

Involvement of Immune Regulation in Multiple Sclerosis

Gabrielle Spagnuolo^{1,2}, Aaron Piavis² and Tyisha Williams^{1,2}

¹Department of Biology, Wilkes University, Wilkes-Barre, PA, USA. ²Neuroscience Program, Wilkes University, Wilkes-Barre, PA, USA.

Immunology and Immunogenetics Insights
Volume 9: 1–10
© The Author(s) 2017
Reprints and permissions:
sagepub.co.uk/journalsPermissions.nav
DOI: 10.1177/1178634517734175


ABSTRACT: Multiple sclerosis (MS) is a neurodegenerative disease characterized by neuroinflammation and demyelination that results in axon loss. Multiple sclerosis has been shown to be the result of an autoimmune response caused by a mixture of genetic and environmental factors. Dendritic cells are prominent antigen-presenting cells that interact with various molecules to regulate the immune system. The dysfunction of various features of immune regulation, including interleukins (ILs), CD4⁺ T cells, and suppressor of cytokine signaling (SOCS1), has been implicated in the pathogenesis of MS. T cells, particularly through the malfunction of B7-costimulatory pathways, have been shown to affect the progression of the disease. SOCS1 is important in regulating the function of T cells through its interactions with other nearby genes, especially CLEC16A, with abnormal decreases in SOCS1 expression leading to the exhibition of MS symptoms. The activation of IL-23 receptors on CD4⁺ T cells is pivotal to their differentiation into pathogenic T_H17 cells. Several promising compounds that downregulate gene expression of IL-23 and IL-23R have been discovered but require further investigation for efficacy and safety. Given their role in the severity and progression of MS, therapies that decrease these dysregulations may ultimately decrease symptoms and in turn improve patients' quality of life.

KEYWORDS: Multiple sclerosis, dendritic cells, T cells, interleukins, cytokines

RECEIVED: May 1, 2017. **ACCEPTED:** August 25, 2017.

PEER REVIEW: Three peer reviewers contributed to the peer review report. Reviewers' reports totaled 515 words, excluding any confidential comments to the academic editor.

TYPE: Review

FUNDING: The author(s) received no financial support for the research, authorship, and/or publication of this article.

DECLARATION OF CONFLICTING INTERESTS: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

CORRESPONDING AUTHOR: Tyisha Williams, Department of Biology, Wilkes University, 84 West South Street, Wilkes-Barre, PA 18766, USA. Email: tyisha.williams@wilkes.edu

Introduction

Multiple sclerosis (MS) is a neurodegenerative disease characterized by inflammation of the central nervous system (CNS). It has been reported that in MS, there is an autoimmune response that leads to myelin deterioration and axonal loss, but the specific cause of the disease has yet to be determined.¹ One widely studied possible cause of MS is a triggered response to the Epstein-Barr virus (EBV).^{2,3} It has been speculated that the viral infection results in the upregulation of antigen-specific cytotoxic T cells, which remain throughout the body for several years postinfection.² When these cytotoxic T cells target self-peptides in the CNS, this often results in autoimmunity, such as MS. Although this link has been implicated in literature, conclusive evidence supporting this as the unique cause of MS is currently lacking.^{4,5}

Although EBV may play a key role in disease onset, multiple lines of evidence suggest a genetic component to the onset and severity of the disease. Although the role of heredity is yet to be fully understood, variations in specific genes, especially those relating to inflammation, have been associated with an increased risk of MS and corresponds well with the increased inflammatory response observed in the advancement of MS.^{6–9} This is further supported by the increased incidence of MS in those with dysregulation of T-cell differentiation, particularly effector T cells.^{7,9}

Naïve CD4⁺ T cells can mature into different effector T-cell types including helper T (T_H) (T_H1, T_H2, and T_H17) and regulatory T (Treg) cells.¹⁰ IFN- γ -secreting T_H1 cells and interleukin (IL) 17-secreting T_H17 cells are present in MS lesions in the CNS and are known to be integral in the

pathogenesis of MS in human and mouse models.^{11–14} Conversely, addition of Tregs in experimental autoimmune encephalomyelitis (EAE), a mouse model simulating MS, shows a tolerogenic effect, whereas depletion of Tregs increases symptom severity.¹⁵ This differentiation of T_H cells is initiated by the interactions between naïve CD4⁺ T-cell populations and antigen-presenting cells (APCs), primarily dendritic cells (DCs).¹⁶ This communication involves the B7/T-cell costimulatory pathway and the stimulation of T cells with cytokines produced by DCs. Under normal circumstances, peripheral DCs in an immature state do not initiate T_H cell differentiation and show very low expression of costimulatory molecules and cytokines.^{17,18} On receiving signals classified as pathogenic, immature DCs become activated and mature. This process is characterized by increased antigen uptake, increased class II major histocompatibility complex (MHCII) molecule expression, increased expression of costimulatory molecules (such as B7 molecules), and increased secretion of cytokines.¹⁹ Different cytokines secreted by mature DCs are responsible for manifestation of the different subsets of T_H cells. T_H17 differentiation occurs via IL-6, IL-23, and IL-17 signaling; T_H1 via IL-12 signaling, whereas Treg development occurs through interactions with IL-10.^{20–23}

