oxide synthase and mechanistic target of rapamycin

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signaling to promote cell survival [74-78] and is a demonstrated mitogen [79]. C1P-mediated inhibition of acid SMase and SPT has been implicated in regulating survival as well [80, 81]. CERK is a regulator of cell growth and survival [82], though, intriguingly, there is conflicting evidence on the role of SPHK in apoptosis. SPHK activation has been known to inhibit apoptosis for decades [83] and has been linked to cell survival [84-86]. However, SPHK overexpression has also been shown to suppress growth and enhance apoptosis [87, 88]. In addition, inhibition of SPHKs has paradoxically been shown to both promote [89, 90] and inhibit apoptosis [91]. While these contradictory roles are largely attributed to divergent functions of the isoenzymes SPHK1 and SPHK2 [92], this explanation is insufficient to describe the full range of observed phenomena. Consequently, crosstalk between these pathways or yet unknown functions of these kinases may play a role in regulating apoptosis/cell survival.

Ischemia

Loss of blood flow in SCI is a major contributor to SCI pathogenesis [2, 3] and has several connections to sphingolipid signaling and metabolism. Indeed, the balance of ceramide and S1P seems play a significant role in angiogenesis, with ceramide acting as an inhibitor [93, 94] and S1P (and thus SPHK) acting as an activator for this process [95-98]. Ischemic events can lead to increased production of ceramide via SMase upregulation or UDP-glucose ceramide glucosyltransferase and SMS downregulation [99-102], and neutral SMase inhibition prevents neuron death caused by ischemic stress [103]. Unexpectedly, exogenous ceramide has also been found to inhibit apoptosis and reduce the infarct size in focal cerebral ischemia via Bcl-2 upregulation [104]. This dual role of ceramide may be a result of crosstalk between pro-apoptotic and anti-apoptotic pathways or dose-dependency, that is, varying concentrations of ceramide may have differential effects. S1P, FTY720 and SPHKs exhibit protective effects during ischemic events [105-109]. S1P promotes functional recovery in the infarcted brain by enhancing neural progenitor cell migration. In line with this, the concentration of S1P in the brain was increased after ischemia, and inhibition of S1PRs enhanced S1P-mediated neural progenitor cell migration toward the injury site [110]. This is analogous to the chemoat-

tractant effect of S1P in SCI [39] and may represent a generalized mechanism for S1P in CNS injuries.

Endothelial cells: It is well established that endothelial cells (ECs) and vascular endothelial growth factor (VEGF) regulate the process of angiogenesis in numerous ways [111, 112], and this holds true in SCI as well [113-115]. There is mounting evidence that sphingolipids can interact with ECs and VEGF to regulate angiogenesis and vasculogenesis [116, 117]. S1P has long been implicated in EC function, and has been shown to stimulate EC migration [118, 119], increase barrier integrity [120-122] and enhance EC differentiation [123], proliferation [124], survival [125], adhesion [126] and VEGF expression [127]. Similarly, SPHKs have diverse functions in these processes [128-132]. Akin to their roles in apoptosis, ceramide and S1P regulate EC function in an antagonistic fashion, as ceramide has been shown to decrease barrier integrity and induce senescence in ECs [133, 134]. Cerebrosides and gangliosides have proangiogenic functions via VEGF [135-137], although ganglioside GM3 is able to suppress these effects, suggesting a more complex and nuanced relationship [138]. It is important to note that sphingolipids have not been shown to directly affect barrier integrity in a SCI model. Nonetheless, these results underscore the distinct possibility that sphingolipids contribute to SCI pathology by altering vascular permeability.

Inflammation

Inflammation in the injured spinal cord is a highly pathological process that begins shortly after the primary injury event. Despite, or perhaps owing to the diversity of inflammatory responses in SCI, immunotherapy has enjoyed only modest success in patients with SCI [7-10]. Cytokines and eicosanoids play central roles in the activation, differentiation, function and migration of immune cells, and sphingolipids are able to regulate these inflammatory mediators in diverse and complex ways to promote or inhibit inflammation. The function of sphingolipids in regulating immune cells has been the topic of numerous research papers and literature reviews in recent years [139-144].

Cytokines and eicosanoids: SCI pathogenesis is highly associated with cytokine dysregulation [2, 3], and SCI can induce the expression of cytokines in a matter of hours [145]. Sphingo-

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lipids are critical mediators of cytokine signaling [146, 147], though their effects are complex and often cell-type specific. S1P has been shown to enhance expression of IFN- γ and IL-2 [148], IL-27 [149], IL-17 [150] and IL-8 [151] and reduces expression of IL-12 and IL-23 [149]. Inhibition of SPHK1 reduces IL-17, TNF- α and IL-1 β production in activated microglia [144, 150], and SPHK1 interacts with the TNF α receptor via scaffolding protein TRAF2 [152]. SPHK2 associates with IL-12 receptors to modulate IL-12 signaling [153]. Exogenous C1P decreases secretion of TNF- α , IL-6, IL-8 and IL-1 β in peripheral blood mononuclear cells [154]. C1P and S1P stimulate the production of prostaglandin E2 [155] at least in part by activating cPLA2 [156, 157]. Sphingolipid phosphatases play a role, as TNF α induced transcription of IL-1 β was significantly reduced by S1P phosphatase siRNAs [158], and LPP regulates NF- κ B activation and IL-8 secretion [159]. Neutral SMase activity induces the production of TNF α , IL-1 β and IL-6 in astrocytes [160]. Sulfatide increases and cerebroside decreases the production of cytokines IL-1 β , IL-6, IL-8, TNF- α and CCL3 [161].

Macrophages: A number of studies have shown that the inflammatory response in SCI is mediated by the activation and invasion of bone marrow derived-macrophages at the site of injury [162-166], and these macrophages can assume either a pro-inflammatory or anti-inflammatory phenotype. Exposure of macrophages to myelin debris, as in SCI, has been shown to promote a pro-inflammatory phenotype [167, 168]. Myelin has a characteristically high sphingolipid content (**Table 1**), though it remains to be seen whether its effect on macrophages can be attributed to sphingolipids specifically. S1P in particular has diverse roles in mediating macrophage function and phenotype. Intracellular S1P, generated via SPHK, induces a pro-inflammatory macrophage phenotype, while extracellular S1P binding to S1PRs induces an anti-inflammatory phenotype, inhibiting NF- κ B activation and the production of pro-inflammatory cytokines while promoting the production of anti-inflammatory molecules [169]. Macrophages are protected from apoptosis via S1P-mediated inhibition of SMase [170], upregulation of anti-apoptotic Bcl-2 and Bcl extra-large [171] or activation of PI3K/MAPK/Ca²⁺ signaling [172]. S1P can act as a chemoattractant for monocyte and macro-

phage trafficking [173-175] and alter cytokine production in human macrophages [171, 176]. Conversely, the sphingosine analog FTY720 reduces macrophage infiltration *in vivo* [177-179]. SPHK1 mediates a variety of inflammatory responses in macrophages such as migration, NF- κ B activation and secretion of cytokines [180, 181], and inhibition of SPHK sensitizes macrophages to lipopolysaccharide-induced cell death [182, 183]. C1P also stimulates macrophage NF- κ B activation and chemokine CCL2 release to promote cell migration [184].

Glia: CNS inflammation, e.g. from a SCI, induces the migration and activation of microglia and astrocytes [2, 3]. These glial populations initially have constructive effects in response to injury, but prolonged activation contributes to further inflammation and tissue damage [185]. Astroglia is a common feature of CNS inflammation and is characterized by astrocyte proliferation and increased glial fibrillary acidic protein expression. Uncontrolled astroglia results in the formation of a glial scar surrounding the injury site which inhibits neural regeneration and functional recovery after SCI [186]. Studies in the 1980s and 1990s provided early evidence of sphingolipid-mediated glial activation by demonstrating that gangliosides stimulate glial cell proliferation and differentiation [187-189], and other known activators of glia include ceramide [160, 190-192] and sulfatide [193]. FTY720 has been shown to have diverse effects on glia: reducing reactive astroglia [29], altering calcium homeostasis [194], inhibiting vesicle mobility and secretion [195], decreasing NO production [144, 196], promoting migration [197], downregulating pro-inflammatory cytokines production [144, 198] and upregulating production of brain-derived neurotrophic factor and glial cell-derived neurotrophic factor [198]. Together, these results highlight a neuroprotective role for FTY720 through regulation of glial function.

