

# Interleukin 1 $\beta$ —A Potential Salivary Biomarker for Cancer Progression?

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**ABSTRACT:** The relationship between cancer and inflammation is a complex but intimate one. Decades of work has shown to us that cancer progression is influenced by a multitude of factors, including genetic, environmental, and immunological factors. We often overlook that cancer progression is also a pathological consequence of a dysregulated inflammatory control in the body. A current emerging topic in cancer research is the role of inflammasomes in carcinogenesis. The inflammasome is a multicomplex protein platform that when activated results in the release of proinflammatory cytokines, such as interleukin (IL)-1 $\beta$ . There is increasing evidence suggesting that IL-1 $\beta$  plays a pivotal role in cancer progression. This short review proposes the possibility of using IL-1 $\beta$  as a potential cancer progression biomarker and discusses the use of saliva as a model biological fluid for measuring physiological IL-1 $\beta$  levels in the body.

**KEYWORDS:** interleukin 1, saliva, cancer, biomarker, inflammasome

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## Introduction

Saliva is a clear, slightly acidic liquid, which is secreted from the salivary glands into the oral mouth cavity. Several years of research into identification of an array of biomolecules in saliva has opened new avenues into the field of salivary diagnostics. Given its noninvasive and rapid collection process, saliva possesses the qualities of a bodily fluid with high potential for the diagnosis, monitoring, and study of several diseases.<sup>1</sup> Saliva is able to provide reliable clinical data and cellular information concerning biologically active molecules that can be used to evaluate and monitor the health-disease process.<sup>2,3</sup> Approximately 27% of proteins are common between saliva and blood plasma.<sup>4</sup> Furthermore, saliva has an advantage over blood in that it contains low background of normal material and inhibitory substances and fewer complexes than blood.<sup>5</sup> Therefore, saliva is a good alternative to blood for diagnostic tests and presents a potential reservoir for researchers to identify new biomolecular markers.

Saliva can be a valuable resource for the diagnosis of both oral and systemic diseases. These diseases include periodontal disease, caries, and oral candidiasis, as well as different types of cancer, cardiovascular, endocrine, and infectious diseases (eg, bacteria, virus, and fungi).<sup>6</sup> Research conducted with saliva samples is proving to be promising in the detection or prediction of susceptibility to systemic diseases. Cancer diagnostics is an actively pursued area of salivary biomarker research. Several groups have conducted extensive investigations into potential biomarkers in saliva at the DNA, mRNA, and protein level in different types of cancer, including oral,<sup>5,7–10</sup>

salivary gland, head and neck,<sup>11</sup> ovarian,<sup>12,13</sup> breast,<sup>14,15</sup> gastric,<sup>16</sup> and lung cancers<sup>17</sup> (Table 1). Despite these investigations, a definitive biomarker to measure the extent and progression of the cancer remains to be discovered.

This review focuses on one potential cancer biomarker, interleukin (IL)-1 $\beta$  (also known as catabolin, IL-1, and IL1F2), which has recently received attention in immune-cancer biology research.

## IL-1 $\beta$ : A Potential Cancer Progression Biomarker?

IL-1 $\beta$  is a key inflammatory cytokine released upon infection, cellular injury, or antigenic challenge. This cytokine directly acts on various cell types, either alone or in combination with other inflammatory cytokines, to induce an inflammatory state or the “fever response.” IL-1 $\beta$  is a pro-inflammatory cytokine released upon activation of a multi-protein innate immune pathogen-sensing complex called the inflammasome.<sup>18</sup> The preformed IL-1 $\beta$  has to be proteolytically processed by an important cysteine protease, caspase 1, upon inflammasome activation to result in the secretion of an active and mature form of IL-1 $\beta$ . Inflammasomes can be induced to a variety of stimuli, including microbial components (eg, bacterial toxins) and environmental particulates (eg, silica and asbestos).<sup>18</sup>

Inflammasomes have been linked to carcinogenesis and the maintenance of a tumorigenic microenvironment.<sup>19,20</sup> It is likely that inflammasomes may play a critical role in the formation and progression of cancer through their contribution