The notion that DCs are of critical importance in the pathogenesis of MS is well established. For instance, high amounts of CD8 α ⁺-type conventional DCs (cDCs) and plasmacytoid DCs (pDCs) are seen in the white matter and cerebrospinal fluid (CSF) of MS subjects.^{24,25} Patients with MS have been shown to have higher amounts of cDCs that secrete



Table 1. Important molecules dysregulated in MS pathogenesis.

BIOLOGICAL PROCESS	INVOLVEMENT WITH MS
IL-23	Cytokine that facilitates the development of inflammation by regulating immune system cell activity Increased amounts promote pathogenic T _H 17 cell survival in mice and humans
IL-23R	Receptor that results in chemical signals that coordinate immune response on activation Responsible for specialization of T _H 17 cells from T _H 1 cells that is believed to lead to MS pathology
CTLA-4	Immune checkpoint that downregulates immune response when bound by B7 molecules
CD28	Immune checkpoint that upregulates immune response when bound by B7 molecules
PD-1/PD-L1	Once bound to one another, suppresses T-cell proliferation to reduce inflammation Dysregulation believed to result in autoimmunity
SOCS1	Regulates cytokine signaling and inflammation Single-nucleotide polymorphisms may be responsible for specialization of T _H 17 cells from T cells that can lead to MS pathology

Abbreviation: MS, multiple sclerosis.

This table illustrates the important molecules and pathways involved in immune regulation that play a role in the induction of MS on dysregulation.

IL-12 and express CD80, both of which are markers associated with the differentiation of the pathogenic T_H1 population.²⁶ Furthermore, cDCs acquired from patients with MS show an increased secretion of proinflammatory cytokines involved in MS pathogenesis, such as IL-6 and IL-23, compared with healthy individuals.^{27,28} It has been observed that populations of cDCs can infiltrate the CNS and facilitate epitope spreading in EAE mouse models.²⁹ As DCs interact with and are involved in different aspects of immune regulation that have been shown to increase risk of MS onset, the focus of this review is to highlight the role of immune regulation in the development and progression of MS, particularly regarding DCs, the associations of DCs and the B7/T-cell costimulatory pathway, as well as the resulting cytokine release and functions associated with T cells (see Table 1).

Methods

A literature review was conducted using the database *Web of Science* with a combination of the key word “multiple sclerosis” with “dendritic cells,” “interleukins,” “cytokines,” and “costimulatory pathway.” From the 186 articles generated, those the authors did not have access to (33 articles) were excluded, resulting in the inclusion of 153 total articles considered for this review.

DCs and MS

Dendritic cells can be divided into 5 subsets that exist in mice and humans: cDCs, CD11b⁺-type cDCs, pDCs, Langerhans cells, and monocyte-derived DCs (MoDCs).^{30–37} Of these 5 subsets, pDCs and cDCs are of hematopoietic origin and stem from a DC progenitor cell that matures in response to FLT-3 signaling.³⁸ It has been shown that cDCs and pDCs are present in high amounts in white matter lesions of patients with MS,^{24,39} suggesting that they may play a role in the disease. Conventional DCs promote cytotoxic T-cell activation and primarily function to induce adaptive immune responses that

target tumors and intracellular pathogens.^{30,38} CD11b⁺-type cDCs are responsible for CD4⁺ T-cell activation and promote humoral immunity, particularly regarding antibody production and cytokine secretion.³⁸

There is evidence supporting the involvement of the various subsets of DCs in the pathogenesis of MS. For example, in EAE, cDCs alone are able to sufficiently present antigen, activate autoreactive T cells, and facilitate CNS inflammation.^{29,40,41} All of these symptoms are significant, as they are each key features of EAE development and severity, which in turn would translate to MS severity in humans. It has also been demonstrated that pDCs have the ability to influence the differentiation of immunosuppressive Tregs that monitor self-tolerance to prevent autoimmunity.⁴² In addition, increased pDCs have been observed in the inflamed regions of the CSF of patients with MS when compared with healthy controls,^{25,43} indicating that the upregulated immunosuppression demonstrated by the overexpression of pDCs exacerbates MS symptoms. This has been further supported by a recent study showing a greater number of pDCs in the CSF of patients in relapse compared with those who were in remission.²⁵ Together, this indicates that the influence of overexpressed pDCs on Tregs results in the suppression of the typical mechanism that monitors self-tolerance, contributing to autoimmunity.