Clinical applications

Components of the sphingolipid metabolic pathway have been targeted in various clinical trials, although only GM1 ganglioside has specifically been tested in patients with SCI [41-44]. As seen throughout this review, animal studies have uncovered a wealth of information regarding SCI pathology and treatment app-

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roaches. Ceramide biosynthesis inhibitors and FTY720 have been shown to attenuate nervous system insults in various rodent models [33, 36, 38, 103, 109, 177, 199], thus targeting the sphingolipid rheostat in future SCI studies is warranted and could mitigate tissue damage, alleviate pain and promote functional recovery in patients.

Despite early promise, GM1 ganglioside therapy development languished for decades due to criticisms of experimental design or failure to achieve defined endpoints in clinical trials [43, 44]. In recent years, however, new studies using GM1 ganglioside alone or with methylprednisolone-another controversial treatment for SCI [200]-have yielded positive results [41, 42].

FTY720, also known as fingolimod or Gilenya®, is the first oral drug approved by the FDA to treat relapsing multiple sclerosis [201, 202]. Since multiple sclerosis is a demyelinating disease that affects spinal neurons, these findings can be extended toward SCI therapies. For this reason, in addition to all of the previously described actions of FTY720, fingolimod may be a promising therapeutic for SCI.

Problematic methods for quantifying sphingolipids have impeded the development of sphingolipid biomarkers for human diseases, although recent progress has been made through advances in genomics and proteomics. Researchers are actively evaluating the utility of sphingolipid biomarkers in a variety of diseases such as cancer, diabetes, liver disease, acute brain injury and Alzheimer's disease [203-211]. Even so, predictive and prognostic biomarkers for SCI remain to be discovered.

Conclusion and outlook

The diversity of sphingolipids and the complexity of their metabolism are reflected in the diverse and complex ways by which they affect cell physiology and pathophysiology. Far beyond the well-known functions of ceramide and S1P in the sphingolipid rheostat model, simple and complex sphingolipids regulate the processes of apoptosis, cell survival, ischemia, angiogenesis, inflammation and SCI repair. Despite the wealth of evidence that suggests sphingolipids are involved in the pathogenic processes of SCI, there is a paucity of clinical research in this field. Can we quantify sphingolipid dysregula-

tion to develop useful predictive and prognostic biomarkers for SCI? Can we improve SCI outcomes by using our knowledge of sphingolipid metabolism to shift the balance toward pro-survival S1P and away from apoptotic ceramide? SCI is a tragic and disabling condition with no existing cure, and current therapies have only a modest effect. Tackling these important questions may prove to be a critical step forward in treating SCI and improving the lives of millions of people around the world.

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Disclosure of conflict of interest

None.

Abbreviations

Bcl, B-cell lymphoma; C1P, ceramide-1-phosphate; CERK, ceramide kinase; CerS, ceramide synthase; cPLA2, cytosolic phospholipase A2; EC, endothelial cell; GSL, glycosphingolipid; LPP, lipid phosphate phosphatase; MAPK, mitogen-activated protein kinase; PI3K, phosphoinositide 3-kinase; S1P, sphingosine-1-phosphate; S1PR, S1P receptor; SCI, spinal cord injury; SMase, sphingomyelinase; SMS, sphingomyelin synthase; SPHK, sphingosine kinase; SPT, serine palmitoyltransferase; VEGF, vascular endothelial growth factor.

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References

- [1] Mortazavi MM, Verma K, Harmon OA, Griessenauer CJ, Adeeb N, Theodore N and Tubbs RS. The microanatomy of spinal cord injury: a review. *Clin Anat* 2015; 28: 27-36.
- [2] Oyinbo CA. Secondary injury mechanisms in traumatic spinal cord injury: a nugget of this multiply cascade. *Acta Neurobiol Exp (Wars)* 2011; 71: 281-299.

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- [3] Sekhon LH and Fehlings MG. Epidemiology, demographics, and pathophysiology of acute spinal cord injury. *Spine (Phila Pa 1976)* 2001; 26: S2-12.
- [4] Anwar MA, Al Shehabi TS and Eid AH. Inflammogenesis of Secondary Spinal Cord Injury. *Front Cell Neurosci* 2016; 10: 98.
- [5] Allison DJ and Ditor DS. Immune dysfunction and chronic inflammation following spinal cord injury. *Spinal Cord* 2015; 53: 14-18.
- [6] Carlson SL, Parrish ME, Springer JE, Doty K and Dossett L. Acute inflammatory response in spinal cord following impact injury. *Exp Neurol* 1998; 151: 77-88.
- [7] Wang YT, Lu XM, Chen KT, Shu YH and Qiu CH. Immunotherapy strategies for spinal cord injury. *Curr Pharm Biotechnol* 2015; 16: 492-505.
- [8] Cox A, Varma A and Banik N. Recent advances in the pharmacologic treatment of spinal cord injury. *Metab Brain Dis* 2015; 30: 473-482.
- [9] Plemel JR, Wee Yong V and Stirling DP. Immune modulatory therapies for spinal cord injury-past, present and future. *Exp Neurol* 2014; 258: 91-104.
- [10] Singh PL, Agarwal N, Barrese JC and Heary RF. Current therapeutic strategies for inflammation following traumatic spinal cord injury. *Neural Regen Res* 2012; 7: 1812-1821.
- [11] Thudichum JL. A Treatise on the Chemical Constitution of the Brain. Archon Books 1962.
- [12] Hannun YA and Obeid LM. Principles of bioactive lipid signalling: lessons from sphingolipids. *Nat Rev Mol Cell Biol* 2008; 9: 139-150.
- [13] Spiegel S and Merrill AH Jr. Sphingolipid metabolism and cell growth regulation. *FASEB J* 1996; 10: 1388-1397.
- [14] Obeid LM, Linardic CM, Karolak LA and Hannun YA. Programmed cell death induced by ceramide. *Science* 1993; 259: 1769-1771.
- [15] Rivera IG, Ordonez M, Presa N, Gangoiti P, Gomez-Larrauri A, Trueba M, Fox T, Kester M and Gomez-Munoz A. Ceramide 1-phosphate regulates cell migration and invasion of human pancreatic cancer cells. *Biochem Pharmacol* 2016; 102: 107-119.
- [16] Zhang H, Desai NN, Olivera A, Seki T, Brooker G and Spiegel S. Sphingosine-1-phosphate, a novel lipid, involved in cellular proliferation. *J Cell Biol* 1991; 114: 155-167.
- [17] Bieberich E. Ceramide and sphingosine-1-phosphate signaling in embryonic stem cell differentiation. *Methods Mol Biol* 2012; 874: 177-192.
- [18] Hannun YA and Luberto C. Ceramide in the eukaryotic stress response. *Trends Cell Biol* 2000; 10: 73-80.
- [19] Wheeler D, Knapp E, Bandaru VV, Wang Y, Knorr D, Poirier C, Mattson MP, Geiger JD and Haughey NJ. Tumor necrosis factor-alpha-induced neutral sphingomyelinase-2 modulates synaptic plasticity by controlling the membrane insertion of NMDA receptors. *J Neurochem* 2009; 109: 1237-1249.
- [20] Lee OH, Kim YM, Lee YM, Moon EJ, Lee DJ, Kim JH, Kim KW and Kwon YG. Sphingosine 1-phosphate induces angiogenesis: its angiogenic action and signaling mechanism in human umbilical vein endothelial cells. *Biochem Biophys Res Commun* 1999; 264: 743-750.
- [21] Spiegel S and Milstien S. The outs and the ins of sphingosine-1-phosphate in immunity. *Nat Rev Immunol* 2011; 11: 403-415.
- [22] Gault CR, Obeid LM and Hannun YA. An overview of sphingolipid metabolism: from synthesis to breakdown. *Adv Exp Med Biol* 2010; 688: 1-23.
- [23] Pyne S, Adams DR and Pyne NJ. Sphingosine 1-phosphate and sphingosine kinases in health and disease: Recent advances. *Prog Lipid Res* 2016; 62: 93-106.
- [24] Cuvillier O, Pirianov G, Kleuser B, Vanek PG, Coso OA, Gutkind S and Spiegel S. Suppression of ceramide-mediated programmed cell death by sphingosine-1-phosphate. *Nature* 1996; 381: 800-803.
- [25] Newton J, Lima S, Maceyka M and Spiegel S. Revisiting the sphingolipid rheostat: Evolving concepts in cancer therapy. *Exp Cell Res* 2015; 333: 195-200.
- [26] Slikker W Jr and Chang LW. Handbook of developmental neurotoxicology. Academic Press, 1998.
- [27] Calderon RO and DeVries GH. Lipid composition and phospholipid asymmetry of membranes from a Schwann cell line. *J Neurosci Res* 1997; 49: 372-380.
- [28] D'Angelo G, Capasso S, Sticco L and Russo D. Glycosphingolipids: synthesis and functions. *FEBS J* 2013; 280: 6338-6353.
- [29] Wang J, Wang J, Lu P, Cai Y, Wang Y, Hong L, Ren H, Heng BC, Liu H, Zhou J and Ouyang H. Local delivery of FTY720 in PCL membrane improves SCI functional recovery by reducing reactive astrogliosis. *Biomaterials* 2015; 62: 76-87.
- [30] Liu NK, Deng LX, Zhang YP, Lu QB, Wang XF, Hu JG, Oakes E, Bonventre JV, Shields CB and Xu XM. Cytosolic phospholipase A2 protein as a novel therapeutic target for spinal cord injury. *Ann Neurol* 2014; 75: 644-658.
- [31] Mar FM, da Silva TF, Morgado MM, Rodrigues LG, Rodrigues D, Pereira MI, Marques A, Sousa VF, Coentro J, Sa-Miranda C, Sousa MM and Brites P. Myelin Lipids Inhibit Axon Regeneration Following Spinal Cord Injury: a Novel Perspective for Therapy. *Mol Neurobiol* 2016; 53: 1052-1064.
- [32] Irie F and Hirabayashi Y. Application of exogenous ceramide to cultured rat spinal motoneurons promotes survival or death by regulation