**Table 1.** Cancers with identifiable changes in saliva.

TYPE OF CANCER	TYPE	SOURCE	BIOMARKER	REF
Oral	Protein	Saliva	CycD1, Ki67, LDH, MMP-9 OGG1, Maspin	58
	Protein	Saliva	IL-1 $\beta$ , IL-6, IL-8, Osteopontin	59
	Protein	Saliva	IL-8, IL-1 $\beta$ , M2BP	39
	Protein	Saliva	TNF- $\alpha$ , IL-1, IL-6, IL-8	10
	Protein	Saliva, Serum	IL-1 $\beta$ , IL-6, TNF- $\alpha$	21
Salivary gland	Protein	Saliva, Serum	FGF2, FGFR1	60
Head and neck	Protein	Serum	IL-6	61
	Protein	Serum, Tissue	IL-1 $\alpha$ , IL-6, IL-8, GM-CSF, VEGF, basic FGF	62
Ovarian	Protein	Tissue	c-erbB-2	63
Breast	mRNA, Protein	Saliva	CSTA, TPT1, IGF2BP1, GRM1, GRIK1, H6PD, MDM4, S100A8, CA6	64
	Protein	Saliva, Serum	CA15-3	15
	Protein	Saliva, Serum	c-erbB-2	14
	Protein	Serum	IL-6, IL-10	65
	Protein	Serum	IL-6	66
Lung	Protein	Saliva	EGFR, BRAF, CCNI, FGF19, FRS2, GREB1, LZTS1	17
	Protein	Saliva	HP, AZGP1, human calprotectin	67

to inflammation, immune responses, and tissue homeostasis. Although macrophages are the largest producers of IL-1 $\beta$ , other cell types from various tissue sources can also produce epidermal tissue, mucosa epithelial cells, acinar, and ductal cells of the salivary glands. However, limited work has been put into studying the levels of IL-1 $\beta$  and its relationship with different types of cancers. Given the growing number of evidences that cancer progression correlates with an increase in IL-1 $\beta$ ,<sup>21</sup> it is reasonable to suggest that IL-1 $\beta$  could be a potential biomarker as a predictor of cancer risk or cancer prognosis.

It is well known that polymorphisms of IL-1 receptor 1 (IL-1R1) and IL-1 $\beta$  have been associated with disease progression and prognosis.<sup>22,23</sup> IL-1 $\beta$  has been found to be elevated in various types of cancers, and it is known that IL-1 $\beta$  producing tumors have bad prognoses.<sup>4</sup> The IL-1 family of cytokines are a group of pleiotropic cytokines that can induce several genes that significantly promote cancer growth and metastasis, such as vascular endothelial growth factor (VEGF) and tumor growth factor (TGF)- $\beta$ . It is important to note that like many other proinflammatory cytokines, IL-1 $\beta$  exerts both beneficial and harmful effects in human beings. In cancer, IL-1 $\beta$  is the most clinically important IL-1 subtype and has attracted a lot of attention in the past decade, especially in the way this cytokine is activated. Therefore, tumors that express IL-1 $\beta$  must be proteolytically processed into its active form, via the inflammasome, before it can exert its physiological effects. IL-1 $\beta$  can induce the expression of adhesion molecules, increase prostaglandin production, and chemokine release.<sup>24</sup> All these contribute to cell chemotaxis, angiogenesis, and increase in cell adhesiveness, the typical hallmarks of cancer growth and spread.

There is growing evidence that inflammasomes may be playing a critical role in tumorigenesis. Inflammasomes play an active role in regulating proinflammatory signals and shaping the adaptive arm of the immune system (eg, T-cells). Therefore, it is no surprise that a dysregulation of inflammasome activity could influence disease outcomes and tumorigenesis. Cancer development can arise from various physiological stresses evoked by typical cancer risk factors such as viral infections, tobacco smoke, obesity, and aging, which results in inflammation. Once tumors arise, the maintenance of the tumor is critical for the cancer to spread and survive. It is suggested that inflammatory immune cells like macrophages and T-cells construct and orchestrate this favorable tumorigenic microenvironment by supplying cancer cells with cytokines, chemokines, and growth factors.

Despite the beneficial effects of inflammasome activation (as in the case of combating microbes), adverse effects of inflammasomes have been reported. Overt or excessive inflammasome activation could possibly drive and sustain tumorigenesis (eg, skin cancer). However, the loss of inflammasome activation could also increase the likelihood on cancer progression (eg, gastrointestinal cancer).

In gut-associated malignancies, loss of inflammasome components has been shown to increase tumor growth in several colitis mouse models.<sup>25–29</sup> This was shown to be attributed to the lack of IL-18 production, an important cytokine secreted in response to inflammasome activation. Normally, IL-18 functions as a factor to maintain tissue homeostasis and promote an antitumorigenic microenvironment.