Sphingolipids in SCI

- of apoptosis depending on its concentrations. *J Neurosci Res* 1998; 54: 475-485.
- [33] Cuzzocrea S, Deigner HP, Genovese T, Mazzon E, Esposito E, Crisafulli C, Di Paola R, Bramanti P, Matuschak G and Salvemini D. Inhibition of ceramide biosynthesis ameliorates pathological consequences of spinal cord injury. *Shock* 2009; 31: 634-644.
- [34] Liu NK, Byers JS, Lam T, Lu QB, Sengelaub DR and Xu XM. Inhibition of cPLA2 has neuroprotective effects on motoneuron and muscle atrophy following spinal cord injury. *J Neurotrauma* 2014; [Epub ahead of print].
- [35] Norimatsu Y, Ohmori T, Kimura A, Madoiwa S, Mimuro J, Seichi A, Yatomi Y, Hoshino Y and Sakata Y. FTY720 improves functional recovery after spinal cord injury by primarily nonimmunomodulatory mechanisms. *Am J Pathol* 2012; 180: 1625-1635.
- [36] Lee KD, Chow WN, Sato-Bigbee C, Graf MR, Graham RS, Colello RJ, Young HF and Mathern BE. FTY720 reduces inflammation and promotes functional recovery after spinal cord injury. *J Neurotrauma* 2009; 26: 2335-2344.
- [37] Zhang J, Zhang A, Sun Y, Cao X and Zhang N. Treatment with immunosuppressants FTY720 and tacrolimus promotes functional recovery after spinal cord injury in rats. *Tohoku J Exp Med* 2009; 219: 295-302.
- [38] Zhang DD, Linke B, Suo J, Zivkovic A, Schreiber Y, Ferreiros N, Henke M, Geisslinger G, Stark H and Scholich K. Antinociceptive effects of FTY720 during trauma-induced neuropathic pain are mediated by spinal S1P receptors. *Biol Chem* 2015; 396: 783-794.
- [39] Kimura A, Ohmori T, Ohkawa R, Madoiwa S, Mimuro J, Murakami T, Kobayashi E, Hoshino Y, Yatomi Y and Sakata Y. Essential roles of sphingosine 1-phosphate/S1P1 receptor axis in the migration of neural stem cells toward a site of spinal cord injury. *Stem Cells* 2007; 25: 115-124.
- [40] Hurlbert RJ, Hadley MN, Walters BC, Aarabi B, Dhall SS, Gelb DE, Rozzelle CJ, Ryken TC and Theodore N. Pharmacological therapy for acute spinal cord injury. *Neurosurgery* 2013; 72 Suppl 2: 93-105.
- [41] Barros TE Jr, Araujo FF, Higino LD, Marcon RM and Cristante AF. The Effect of Monosialoganglioside (Gm-1) Administration in Spinal Cord Injury. *Acta Ortop Bras* 2016; 24: 123-126.
- [42] Xu D, Yang L, Li Y and Sun Y. Clinical study of ganglioside (GM) combined with methylprednisolone (MP) for early acute spinal injury. *Pak J Pharm Sci* 2015; 28: 701-704.
- [43] Geisler FH, Coleman WP, Grieco G, Poonian D and Sygen Study G. The Sygen multicenter acute spinal cord injury study. *Spine (Phila Pa 1976)* 2001; 26: S87-98.
- [44] Geisler FH, Dorsey FC and Coleman WP. Recovery of motor function after spinal-cord injury—a randomized, placebo-controlled trial with GM-1 ganglioside. *N Engl J Med* 1991; 324: 1829-1838.
- [45] Liu XZ, Xu XM, Hu R, Du C, Zhang SX, McDonald JW, Dong HX, Wu YJ, Fan GS, Jacquin MF, Hsu CY and Choi DW. Neuronal and glial apoptosis after traumatic spinal cord injury. *J Neurosci* 1997; 17: 5395-5406.
- [46] Almad A, Sahinkaya FR and McTigue DM. Oligodendrocyte fate after spinal cord injury. *Neurotherapeutics* 2011; 8: 262-273.
- [47] Abe Y, Yamamoto T, Sugiyama Y, Watanabe T, Saito N, Kayama H and Kumagai T. Apoptotic cells associated with Wallerian degeneration after experimental spinal cord injury: a possible mechanism of oligodendroglial death. *J Neurotrauma* 1999; 16: 945-952.
- [48] Crowe MJ, Bresnahan JC, Shuman SL, Masters JN and Beattie MS. Apoptosis and delayed degeneration after spinal cord injury in rats and monkeys. *Nat Med* 1997; 3: 73-76.
- [49] Van Brocklyn JR and Williams JB. The control of the balance between ceramide and sphingosine-1-phosphate by sphingosine kinase: oxidative stress and the seesaw of cell survival and death. *Comp Biochem Physiol B Biochem Mol Biol* 2012; 163: 26-36.
- [50] Mullen TD and Obeid LM. Ceramide and apoptosis: exploring the enigmatic connections between sphingolipid metabolism and programmed cell death. *Anticancer Agents Med Chem* 2012; 12: 340-363.
- [51] Leucht K, Fischbeck A, Caj M, Liebisch G, Hartlieb E, Benes P, Fried M, Humpf HU, Rogler G and Hausmann M. Sphingomyelin and phosphatidylcholine contrarily affect the induction of apoptosis in intestinal epithelial cells. *Mol Nutr Food Res* 2014; 58: 782-798.
- [52] Ha SH, Lee JM, Kwon KM, Kwak CH, Abekura F, Park JY, Cho SH, Lee K, Chang YC, Lee YC, Choi HJ, Chung TW, Ha KT, Chang HW and Kim CH. Exogenous and Endogenous Disialosyl Ganglioside GD1b Induces Apoptosis of MCF-7 Human Breast Cancer Cells. *Int J Mol Sci* 2016; 17.
- [53] Omran OM, Saqr HE and Yates AJ. Endogenous GD3 ganglioside induces apoptosis in U-1242 MG glioma cells. *Int J Health Sci (Qassim)* 2011; 5: 4-6.
- [54] Chen L, Luo LF, Lu J, Li L, Liu YF, Wang J, Liu H, Song H, Jiang H, Chen SJ, Luo C and Li KK. FTY720 induces apoptosis of M2 subtype acute myeloid leukemia cells by targeting sphingolipid metabolism and increasing endogenous ceramide levels. *PLoS One* 2014; 9: e103033.
- [55] Kanno T, Nishimoto T, Fujita Y, Gotoh A, Nakano T and Nishizaki T. Sphingosine induces apopto-