In melanomas, the most malignant type of skin cancer, important inflammatory cytokines such as IL-6, IL-8, and IL-1 $\beta$  are upregulated. Interestingly, all these cytokines can be regulated by active IL-1 $\beta$ .<sup>30,31</sup> Elevated levels of IL-1 $\beta$

in melanomas can influence tumorigenesis and also increase the recruitment of immune suppressor cells to further reduce antitumor immune surveillance.<sup>32</sup> Indeed, loss of IL-1R and caspase 1 genes resulted in significantly reduced tumor growth and incidence in a chemical-induced skin cancer mouse model.<sup>33</sup> This suggests that there is a possible strong interplay between inflammasomes and skin cancer progression.

There is some evidence to show that inflammasome activation may play an important role in breast cancer. The crucial inflammasome mediator IL-1 $\beta$  plays an important role in the progression of breast cancer, although direct evidence linking to inflammasomes has yet to be established. However, there is a strong correlation between breast cancer tumor aggressiveness and IL-1 $\beta$  levels.<sup>34</sup> Work by Kurtzman et al<sup>35</sup> demonstrated that IL-1 $\beta$  expression is increased in 90% of invasive breast carcinomas and that it is localized to both tumor cells and stromal cells. Elevated IL-1 $\beta$  levels in breast cancer are associated with a more aggressive phenotype and higher tumor grade.<sup>36,37</sup> Furthermore, in a mouse model of human breast cancer, fibroblast growth factor (FGF) 1-induced mammary tumorigenesis is associated with local production of IL-1 $\beta$ .<sup>38</sup> Overall, it is possible that the levels of IL-1 $\beta$  might be related to different stages of breast cancer.

### Saliva: A Model Biological Fluid For Measuring IL-1 $\beta$

In the tumor microenvironment, both malignant and infiltrating immune cells secrete these cytokines, influencing tumor progression and eventual tumor dissemination. The levels of IL-1 $\beta$  have been shown to increase in oral types of cancers.<sup>21,39,40</sup> Elevated IL-1 $\beta$  levels are detected in the human lung, colon, breast carcinoma, and skin melanomas.<sup>4</sup> However, in all these studies, IL-1 $\beta$  have only been measured in the serum, not saliva. Out of all cancers studied, salivary IL-1 $\beta$  has only been measured in oral cancer patients (Table 1). Indeed, several groups have also measured IL-1 $\beta$  from saliva in other noncancer-related pathologies, including peri-implant inflammation in patients with dental implants,<sup>41</sup> periodontitis,<sup>42,43</sup> and graft-versus-host disease.<sup>44</sup> This suggests that saliva can be used as a diagnostic tool for detecting early stages of cancer.

Tumors are heterogeneous and consist of many cell types in addition to cancer cells, including cancer-associated fibroblasts, different tumor infiltrating immune cells, adipocytes, and endothelial cells to list a few.<sup>45,46</sup> All these cells have the ability to secrete immune-important cytokines and chemokines, which can directly affect cancer cells or cause cancer-associated inflammation. In malignancies where there is a low survival rate from time of diagnosis, high serum concentrations of IL-1 cytokines (ie, IL-1 $\beta$  and IL-1 $\alpha$ ) are usually present.<sup>4</sup> In general, tumors that produce IL-1 have bad prognoses and high IL-1 concentrations within the tumor microenvironment are usually associated with a more virulent tumor phenotype.<sup>47</sup>

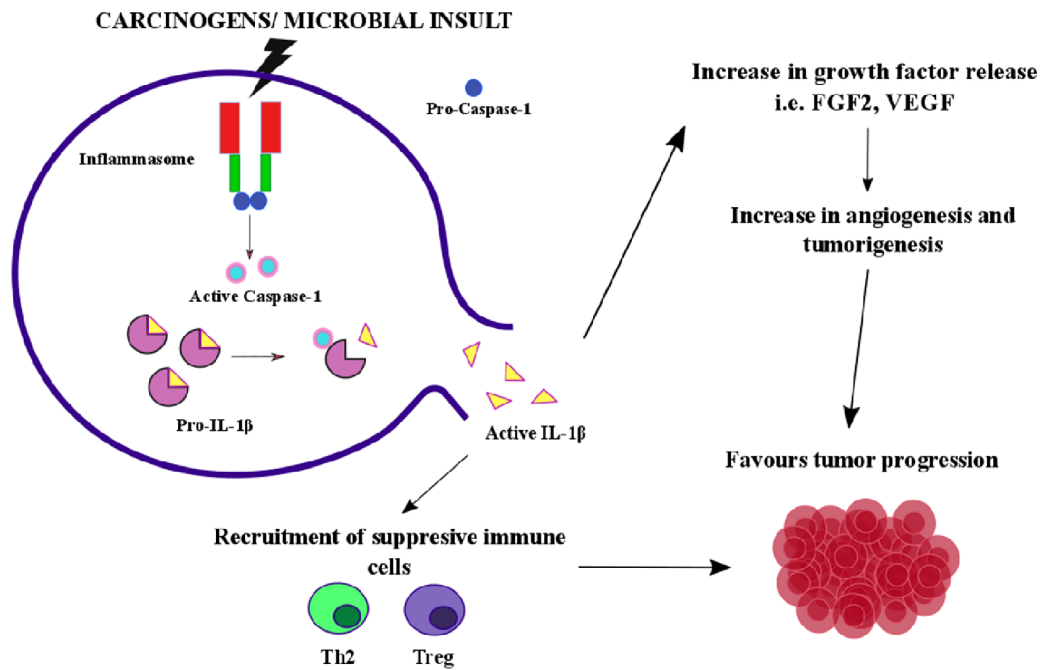
Researchers working with saliva biomarkers and diagnostics are always faced with the dilemma of whether the levels of target biomarker in saliva correlate with that in the blood circulation.

Some studied biomarkers show correlated changes between saliva and serum or plasma. For example, a positive correlation exists between salivary and serological levels of antigen CS 15-3 and the oncogene, c-erb, where it was shown to be significantly higher in breast cancer patients than in healthy controls.<sup>14,15,48</sup> On the other hand, biomarkers such as IL-8, a potential biomarker for oral squamous cell carcinoma (OSCC), could be detected at higher concentrations in saliva than serum.<sup>49</sup> However, the relationship between salivary and blood IL-1 $\beta$  is not clear. However, what is now clear is that circulating IL-1 $\beta$  in the blood of healthy individuals is very low and often below the detection limit,<sup>50</sup> whereas it is generally higher in saliva.<sup>21</sup> Resende et al<sup>44</sup> showed that salivary IL-1 $\beta$  levels increased progressively from the time before diagnosis until weeks after diagnosis, whereas blood IL-1 $\beta$  peak levels could be observed only within the time allotted for diagnosis in an acute graft-versus-host disease clinical study.

The other challenge in salivary biomarker research is the variability in the levels of potential salivary biomarkers in both healthy individuals and cancer patients. The best examples of this are the salivary biomarkers IL-6 and IL-8, which have been proposed as diagnostic markers for OSCC. Wide variation in the reference levels of these biomarkers have been reported in several different studies,<sup>51</sup> making it impossible to determine what ranges of salivary IL-6 or IL-8 levels are likely to indicate OSCC development. These wide variations in the levels of the same salivary constituent across different studies could be due at least partly to the different processing methods used or inherent biological variations within different individuals and groups. Although some degree of heterogeneity in the levels of IL-1 $\beta$  from healthy individuals have been reported, the variation is not as diverse as those reported for IL-6 and IL-8.<sup>21,52</sup> Furthermore, variation in salivary IL-1 $\beta$  levels from healthy individuals is only seen at different time periods of the day (ie, higher levels at awakening and lower during sleep).<sup>53</sup> This is not surprising as disruption in circadian system affects directly the production of inflammatory cytokines.<sup>54</sup> From a sample collection aspect, the timing of saliva collection is crucial to avoid such high background noise of salivary IL-1 $\beta$  levels to ensure that it does not mask the changes during disease progression, such as in cancer progression.