Sphingolipids in SCI

- sis in MKN-28 human gastric cancer cells in an SDK-dependent manner. *Cell Physiol Biochem* 2012; 30: 987-994.
- [56] Kanno T and Nishizaki T. Sphingosine induces apoptosis in hippocampal neurons and astrocytes by activating caspase-3/-9 via a mitochondrial pathway linked to SDK/14-3-3 protein/Bax/cytochrome c. *J Cell Physiol* 2011; 226: 2329-2337.
- [57] Lee TK, Man K, Ho JW, Sun CK, Ng KT, Wang XH, Wong YC, Ng IO, Xu R and Fan ST. FTY720 induces apoptosis of human hepatoma cell lines through PI3-K-mediated Akt dephosphorylation. *Carcinogenesis* 2004; 25: 2397-2405.
- [58] Toman RE, Movsesyan V, Murthy SK, Milstien S, Spiegel S and Faden AI. Ceramide-induced cell death in primary neuronal cultures: upregulation of ceramide levels during neuronal apoptosis. *J Neurosci Res* 2002; 68: 323-330.
- [59] Park JY, Kim MJ, Kim YK and Woo JS. Ceramide induces apoptosis via caspase-dependent and caspase-independent pathways in mesenchymal stem cells derived from human adipose tissue. *Arch Toxicol* 2011; 85: 1057-1065.
- [60] Wang SW, Hojabrpour P, Zhang P, Kolesnick RN, Steinbrecher UP, Gomez-Munoz A and Duronio V. Regulation of ceramide generation during macrophage apoptosis by ASMase and de novo synthesis. *Biochim Biophys Acta* 2015; 1851: 1482-1489.
- [61] Yabu T, Shiba H, Shibasaki Y, Nakanishi T, Imamura S, Touhata K and Yamashita M. Stress-induced ceramide generation and apoptosis via the phosphorylation and activation of nSMase1 by JNK signaling. *Cell Death Differ* 2015; 22: 258-273.
- [62] Tafesse FG, Vacaru AM, Bosma EF, Hermansson M, Jain A, Hilderink A, Somerharju P and Holthuis JC. Sphingomyelin synthase-related protein SMSr is a suppressor of ceramide-induced mitochondrial apoptosis. *J Cell Sci* 2014; 127: 445-454.
- [63] Mojakgomo R, Mbita Z and Dlamini Z. Linking the ceramide synthases (CerSs) 4 and 5 with apoptosis, endometrial and colon cancers. *Exp Mol Pathol* 2015; 98: 585-592.
- [64] Dai L, Trillo-Tinoco J, Bai A, Chen Y, Bielawski J, Del Valle L, Smith CD, Ochoa AC, Qin Z and Parsons C. Ceramides promote apoptosis for virus-infected lymphoma cells through induction of ceramide synthases and viral lytic gene expression. *Oncotarget* 2015; 6: 24246-24260.
- [65] Veret J, Coant N, Berdyshev EV, Skobeleva A, Therville N, Bailbe D, Gorshkova I, Natarajan V, Portha B and Le Stunff H. Ceramide synthase 4 and de novo production of ceramides with specific N-acyl chain lengths are involved in glucolipototoxicity-induced apoptosis of INS-1 beta-cells. *Biochem J* 2011; 438: 177-189.
- [66] Abou-Ghali M and Stiban J. Regulation of ceramide channel formation and disassembly: Insights on the initiation of apoptosis. *Saudi J Biol Sci* 2015; 22: 760-772.
- [67] Chang KT, Anishkin A, Patwardhan GA, Beverly LJ, Siskind LJ and Colombini M. Ceramide channels: destabilization by Bcl-xL and role in apoptosis. *Biochim Biophys Acta* 2015; 1848: 2374-2384.
- [68] Bradley E, Dasgupta S, Jiang X, Zhao X, Zhu G, He Q, Dinkins M, Bieberich E and Wang G. Critical role of Spns2, a sphingosine-1-phosphate transporter, in lung cancer cell survival and migration. *PLoS One* 2014; 9: e110119.
- [69] Rutherford C, Childs S, Ohotski J, McGlynn L, Riddick M, MacFarlane S, Tasker D, Pyne S, Pyne NJ, Edwards J and Palmer TM. Regulation of cell survival by sphingosine-1-phosphate receptor S1P1 via reciprocal ERK-dependent suppression of Bim and PI-3-kinase/protein kinase C-mediated upregulation of Mcl-1. *Cell Death Dis* 2013; 4: e927.
- [70] Long J, Darroch P, Wan KF, Kong KC, Ktistakis N, Pyne NJ and Pyne S. Regulation of cell survival by lipid phosphate phosphatases involves the modulation of intracellular phosphatidic acid and sphingosine 1-phosphate pools. *Biochem J* 2005; 391: 25-32.
- [71] Moriuie T, Igarashi J, Yoneda K, Hashimoto T, Nakai K, Kosaka H and Kubota Y. Sphingosine 1-phosphate attenuates peroxide-induced apoptosis in HaCaT cells cultured in vitro. *Clin Exp Dermatol* 2013; 38: 638-645.
- [72] Schmitz EI, Potteck H, Schuppel M, Manggau M, Wahyidin E and Kleuser B. Sphingosine 1-phosphate protects primary human keratinocytes from apoptosis via nitric oxide formation through the receptor subtype S1P(3). *Mol Cell Biochem* 2012; 371: 165-176.
- [73] Nakahara T, Iwase A, Nakamura T, Kondo M, Bayasula, Kobayashi H, Takikawa S, Manabe S, Goto M, Kotani T and Kikkawa F. Sphingosine-1-phosphate inhibits H2O2-induced granulosa cell apoptosis via the PI3K/Akt signaling pathway. *Fertil Steril* 2012; 98: 1001-1008, e1001.
- [74] Gangoiti P, Arana L, Ouro A, Granado MH, Trueba M and Gomez-Munoz A. Activation of mTOR and RhoA is a major mechanism by which Ceramide 1-phosphate stimulates macrophage proliferation. *Cell Signal* 2011; 23: 27-34.
- [75] Gangoiti P, Granado MH, Arana L, Ouro A and Gomez-Munoz A. Activation of protein kinase C-alpha is essential for stimulation of cell proliferation by ceramide 1-phosphate. *FEBS Lett* 2010; 584: 517-524.
- [76] Gomez-Munoz A, Gangoiti P, Granado MH, Arana L and Ouro A. Ceramide-1-phosphate in cell survival and inflammatory signaling. *Adv Exp Med Biol* 2010; 688: 118-130.

Sphingolipids in SCI

- [77] Gangoiti P, Granado MH, Arana L, Ouro A and Gomez-Munoz A. Involvement of nitric oxide in the promotion of cell survival by ceramide 1-phosphate. *FEBS Lett* 2008; 582: 2263-2269.
- [78] Gangoiti P, Granado MH, Wang SW, Kong JY, Steinbrecher UP and Gomez-Munoz A. Ceramide 1-phosphate stimulates macrophage proliferation through activation of the PI3-kinase/PKB, JNK and ERK1/2 pathways. *Cell Signal* 2008; 20: 726-736.
- [79] Gomez-Munoz A, Frago LM, Alvarez L and Varela-Nieto I. Stimulation of DNA synthesis by natural ceramide 1-phosphate. *Biochem J* 1997; 325: 435-440.
- [80] Granado MH, Gangoiti P, Ouro A, Arana L and Gomez-Munoz A. Ceramide 1-phosphate inhibits serine palmitoyltransferase and blocks apoptosis in alveolar macrophages. *Biochim Biophys Acta* 2009; 1791: 263-272.
- [81] Gomez-Munoz A, Kong JY, Salh B and Steinbrecher UP. Ceramide-1-phosphate blocks apoptosis through inhibition of acid sphingomyelinase in macrophages. *J Lipid Res* 2004; 45: 99-105.
- [82] Mitra P, Maceyka M, Payne SG, Lamour N, Milstien S, Chalfant CE and Spiegel S. Ceramide kinase regulates growth and survival of A549 human lung adenocarcinoma cells. *FEBS Lett* 2007; 581: 735-740.
- [83] Xia P, Wang L, Gamble JR and Vadas MA. Activation of sphingosine kinase by tumor necrosis factor-alpha inhibits apoptosis in human endothelial cells. *J Biol Chem* 1999; 274: 34499-34505.
- [84] Tsukamoto S, Huang Y, Kumazoe M, Lesnick C, Yamada S, Ueda N, Suzuki T, Yamashita S, Kim YH, Fujimura Y, Miura D, Kay NE, Shanafelt TD and Tachibana H. Sphingosine Kinase-1 Protects Multiple Myeloma from Apoptosis Driven by Cancer-Specific Inhibition of RTKs. *Mol Cancer Ther* 2015; 14: 2303-2312.
- [85] Song L, Xiong H, Li J, Liao W, Wang L, Wu J and Li M. Sphingosine kinase-1 enhances resistance to apoptosis through activation of PI3K/Akt/NF-kappaB pathway in human non-small cell lung cancer. *Clin Cancer Res* 2011; 17: 1839-1849.
- [86] Guan H, Song L, Cai J, Huang Y, Wu J, Yuan J, Li J and Li M. Sphingosine kinase 1 regulates the Akt/FOXO3a/Bim pathway and contributes to apoptosis resistance in glioma cells. *PLoS One* 2011; 6: e19946.
- [87] Igarashi N, Okada T, Hayashi S, Fujita T, Jahangeer S and Nakamura S. Sphingosine kinase 2 is a nuclear protein and inhibits DNA synthesis. *J Biol Chem* 2003; 278: 46832-46839.
- [88] Liu H, Toman RE, Goparaju SK, Maceyka M, Nava VE, Sankala H, Payne SG, Bektas M, Ishii I, Chun J, Milstien S and Spiegel S. Sphingosine kinase type 2 is a putative BH3-only protein that induces apoptosis. *J Biol Chem* 2003; 278: 40330-40336.
- [89] Li PH, Wu JX, Zheng JN and Pei DS. A sphingosine kinase-1 inhibitor, SKI-II, induces growth inhibition and apoptosis in human gastric cancer cells. *Asian Pac J Cancer Prev* 2014; 15: 10381-10385.
- [90] Hara-Yokoyama M, Terasawa K, Ichinose S, Watanabe A, Podyma-Inoue KA, Akiyoshi K, Igarashi Y and Yanagishita M. Sphingosine kinase 2 inhibitor SG-12 induces apoptosis via phosphorylation by sphingosine kinase 2. *Bioorg Med Chem Lett* 2013; 23: 2220-2224.
- [91] Karimian G, Buist-Homan M, Schmidt M, Tietge UJ, de Boer JF, Klappe K, Kok JW, Combettes L, Tordjmann T, Faber KN and Moshage H. Sphingosine kinase-1 inhibition protects primary rat hepatocytes against bile salt-induced apoptosis. *Biochim Biophys Acta* 2013; 1832: 1922-1929.
- [92] Maceyka M, Sankala H, Hait NC, Le Stunff H, Liu H, Toman R, Collier C, Zhang M, Satin LS, Merrill AH Jr, Milstien S and Spiegel S. SphK1 and SphK2, sphingosine kinase isoenzymes with opposing functions in sphingolipid metabolism. *J Biol Chem* 2005; 280: 37118-37129.
- [93] Mehra VC, Jackson E, Zhang XM, Jiang XC, Dobrucki LW, Yu J, Bernatchez P, Sinusas AJ, Shulman GI, Sessa WC, Yarovinsky TO and Bender JR. Ceramide-activated phosphatase mediates fatty acid-induced endothelial VEGF resistance and impaired angiogenesis. *Am J Pathol* 2014; 184: 1562-1576.
- [94] Novgorodov SA and Gudzi TI. Ceramide and mitochondria in ischemic brain injury. *Int J Biochem Mol Biol* 2011; 2: 347-361.
- [95] Wang H, Cai KY, Li W and Huang H. Sphingosine-1-Phosphate Induces the Migration and Angiogenesis of Epc3 Through the Akt Signaling Pathway via Sphingosine-1-Phosphate Receptor 3/Platelet-Derived Growth Factor Receptor-beta. *Cell Mol Biol Lett* 2015; 20: 597-611.
- [96] Abuhusain HJ, Matin A, Qiao Q, Shen H, Kain N, Day BW, Stringer BW, Daniels B, Laaksonen MA, Teo C, McDonald KL and Don AS. A metabolic shift favoring sphingosine 1-phosphate at the expense of ceramide controls glioblastoma angiogenesis. *J Biol Chem* 2013; 288: 37355-37364.
- [97] Su SC and Bayless KJ. Utilizing sphingosine-1-phosphate to stimulate sprouting angiogenesis. *Methods Mol Biol* 2012; 874: 201-213.
- [98] Nagahashi M, Ramachandran S, Kim EY, Allegood JC, Rashid OM, Yamada A, Zhao R, Milstien S, Zhou H, Spiegel S and Takabe K. Sphingosine-1-phosphate produced by sphin-

Sphingolipids in SCI

- gოსine kinase 1 promotes breast cancer progression by stimulating angiogenesis and lymphangiogenesis. *Cancer Res* 2012; 72: 726-735.
- [99] Dmitrieva VG, Torshina EV, Yuzhakov VV, Povarova OV, Skvortsova VI, Limborska SA and Dergunova LV. Expression of sphingomyelin synthase 1 gene in rat brain focal ischemia. *Brain Res* 2008; 1188: 222-227.
- [100] Ohtani R, Tomimoto H, Kondo T, Wakita H, Akiguchi I, Shibasaki H and Okazaki T. Upregulation of ceramide and its regulating mechanism in a rat model of chronic cerebral ischemia. *Brain Res* 2004; 1023: 31-40.
- [101] Yu ZF, Nikolova-Karakashian M, Zhou D, Cheng G, Schuchman EH and Mattson MP. Pivotal role for acidic sphingomyelinase in cerebral ischemia-induced ceramide and cytokine production, and neuronal apoptosis. *J Mol Neurosci* 2000; 15: 85-97.
- [102] Nakane M, Kubota M, Nakagomi T, Tamura A, Hisaki H, Shimasaki H and Ueta N. Lethal fore-brain ischemia stimulates sphingomyelin hydrolysis and ceramide generation in the gerbil hippocampus. *Neurosci Lett* 2000; 296: 89-92.
- [103] Soeda S, Tsuji Y, Ochiai T, Mishima K, Iwasaki K, Fujiwara M, Yokomatsu T, Murano T, Shibuya S and Shimeno H. Inhibition of sphingomyelinase activity helps to prevent neuron death caused by ischemic stress. *Neurochem Int* 2004; 45: 619-626.
- [104] Chen Y, Ginis I and Hallenbeck JM. The protective effect of ceramide in immature rat brain hypoxia-ischemia involves up-regulation of bcl-2 and reduction of TUNEL-positive cells. *J Cereb Blood Flow Metab* 2001; 21: 34-40.
- [105] Zheng S, Wei S, Wang X, Xu Y, Xiao Y, Liu H, Jia J and Cheng J. Sphingosine kinase 1 mediates neuroinflammation following cerebral ischemia. *Exp Neurol* 2015; 272: 160-169.
- [106] Hasegawa Y, Suzuki H, Altay O, Rolland W and Zhang JH. Role of the sphingosine metabolism pathway on neurons against experimental cerebral ischemia in rats. *Transl Stroke Res* 2013; 4: 524-532.
- [107] Pfeilschifter W, Czech-Zechmeister B, Sujak M, Mirceska A, Koch A, Rami A, Steinmetz H, Foerch C, Huwiler A and Pfeilschifter J. Activation of sphingosine kinase 2 is an endogenous protective mechanism in cerebral ischemia. *Biochem Biophys Res Commun* 2011; 413: 212-217.
- [108] Hasegawa Y, Suzuki H, Sozen T, Rolland W and Zhang JH. Activation of sphingosine 1-phosphate receptor-1 by FTY720 is neuroprotective after ischemic stroke in rats. *Stroke* 2010; 41: 368-374.
- [109] Czech B, Pfeilschifter W, Mazaheri-Omrani N, Strobel MA, Kahles T, Neumann-Haefelin T, Rami A, Huwiler A and Pfeilschifter J. The immunomodulatory sphingosine 1-phosphate analog FTY720 reduces lesion size and improves neurological outcome in a mouse model of cerebral ischemia. *Biochem Biophys Res Commun* 2009; 389: 251-256.
- [110] Kimura A, Ohmori T, Kashiwakura Y, Ohkawa R, Madoiwa S, Mimuro J, Shimazaki K, Hoshino Y, Yatomi Y and Sakata Y. Antagonism of sphingosine 1-phosphate receptor-2 enhances migration of neural progenitor cells toward an area of brain. *Stroke* 2008; 39: 3411-3417.
- [111] Sun Y, Jin K, Xie L, Childs J, Mao XO, Logvinova A and Greenberg DA. VEGF-induced neuroprotection, neurogenesis, and angiogenesis after focal cerebral ischemia. *J Clin Invest* 2003; 111: 1843-1851.
- [112] Ausprunk DH and Folkman J. Migration and proliferation of endothelial cells in preformed and newly formed blood vessels during tumor angiogenesis. *Microvasc Res* 1977; 14: 53-65.
- [113] Kamei N, Kwon SM, Kawamoto A, Ii M, Ishikawa M, Ochi M and Asahara T. Contribution of bone marrow-derived endothelial progenitor cells to neovascularization and astrogliosis following spinal cord injury. *J Neurosci Res* 2012; 90: 2281-2292.
- [114] Herrera JJ, Sundberg LM, Zentilin L, Giacca M and Narayana PA. Sustained expression of vascular endothelial growth factor and angiotensin-1 improves blood-spinal cord barrier integrity and functional recovery after spinal cord injury. *J Neurotrauma* 2010; 27: 2067-2076.
- [115] Widenfalk J, Lipson A, Jubran M, Hofstetter C, Ebendal T, Cao Y and Olson L. Vascular endothelial growth factor improves functional outcome and decreases secondary degeneration in experimental spinal cord contusion injury. *Neuroscience* 2003; 120: 951-960.
- [116] Limaye V. The role of sphingosine kinase and sphingosine-1-phosphate in the regulation of endothelial cell biology. *Endothelium* 2008; 15: 101-112.
- [117] Mulders AC, Peters SL and Michel MC. Sphingomyelin metabolism and endothelial cell function. *Eur Heart J* 2007; 28: 777-779.
- [118] Alford SK, Kaneda MM, Wacker BK and Elbert DL. Endothelial cell migration in human plasma is enhanced by a narrow range of added sphingosine 1-phosphate: implications for biomaterials design. *J Biomed Mater Res A* 2009; 88: 205-212.
- [119] Okamoto H, Yatomi Y, Ohmori T, Satoh K, Matsumoto Y and Ozaki Y. Sphingosine 1-phosphate stimulates G(i)- and Rho-mediated vascular endothelial cell spreading and migration. *Thromb Res* 2000; 99: 259-265.