### Summary

IL-1 $\beta$  is well known as a potent promoter of carcinogenesis.<sup>55</sup> The potential molecular mechanisms by which IL-1 $\beta$  can promote tumor growth and invasion is depicted in Figure 1. However to date, there has been no detailed study investigating the levels of IL-1 $\beta$  at different stages of cancer, let alone in the salivary compartment. The use of salivary biomarkers has been explored for several types of cancers in the past including salivary gland cancer,<sup>56</sup> and cancers remote from the oral cavity such as breast and pancreatic cancer.<sup>57</sup> Furthermore, several studies have reported that salivary constituents can discriminate oral and systemic types of cancers (eg, lung cancer, breast



**Figure 1.** The inflammasome is a multicomplex system whose activation can be induced by exposure to carcinogens or intrinsic genetic aberrations, the basis of cancer initiation. Upon inflammasome activation, caspase 1 becomes activated, which leads to a cascade of proinflammatory events via the activation of cytokines, such as IL-1 $\beta$ , which then interact with their own membrane receptors amplifying the inflammatory response. On the other hand, active caspase 1 can lead to cell pyroptosis with the consequence of membrane rupture and release of various factors, including IL-1 $\beta$ . Inflammasome effectors (IL-1-like cytokines and pyroptosis) can influence the adaptive immune response and cell proliferative responses in different ways. The activation of the inflammasome complex can lead to the recruitment of suppressive immune cells, such as Type 2 helper T-cells (Th2) and regulatory T-cells (Treg), which can favor tumor progression. The inflammasome is also involved in tumor progression through the release of growth factors [eg, fibroblast growth factor 2 (FGF2) and vascular endothelial growth factor (VEGF)], which facilitate angiogenesis and tumor invasiveness.

cancer, pancreatic cancer, and ovarian cancer). We have briefly summarized the potential use of saliva as an alternative diagnostic fluid compared to blood. Although there have been many studies that investigated different types of cytokines and chemokines as potential biomarkers for early diagnosis and progression of cancer, limited amount of research has been done to investigate the correlation of IL-1 $\beta$  and cancer. Recent studies also suggest that the level of IL- $\beta$  is significantly higher in cancer patients compared to healthy controls, especially oral cancer. Furthermore, IL-1 $\beta$  is more detectable in saliva than in blood serum, suggesting that saliva serves as a good biological fluid for assessing changes in IL-1 $\beta$  levels.<sup>21</sup> Studying the dynamics of IL-1 $\beta$  changes in different cancers will shed light on linking the possible role of inflammasomes in cancer pathogenesis, a field that has gained attention among immunologists in the past decade. Identifying a cancer progression biomarker, especially in saliva, can hopefully predict the progress of the cancer and allow a more personalized anticancer treatment to be given to the patient. Current research has identified deregulated cytokines, such as IL-1 $\beta$ , in some cancers mentioned above. Robust and reproducible methods for the assessment of IL-1 $\beta$  in saliva, and the possibility of a rapid salivary test as an indicator of disease and risk of malignancy are important to qualify IL-1 $\beta$  as a putative cancer progression salivary biomarker. Ongoing development

of such a method will have profound impact on cancer screening and the early diagnosis of cancers, potentially resulting in early treatment and a decrease in the high levels of morbidity and mortality associated with different types of cancers.

### Author Contributions

Prepared the first draft of the manuscript: AI. Contributed to the writing of the manuscript: AI, NBG, and DK. Jointly developed the structure and arguments for the paper: AI, NBG, and DK. Made critical revisions and approved the final version: AI and DK. All the authors reviewed and approved the final manuscript.

### REFERENCES

1. Spielmann N, Wong DT. Saliva: diagnostics and therapeutic perspectives. *Oral Dis.* 2011;17(4):345–354.
2. Tiwari M. Science behind human saliva. *J Nat Sci Biol Med.* 2011;2(1):53–58.
3. Zimmermann BG, Park NJ, Wong DT. Genomic targets in saliva. *Ann NY Acad Sci.* 2007;1098:184–191.
4. Dinarello CA. Why not treat human cancer with interleukin-1 blockade? *Cancer Metastasis Rev.* 2010;29(2):317–329.
5. Shah FD, Begum R, Vajaria BN, et al. A review on salivary genomics and proteomics biomarkers in oral cancer. *Indian J Clin Biochem.* 2011;26(4):326–334.
6. Cuevas-Cordoba B, Santiago-Garcia J. Saliva: a fluid of study for OMICS. *OMICS.* 2014;18(2):87–97.
7. Cheng YS, Jordan L, Rees T, et al. Levels of potential oral cancer salivary mRNA biomarkers in oral cancer patients in remission and oral lichen planus patients. *Clin Oral Investig.* 2014;18(3):985–993.



8. Li Y, St John MA, Zhou X, et al. Salivary transcriptome diagnostics for oral cancer detection. *Clin Cancer Res*. 2004;10(24):8442–8450.
9. Malamud D. Saliva as a diagnostic fluid. *Dent Clin North Am*. 2011;55(1):159–178.
10. Pfaffe T, Cooper-White J, Beyerlein P, Kostner K, Punyadeera C. Diagnostic potential of saliva: current state and future applications. *Clin Chem*. 2011;57(5):675–687.
11. Ovchinnikov DA, Cooper MA, Pandit P, et al. Tumor-suppressor gene promoter hypermethylation in saliva of head and neck cancer patients. *Transl Oncol*. 2012;5(5):321–326.
12. Lee YH, Kim JH, Zhou H, Kim BW, Wong DT. Salivary transcriptomic biomarkers for detection of ovarian cancer: for serous papillary adenocarcinoma. *J Mol Med*. 2012;90(4):427–434.
13. Chen DX, Li FQ. [Primary research on saliva and serum CA125 assays for detecting malignant ovarian tumors]. *Zhonghua Fu Chan Ke Za Zhi*. 1990;25(2): 84–85, 123–124.
14. Streckfus C, Bigler L, Dellinger T, Dai X, Kingman A, Thigpen JT. The presence of soluble c-erbB-2 in saliva and serum among women with breast carcinoma: a preliminary study. *Clin Cancer Res*. 2000;6(6):2363–2370.
15. Agha-Hosseini F, Mirzaei-Dizgah I, Rahimi A, Seilanian-Toosi M. Correlation of serum and salivary CA125 levels in patients with breast cancer. *J Contemp Dent Pract*. 2009;10(6):E001–E008.
16. Wu ZZ, Wang JG, Zhang XL. Diagnostic model of saliva protein finger print analysis of patients with gastric cancer. *World J Gastroenterol*. 2009;15(7):865–870.
17. Zhang L, Xiao H, Zhou H, et al. Development of transcriptomic biomarker signature in human saliva to detect lung cancer. *Cell Mol Life Sci*. 2012;69(19):3341–3350.
18. Lamkanfi M, Dixit VM. Mechanisms and functions of inflammasomes. *Cell*. 2014;157(5):1013–1022.
19. Kolb R, Liu GH, Janowski AM, Sutterwala FS, Zhang W. Inflammasomes in cancer: a double-edged sword. *Protein Cell*. 2014;5(1):12–20.
20. Zitvogel L, Kepp O, Galluzzi L, Kroemer G. Inflammasomes in carcinogenesis and anticancer immune responses. *Nat Immunol*. 2012;13(4):343–351.
21. Brailo V, Vucicevic-Boras V, Lukac J, et al. Salivary and serum interleukin 1 beta, interleukin 6 and tumor necrosis factor alpha in patients with leukoplakia and oral cancer. *Med Oral Patol Oral Cir Bucal*. 2012;17(1):e10–e15.
22. Snoussi K, Strosberg AD, Bouaouina N, Ben Ahmed S, Chouchane L. Genetic variation in pro-inflammatory cytokines (interleukin-1beta, interleukin-1alpha and interleukin-6) associated with the aggressive forms, survival, and relapse prediction of breast carcinoma. *Eur Cytokine Netw*. 2005;16(4):253–260.
23. Liu X, Wang Z, Yu J, Lei G, Wang S. Three polymorphisms in interleukin-1beta gene and risk for breast cancer: a meta-analysis. *Breast Cancer Res Treat*. 2010;124(3):821–825.
24. Joosten LA, Netea MG, Dinarello CA. Interleukin-1beta in innate inflammation, autophagy and immunity. *Semin Immunol*. 2013;25(6):416–424.
25. Allen IC, TeKippe EM, Woodford RM, et al. The NLRP3 inflammasome functions as a negative regulator of tumorigenesis during colitis-associated cancer. *J Exp Med*. 2010;207(5):1045–1056.
26. Zaki MH, Vogel P, Body-Malapel M, Lamkanfi M, Kanneganti TD. IL-18 production downstream of the Nlrp3 inflammasome confers protection against colorectal tumor formation. *J Immunol*. 2010;185(8):4912–4920.
27. Dupaul-Chicoine J, Yeretssian G, Doiron K, et al. Control of intestinal homeostasis, colitis, and colitis-associated colorectal cancer by the inflammatory caspases. *Immunity*. 2010;32(3):367–378.