Sphingolipids in SCI

- [120] Xu M, Waters CL, Hu C, Wysolmerski RB, Vincent PA and Minnear FL. Sphingosine 1-phosphate rapidly increases endothelial barrier function independently of VE-cadherin but requires cell spreading and Rho kinase. *Am J Physiol Cell Physiol* 2007; 293: C1309-1318.
- [121] Itagaki K, Yun JK, Hengst JA, Yatani A, Hauser CJ, Spolarics Z and Deitch EA. Sphingosine 1-phosphate has dual functions in the regulation of endothelial cell permeability and Ca²⁺ metabolism. *J Pharmacol Exp Ther* 2007; 323: 186-191.
- [122] McVerry BJ and Garcia JG. Endothelial cell barrier regulation by sphingosine 1-phosphate. *J Cell Biochem* 2004; 92: 1075-1085.
- [123] Langlois S, Gingras D and Beliveau R. Membrane type 1-matrix metalloproteinase (MT1-MMP) cooperates with sphingosine 1-phosphate to induce endothelial cell migration and morphogenic differentiation. *Blood* 2004; 103: 3020-3028.
- [124] Lee H, Goetzl EJ and An S. Lysophosphatidic acid and sphingosine 1-phosphate stimulate endothelial cell wound healing. *Am J Physiol Cell Physiol* 2000; 278: C612-618.
- [125] Kimura T, Sato K, Malchinkhuu E, Tomura H, Tamama K, Kuwabara A, Murakami M and Okajima F. High-density lipoprotein stimulates endothelial cell migration and survival through sphingosine 1-phosphate and its receptors. *Arterioscler Thromb Vasc Biol* 2003; 23: 1283-1288.
- [126] McGuire PG, Rangasamy S, Maestas J and Das A. Pericyte-derived sphingosine 1-phosphate induces the expression of adhesion proteins and modulates the retinal endothelial cell barrier. *Arterioscler Thromb Vasc Biol* 2011; 31: e107-115.
- [127] Heo K, Park KA, Kim YH, Kim SH, Oh YS, Kim IH, Ryu SH and Suh PG. Sphingosine 1-phosphate induces vascular endothelial growth factor expression in endothelial cells. *BMB Rep* 2009; 42: 685-690.
- [128] Bonder CS, Sun WY, Matthews T, Cassano C, Li X, Ramshaw HS, Pitson SM, Lopez AF, Coates PT, Proia RL, Vadas MA and Gamble JR. Sphingosine kinase regulates the rate of endothelial progenitor cell differentiation. *Blood* 2009; 113: 2108-2117.
- [129] Dimasi DP, Pitson SM and Bonder CS. Examining the Role of Sphingosine Kinase-2 in the Regulation of Endothelial Cell Barrier Integrity. *Microcirculation* 2016; 23: 248-265.
- [130] Gamble JR, Sun WY, Li X, Hahn CN, Pitson SM, Vadas MA and Bonder CS. Sphingosine kinase-1 associates with integrin {alpha}V{beta}3 to mediate endothelial cell survival. *Am J Pathol* 2009; 175: 2217-2225.
- [131] Schwalm S, Pfeilschifter J and Huwiler A. Sphingosine kinase 1 is critically involved in nitric oxide-mediated human endothelial cell migration and tube formation. *Br J Pharmacol* 2010; 160: 1641-1651.
- [132] Yan G, Chen S, You B and Sun J. Activation of sphingosine kinase-1 mediates induction of endothelial cell proliferation and angiogenesis by epoxyeicosatrienoic acids. *Cardiovasc Res* 2008; 78: 308-314.
- [133] Lindner K, Uhlig U and Uhlig S. Ceramide alters endothelial cell permeability by a nonapoptotic mechanism. *Br J Pharmacol* 2005; 145: 132-140.
- [134] Venable ME and Yin X. Ceramide induces endothelial cell senescence. *Cell Biochem Funct* 2009; 27: 547-551.
- [135] Lang Z, Guerrero M, Li R and Ladisch S. Ganglioside GD1a enhances VEGF-induced endothelial cell proliferation and migration. *Biochem Biophys Res Commun* 2001; 282: 1031-1037.
- [136] Liu Y, McCarthy J and Ladisch S. Membrane ganglioside enrichment lowers the threshold for vascular endothelial cell angiogenic signaling. *Cancer Res* 2006; 66: 10408-10414.
- [137] Rajesh M, Kolmakova A and Chatterjee S. Novel role of lactosylceramide in vascular endothelial growth factor-mediated angiogenesis in human endothelial cells. *Circ Res* 2005; 97: 796-804.
- [138] Mukherjee P, Faber AC, Shelton LM, Baek RC, Chiles TC and Seyfried TN. Thematic review series: sphingolipids. Ganglioside GM3 suppresses the proangiogenic effects of vascular endothelial growth factor and ganglioside GD1a. *J Lipid Res* 2008; 49: 929-938.
- [139] Kulinski JM, Munoz-Cano R and Olivera A. Sphingosine-1-phosphate and other lipid mediators generated by mast cells as critical players in allergy and mast cell function. *Eur J Pharmacol* 2016; 778: 56-67.
- [140] Wang Z, Fan H, Xie R, Yang J, Ren Y, Yang Y and Li W. The Effect of Sphingosine 1-Phosphate/Sphingosine 1-Phosphate Receptor on Neutrophil Function and the Relevant Signaling Pathway. *Acta Haematol* 2015; 134: 49-56.
- [141] Oskeritzian CA. Mast cell plasticity and sphingosine-1-phosphate in immunity, inflammation and cancer. *Mol Immunol* 2015; 63: 104-112.
- [142] Arlt O, Schwiebs A, Japtok L, Ruger K, Katzy E, Kleuser B and Radeke HH. Sphingosine-1-phosphate modulates dendritic cell function: focus on non-migratory effects in vitro and in vivo. *Cell Physiol Biochem* 2014; 34: 27-44.
- [143] Keul P, Lucke S, von Wnuck Lipinski K, Bode C, Graler M, Heusch G and Levkau B. Sphingosine-1-phosphate receptor 3 promotes recruitment of monocyte/macrophages in inflammation and atherosclerosis. *Circ Res* 2011; 108: 314-323.