28. Zaki MH, Boyd KL, Vogel P, Kastan MB, Lamkanfi M, Kanneganti TD. The NLRP3 inflammasome protects against loss of epithelial integrity and mortality during experimental colitis. *Immunity*. 2010;32(3):379–391.
29. Salcedo R, Worschech A, Cardone M, et al. MyD88-mediated signaling prevents development of adenocarcinomas of the colon: role of interleukin 18. *J Exp Med*. 2010;207(8):1625–1636.
30. Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. *Annu Rev Immunol*. 2009;27:519–550.
31. Raman D, Baugher PJ, Thu YM, Richmond A. Role of chemokines in tumor growth. *Cancer Lett*. 2007;256(2):137–165.
32. Dunn JH, Ellis LZ, Fujita M. Inflammasomes as molecular mediators of inflammation and cancer: potential role in melanoma. *Cancer Lett*. 2012;314(1): 24–33.
33. Drexler SK, Bonsignore L, Masin M, et al. Tissue-specific opposing functions of the inflammasome adaptor ASC in the regulation of epithelial skin carcinogenesis. *Proc Natl Acad Sci U S A*. 2012;109(45):18384–18389.
34. Pantschenko AG, Pushkar I, Anderson KH, et al. The interleukin-1 family of cytokines and receptors in human breast cancer: implications for tumor progression. *Int J Oncol*. 2003;23(2):269–284.
35. Kurtzman S, Anderson K, Wang Y, et al. Cytokines in human breast cancer: IL-1alpha and IL-1beta expression. *Oncol Rep*. 1999;6(1):65–135.
36. Chavey C, Bibeau F, Gourgou-Bourgade S, et al. Oestrogen receptor negative breast cancers exhibit high cytokine content. *Breast Cancer Res*. 2007;9(1):R15.
37. Jin L, Yuan RQ, Fuchs A, et al. Expression of interleukin-1beta in human breast carcinoma. *Cancer*. 1997;80(3):421–434.
38. Reed JR, Leon RP, Hall MK, Schwertfeger KL. Interleukin-1beta and fibroblast growth factor receptor 1 cooperate to induce cyclooxygenase-2 during early mammary tumorigenesis. *Breast Cancer Res*. 2009;11(2):R21.
39. Elashoff D, Zhou H, Reiss J, et al. Prevalidation of salivary biomarkers for oral cancer detection. *Cancer Epidemiol Biomarkers Prev*. 2012;21(4):664–672.
40. Kamatani T, Shiogama S, Yoshihama Y, Kondo S, Shiota T, Shintani S. Interleukin-1 beta in unstimulated whole saliva is a potential biomarker for oral squamous cell carcinoma. *Cytokine*. 2013;64(2):497–502.
41. Rocha FS, Jesus RN, Rocha FM, Moura CC, Zanetta-Barbosa D. Saliva versus peri-implant inflammation: quantification of IL-1beta in partially and totally edentulous patients. *J Oral Implantol*. 2014;40(2):169–173.
42. Sanchez GA, Miozza VA, Delgado A, Busch L. Salivary IL-1beta and PGE2 as biomarkers of periodontal status, before and after periodontal treatment. *J Clin Periodontol*. 2013;40(12):1112–1117.
43. Gumus P, Nizam N, Nalbantsoy A, Ozcaka O, Buduneli N. Saliva and serum levels of pentraxin-3 and interleukin-1beta in generalized aggressive or chronic periodontitis. *J Periodontol*. 2014;85(3):e40–e46.
44. Resende RG, Abreu MH, de Souza LN, Silva ME, Gomez RS, Correia-Silva Jde F. Association between IL1B (+3954) polymorphisms and IL-1beta levels in blood and saliva, together with acute graft-versus-host disease. *J Interferon Cytokine Res*. 2013;33(7):392–397.
45. Coussens LM, Zitvogel L, Palucka AK. Neutralizing tumor-promoting chronic inflammation: a magic bullet? *Science*. 2013;339(6117):286–291.
46. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell*. 2010;140(6):883–899.
47. Lewis AM, Varghese S, Xu H, Alexander HR. Interleukin-1 and cancer progression: the emerging role of interleukin-1 receptor antagonist as a novel therapeutic agent in cancer treatment. *J Transl Med*. 2006;4:48.