Sphingolipids in SCI

- [144] Nayak D, Huo Y, Kwang WX, Pushparaj PN, Kumar SD, Ling EA and Dheen ST. Sphingosine kinase 1 regulates the expression of proinflammatory cytokines and nitric oxide in activated microglia. *Neuroscience* 2010; 166: 132-144.
- [145] Pan JZ, Ni L, Sodhi A, Aguanno A, Young W and Hart RP. Cytokine activity contributes to induction of inflammatory cytokine mRNAs in spinal cord following contusion. *J Neurosci Res* 2002; 68: 315-322.
- [146] Gomez-Munoz A, Presa N, Gomez-Larrauri A, Rivera IG, Trueba M and Ordonez M. Control of inflammatory responses by ceramide, sphingosine 1-phosphate and ceramide 1-phosphate. *Prog Lipid Res* 2016; 61: 51-62.
- [147] Payne SG, Milstien S, Barbour SE and Spiegel S. Modulation of adaptive immune responses by sphingosine-1-phosphate. *Semin Cell Dev Biol* 2004; 15: 521-527.
- [148] Jin Y, Knudsen E, Wang L, Bryceson Y, Damaj B, Gessani S and Maghazachi AA. Sphingosine 1-phosphate is a novel inhibitor of T-cell proliferation. *Blood* 2003; 101: 4909-4915.
- [149] Schaper K, Kietzmann M and Baumer W. Sphingosine-1-phosphate differently regulates the cytokine production of IL-12, IL-23 and IL-27 in activated murine bone marrow derived dendritic cells. *Mol Immunol* 2014; 59: 10-18.
- [150] Lv M, Zhang D, Dai D, Zhang W and Zhang L. Sphingosine kinase 1/sphingosine-1-phosphate regulates the expression of interleukin-17A in activated microglia in cerebral ischemia/reperfusion. *Inflamm Res* 2016; 65: 551-562.
- [151] Schwartz BM, Hong G, Morrison BH, Wu W, Baudhuin LM, Xiao YJ, Mok SC and Xu Y. Lysophospholipids increase interleukin-8 expression in ovarian cancer cells. *Gynecol Oncol* 2001; 81: 291-300.
- [152] Xia P, Wang L, Moretti PA, Albanese N, Chai F, Pitson SM, D'Andrea RJ, Gamble JR and Vadas MA. Sphingosine kinase interacts with TRAF2 and dissects tumor necrosis factor- α signaling. *J Biol Chem* 2002; 277: 7996-8003.
- [153] Yoshimoto T, Furuhashi M, Kamiya S, Hisada M, Miyaji H, Magami Y, Yamamoto K, Fujiwara H and Mizuguchi J. Positive modulation of IL-12 signaling by sphingosine kinase 2 associating with the IL-12 receptor beta 1 cytoplasmic region. *J Immunol* 2003; 171: 1352-1359.
- [154] Hankins JL, Fox TE, Barth BM, Unrath KA and Kester M. Exogenous ceramide-1-phosphate reduces lipopolysaccharide (LPS)-mediated cytokine expression. *J Biol Chem* 2011; 286: 44357-44366.
- [155] Pettus BJ, Kitatani K, Chalfant CE, Taha TA, Kawamori T, Bielawski J, Obeid LM and Hannun YA. The coordination of prostaglandin E2 production by sphingosine-1-phosphate and ceramide-1-phosphate. *Mol Pharmacol* 2005; 68: 330-335.
- [156] Pettus BJ, Bielawska A, Subramanian P, Wijesinghe DS, Maceyka M, Leslie CC, Evans JH, Freiberg J, Roddy P, Hannun YA and Chalfant CE. Ceramide 1-phosphate is a direct activator of cytosolic phospholipase A2. *J Biol Chem* 2004; 279: 11320-11326.
- [157] Subramanian P, Stahelin RV, Szulc Z, Bielawska A, Cho W and Chalfant CE. Ceramide 1-phosphate acts as a positive allosteric activator of group IVA cytosolic phospholipase A2 α and enhances the interaction of the enzyme with phosphatidylcholine. *J Biol Chem* 2005; 280: 17601-17607.
- [158] Mechtcheriakova D, Wlachos A, Sobanov J, Kopp T, Reuschel R, Bornancin F, Cai R, Zemann B, Urtz N, Stingl G, Zlabinger G, Woisetschlager M, Baumruker T and Billich A. Sphingosine 1-phosphate phosphatase 2 is induced during inflammatory responses. *Cell Signal* 2007; 19: 748-760.
- [159] Zhao Y, Usatyuk PV, Cummings R, Saatian B, He D, Watkins T, Morris A, Spannhake EW, Brindley DN and Natarajan V. Lipid phosphate phosphatase-1 regulates lysophosphatidic acid-induced calcium release, NF- κ B activation and interleukin-8 secretion in human bronchial epithelial cells. *Biochem J* 2005; 385: 493-502.
- [160] Gu L, Huang B, Shen W, Gao L, Ding Z, Wu H and Guo J. Early activation of nSMase2/ceramide pathway in astrocytes is involved in ischemia-associated neuronal damage via inflammation in rat hippocampi. *J Neuroinflammation* 2013; 10: 109.
- [161] Roeske-Nielsen A, Fredman P, Mansson JE, Bendtzen K and Buschard K. Beta-galactosylceramide increases and sulfatide decreases cytokine and chemokine production in whole blood cells. *Immunol Lett* 2004; 91: 205-211.
- [162] David S, Greenhalgh AD and Kroner A. Macrophage and microglial plasticity in the injured spinal cord. *Neuroscience* 2015; 307: 311-318.
- [163] Gensel JC and Zhang B. Macrophage activation and its role in repair and pathology after spinal cord injury. *Brain Res* 2015; 1619: 1-11.
- [164] Zhou X, He X and Ren Y. Function of microglia and macrophages in secondary damage after spinal cord injury. *Neural Regen Res* 2014; 9: 1787-1795.
- [165] Greenhalgh AD and David S. Differences in the phagocytic response of microglia and peripheral macrophages after spinal cord injury and its effects on cell death. *J Neurosci* 2014; 34: 6316-6322.
- [166] Ren Y and Young W. Managing inflammation after spinal cord injury through manipulation

- of macrophage function. *Neural Plast* 2013; 2013: 945034.
- [167] Guo L, Rolfe AJ, Wang X, Tai W, Cheng Z, Cao K, Chen X, Xu Y, Sun D, Li J, He X, Young W, Fan J and Ren Y. Rescuing macrophage normal function in spinal cord injury with embryonic stem cell conditioned media. *Mol Brain* 2016; 9: 48.
- [168] Wang X, Cao K, Sun X, Chen Y, Duan Z, Sun L, Guo L, Bai P, Sun D, Fan J, He X, Young W and Ren Y. Macrophages in spinal cord injury: phenotypic and functional change from exposure to myelin debris. *Glia* 2015; 63: 635-651.
- [169] Weigert A, Weis N and Brune B. Regulation of macrophage function by sphingosine-1-phosphate. *Immunobiology* 2009; 214: 748-760.
- [170] Gomez-Munoz A, Kong J, Salh B and Steinbrecher UP. Sphingosine-1-phosphate inhibits acid sphingomyelinase and blocks apoptosis in macrophages. *FEBS Lett* 2003; 539: 56-60.
- [171] Weis N, Weigert A, von Knethen A and Brune B. Heme oxygenase-1 contributes to an alternative macrophage activation profile induced by apoptotic cell supernatants. *Mol Biol Cell* 2009; 20: 1280-1288.
- [172] Weigert A, Johann AM, von Knethen A, Schmidt H, Geisslinger G and Brune B. Apoptotic cells promote macrophage survival by releasing the antiapoptotic mediator sphingosine-1-phosphate. *Blood* 2006; 108: 1635-1642.
- [173] Xie B, Shen J, Dong A, Rashid A, Stoller G and Campochiaro PA. Blockade of sphingosine-1-phosphate reduces macrophage influx and retinal and choroidal neovascularization. *J Cell Physiol* 2009; 218: 192-198.
- [174] Gude DR, Alvarez SE, Paugh SW, Mitra P, Yu J, Griffiths R, Barbour SE, Milstien S and Spiegel S. Apoptosis induces expression of sphingosine kinase 1 to release sphingosine-1-phosphate as a "come-and-get-me" signal. *FASEB J* 2008; 22: 2629-2638.
- [175] Schwab SR and Cyster JG. Finding a way out: lymphocyte egress from lymphoid organs. *Nat Immunol* 2007; 8: 1295-1301.
- [176] Weigert A, Tzieply N, von Knethen A, Johann AM, Schmidt H, Geisslinger G and Brune B. Tumor cell apoptosis polarizes macrophages role of sphingosine-1-phosphate. *Mol Biol Cell* 2007; 18: 3810-3819.
- [177] Zhang Z, Zhang ZY, Fauser U and Schluesener HJ. FTY720 ameliorates experimental autoimmune neuritis by inhibition of lymphocyte and monocyte infiltration into peripheral nerves. *Exp Neurol* 2008; 210: 681-690.
- [178] Rausch M, Hiestand P, Foster CA, Baumann DR, Cannet C and Rudin M. Predictability of FTY720 efficacy in experimental autoimmune encephalomyelitis by in vivo macrophage tracking: clinical implications for ultrasmall superparamagnetic iron oxide-enhanced magnetic resonance imaging. *J Magn Reson Imaging* 2004; 20: 16-24.
- [179] Fujino M, Funeshima N, Kitazawa Y, Kimura H, Amemiya H, Suzuki S and Li XK. Amelioration of experimental autoimmune encephalomyelitis in Lewis rats by FTY720 treatment. *J Pharmacol Exp Ther* 2003; 305: 70-77.
- [180] Sethu S, Mendez-Corao G and Melendez AJ. Phospholipase D1 plays a key role in TNF-alpha signaling. *J Immunol* 2008; 180: 6027-6034.
- [181] Melendez AJ and Ibrahim FB. Antisense knockdown of sphingosine kinase 1 in human macrophages inhibits C5a receptor-dependent signal transduction, Ca²⁺ signals, enzyme release, cytokine production, and chemotaxis. *J Immunol* 2004; 173: 1596-1603.
- [182] Wu W, Mosteller RD and Broek D. Sphingosine kinase protects lipopolysaccharide-activated macrophages from apoptosis. *Mol Cell Biol* 2004; 24: 7359-7369.
- [183] Hammad SM, Crellin HG, Wu BX, Melton J, Anelli V and Obeid LM. Dual and distinct roles for sphingosine kinase 1 and sphingosine 1 phosphate in the response to inflammatory stimuli in RAW macrophages. *Prostaglandins Other Lipid Mediat* 2008; 85: 107-114.
- [184] Arana L, Ordonez M, Ouro A, Rivera IG, Gangoiti P, Trueba M and Gomez-Munoz A. Ceramide 1-phosphate induces macrophage chemoattractant protein-1 release: involvement in ceramide 1-phosphate-stimulated cell migration. *Am J Physiol Endocrinol Metab* 2013; 304: E1213-1226.
- [185] Sofroniew MV and Vinters HV. Astrocytes: biology and pathology. *Acta Neuropathol* 2010; 119: 7-35.
- [186] Karimi-Abdolrezaee S and Billakanti R. Reactive astrogliosis after spinal cord injury-beneficial and detrimental effects. *Mol Neurobiol* 2012; 46: 251-264.
- [187] Facci L, Skaper SD, Favaron M and Leon A. A role for gangliosides in astroglial cell differentiation in vitro. *J Cell Biol* 1988; 106: 821-828.
- [188] Katoh-Semba R, Facci L, Skaper SD and Varon S. Gangliosides stimulate astroglial cell proliferation in the absence of serum. *J Cell Physiol* 1986; 126: 147-153.
- [189] Pyo H, Joe E, Jung S, Lee SH and Jou I. Gangliosides activate cultured rat brain microglia. *J Biol Chem* 1999; 274: 34584-34589.
- [190] Kim S, Steelman AJ, Zhang Y, Kinney HC and Li J. Aberrant upregulation of astroglial ceramide potentiates oligodendrocyte injury. *Brain Pathol* 2012; 22: 41-57.
- [191] Nakajima K, Tohyama Y, Kohsaka S and Kurihara T. Ceramide activates microglia to enhance the production/secretion of brain-de-

Sphingolipids in SCI

- rived neurotrophic factor (BDNF) without induction of deleterious factors in vitro. *J Neurochem* 2002; 80: 697-705.
- [192] Pahan K, Sheikh FG, Khan M, Namboodiri AM and Singh I. Sphingomyelinase and ceramide stimulate the expression of inducible nitric-oxide synthase in rat primary astrocytes. *J Biol Chem* 1998; 273: 2591-2600.
- [193] Jeon SB, Yoon HJ, Park SH, Kim IH and Park EJ. Sulfatide, a major lipid component of myelin sheath, activates inflammatory responses as an endogenous stimulator in brain-resident immune cells. *J Immunol* 2008; 181: 8077-8087.
- [194] Stenovec M, Trkov S, Kreft M and Zorec R. Alterations of calcium homeostasis in cultured rat astrocytes evoked by bioactive sphingolipids. *Acta Physiol (Oxf)* 2014; 212: 49-61.
- [195] Trkov S, Stenovec M, Kreft M, Potokar M, Parpura V, Davletov B and Zorec R. Fingolimod-a sphingosine-like molecule inhibits vesicle mobility and secretion in astrocytes. *Glia* 2012; 60: 1406-1416.
- [196] Colombo E, Di Dario M, Capitolo E, Chaabane L, Newcombe J, Martino G and Farina C. Fingolimod may support neuroprotection via blockade of astrocyte nitric oxide. *Ann Neurol* 2014; 76: 325-337.
- [197] Mullershausen F, Craveiro LM, Shin Y, Cortes-Cros M, Bassilana F, Osinde M, Wishart WL, Guerini D, Thallmair M, Schwab ME, Sivasankaran R, Seuwen K and Dev KK. Phosphorylated FTY720 promotes astrocyte migration through sphingosine-1-phosphate receptors. *J Neurochem* 2007; 102: 1151-1161.
- [198] Noda H, Takeuchi H, Mizuno T and Suzumura A. Fingolimod phosphate promotes the neuroprotective effects of microglia. *J Neuroimmunol* 2013; 256: 13-18.
- [199] Kobayashi Y, Kiguchi N, Maeda T, Ozaki M and Kishioka S. The critical role of spinal ceramide in the development of partial sciatic nerve ligation-induced neuropathic pain in mice. *Biochem Biophys Res Commun* 2012; 421: 318-322.
- [200] Miller SM. Methylprednisolone in acute spinal cord injury: a tarnished standard. *J Neurosurg Anesthesiol* 2008; 20: 140-142.
- [201] Chun J and Brinkmann V. A mechanistically novel, first oral therapy for multiple sclerosis: the development of fingolimod (FTY720, Gilenya). *Discov Med* 2011; 12: 213-228.
- [202] Zecri FJ. From Natural Product to the First Oral Treatment for Multiple Sclerosis: The Discovery of FTY720 (Gilenya)? *Curr Opin Chem Biol* 2016; 32: 60-66.
- [203] Chen Y, Ma Z, Min L, Li H, Wang B, Zhong J and Dai L. Biomarker identification and pathway analysis by serum metabolomics of lung cancer. *Biomed Res Int* 2015; 2015: 183624.
- [204] Di Marzio L, Di Leo A, Cinque B, Fanini D, Agnifili A, Berloco P, Linsalata M, Lorusso D, Barone M, De Simone C and Cifone MG. Detection of alkaline sphingomyelinase activity in human stool: proposed role as a new diagnostic and prognostic marker of colorectal cancer. *Cancer Epidemiol Biomarkers Prev* 2005; 14: 856-862.
- [205] Dubois N, Rio E, Ripoché N, Ferchaud-Roucher V, Gaugler MH, Campion L, Krempf M, Carrie C, Mahe M, Mirabel X and Paris F. Plasma ceramide, a real-time predictive marker of pulmonary and hepatic metastases response to stereotactic body radiation therapy combined with irinotecan. *Radiother Oncol* 2016; 119: 229-235.
- [206] Fox TE, Bewley MC, Unrath KA, Pedersen MM, Anderson RE, Jung DY, Jefferson LS, Kim JK, Bronson SK, Flanagan JM and Kester M. Circulating sphingolipid biomarkers in models of type 1 diabetes. *J Lipid Res* 2011; 52: 509-517.
- [207] Grammatikos G, Muhle C, Ferreiros N, Schroeter S, Bogdanou D, Schwalm S, Hintereder G, Kornhuber J, Zeuzem S, Sarrazin C and Pfeilschifter J. Serum acid sphingomyelinase is upregulated in chronic hepatitis C infection and non alcoholic fatty liver disease. *Biochim Biophys Acta* 2014; 1841: 1012-1020.
- [208] Koresawa R, Yamazaki K, Oka D, Fujiwara H, Nishimura H, Akiyama T, Hamasaki S, Wada H, Sugihara T and Sadahira Y. Sphingosine-1-phosphate receptor 1 as a prognostic biomarker and therapeutic target for patients with primary testicular diffuse large B-cell lymphoma. *Br J Haematol* 2016; 174: 264-74.
- [209] Mielke MM and Haughey NJ. Could plasma sphingolipids be diagnostic or prognostic biomarkers for Alzheimer's disease? *Clin Lipidol* 2012; 7: 525-536.
- [210] Sheth SA, Iavarone AT, Liebeskind DS, Won SJ and Swanson RA. Targeted Lipid Profiling Discovers Plasma Biomarkers of Acute Brain Injury. *PLoS One* 2015; 10: e0129735.
- [211] Zuellig RA, Hornemann T, Othman A, Hehl AB, Bode H, Guntert T, Ogunshola OO, Saponara E, Grabliauskaite K, Jang JH, Ungethuen U, Wei Y, von Eckardstein A, Graf R and Sonda S. Deoxysphingolipids, novel biomarkers for type 2 diabetes, are cytotoxic for insulin-producing cells. *Diabetes* 2014; 63: 1326-1339.