48. Streckfus C, Bigler L, Tucci M, Thigpen JT. A preliminary study of CA15-3, c-erbB-2, epidermal growth factor receptor, cathepsin-D, and p53 in saliva among women with breast carcinoma. *Cancer Invest*. 2000;18(2):101–109.
49. St John MA, Li Y, Zhou X, et al. Interleukin 6 and interleukin 8 as potential biomarkers for oral cavity and oropharyngeal squamous cell carcinoma. *Arch Otolaryngol Head Neck Surg*. 2004;130(8):929–935.
50. Wong H-L, Pfeiffer RM, Fears TR, Vermeulen R, Ji S, Rabkin CS. Reproducibility and correlations of multiplex cytokine levels in asymptomatic persons. *Cancer Epidemiol Biomarkers Prev*. 2008;17(12):3450–3456.
51. Cheng Y-SL, Rees T, Wright J. A review of research on salivary biomarkers for oral cancer detection. *Clin Transl Med*. 2014;3:3–3.
52. Thomas MV, Branscum A, Miller CS, Ebersole J, Al-Sabbagh M, Schuster JL. Within-subject variability in repeated measures of salivary analytes in healthy adults. *J Periodontol*. 2009;80(7):1146–1153.
53. Reinhardt EL, Fernandes PACM, Markus RP, Fischer FM. Daily rhythm of salivary IL-1 $\beta$ , cortisol and melatonin in day and night workers. *Work*. 2012;41: 5788–5790.
54. Castanon-Cervantes O, Wu M, Ehlen JC, et al. Dysregulation of inflammatory responses by chronic circadian disruption. *J Immunol*. 2010;185(10):5796–5805.
55. Charles A. Biologic basis for interleukin-1 in disease. *Blood*. 1996;87(6):2095–2147.
56. Stenner M, Klusmann JP. Current update on established and novel biomarkers in salivary gland carcinoma pathology and the molecular pathways involved. *Eur Arch Otorhinolaryngol*. 2009;266(3):333–341.
57. Bigler LR, Streckfus CF, Dubinsky WP. Salivary biomarkers for the detection of malignant tumors that are remote from the oral cavity. *Clin Lab Med*. 2009;29(1): 71–85.
58. Shpitzer T, Hamzany Y, Bahar G, et al. Salivary analysis of oral cancer biomarkers. *Br J Cancer*. 2009;101(7):1194–1198.
59. Katakura A, Kamiyama I, Takano N, et al. Comparison of salivary cytokine levels in oral cancer patients and healthy subjects. *Bull Tokyo Dent Coll*. 2007;48(4): 199–203.
60. Huang YQ, Li YD, Li GK, Jin Z, Ma J. The evaluation of basic fibroblast growth factor and fibroblastic growth factor receptor 1 levels in saliva and serum of patients with salivary gland tumor. *DNA Cell Biol*. 2012;31(4):520–523.
61. Duffy SA, Taylor JM, Terrell JE, et al. Interleukin-6 predicts recurrence and survival among head and neck cancer patients. *Cancer*. 2008;113(4):750–757.
62. Chen Z, Malhotra PS, Thomas GR, et al. Expression of proinflammatory and proangiogenic cytokines in patients with head and neck cancer. *Clin Cancer Res*. 1999;5(6):1369–1379.
63. Meden H, Marx D, Rath W, et al. Overexpression of the oncogene c-erb B2 in primary ovarian cancer: evaluation of the prognostic value in a Cox proportional hazards multiple regression. *Int J Gynecol Pathol*. 1994;13(1):45–53.
64. Zhang L, Xiao H, Karlan S, et al. Discovery and preclinical validation of salivary transcriptomic and proteomic biomarkers for the non-invasive detection of breast cancer. *PLoS One*. 2010;5(12):e15573.
65. Ikeguchi M, Hatada T, Yamamoto M, et al. Serum interleukin-6 and -10 levels in patients with gastric cancer. *Gastric Cancer*. 2009;12(2):95–100.
66. Wu CW, Wang SR, Chao MF, et al. Serum interleukin-6 levels reflect disease status of gastric cancer. *Am J Gastroenterol*. 1996;91(7):1417–1422.
67. Xiao H, Zhang L, Zhou H, Lee JM, Garon EB, Wong DT. Proteomic analysis of human saliva from lung cancer patients using two-dimensional difference gel electrophoresis and mass spectrometry. *Mol Cell Proteomics*. 2012;11(2): M111012112.