On infection or inflammation, monocyte precursors are activated into MoDCs through granulocyte-macrophage colony-stimulating factor (GM-CSF), which shares characteristic qualities with hematopoietic-derived DCs.⁴⁴ Inflammatory monocytes extracted from GM-CSF-deficient mice are unable to develop into MoDCs, which indicates that GM-CSF is required for the generation of MoDCs.⁴⁵ Co-culturing these MoDCs with naïve CD4⁺ T cells resulted in the in vitro differentiation of T_H17 cells.³⁷ This indicates that the interactions between these activated MoDCs and T cells result in the differentiation of T cells into T_H17 cells, contributing to demyelination and inflammation at MS lesions. Consistent with the

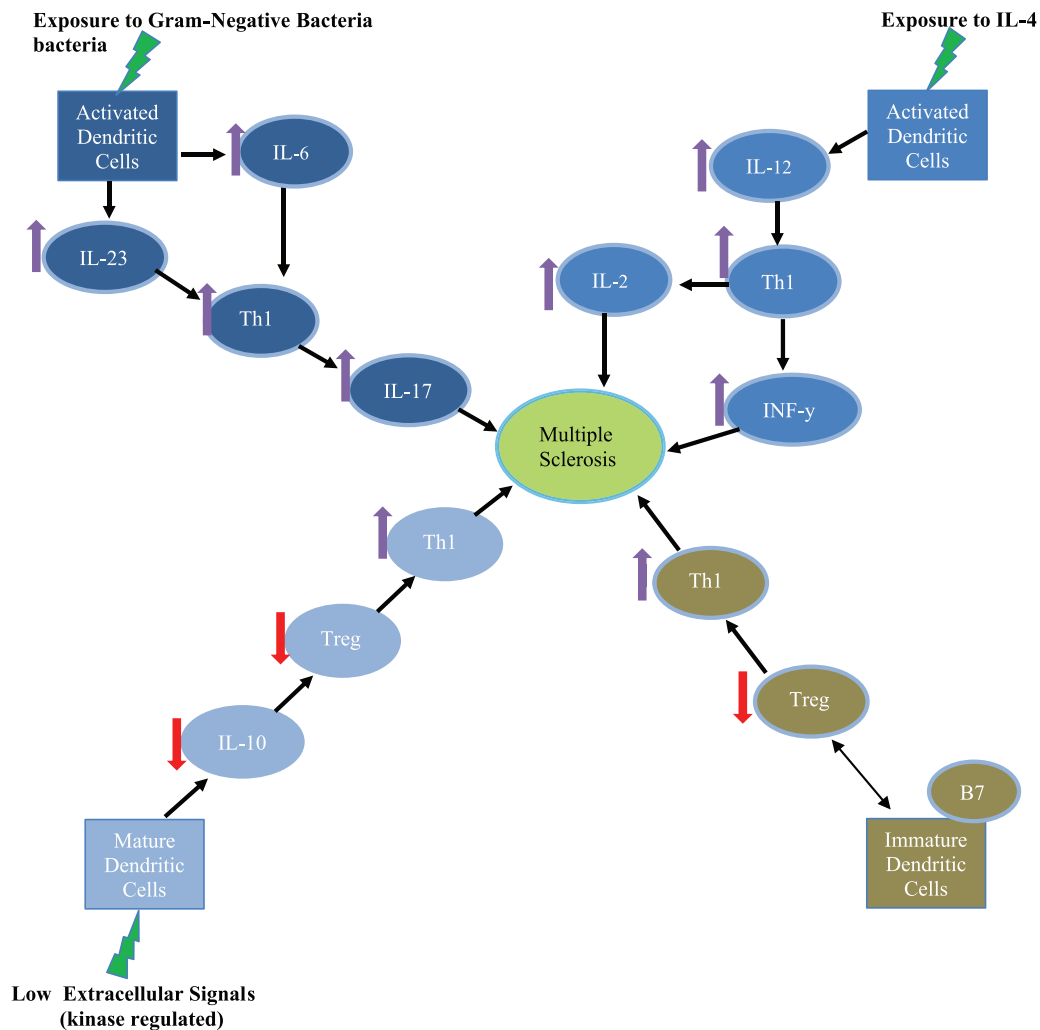


Figure 1. The pathogenesis of MS involves the dysfunction of immune regulation. Proper immune regulation depends on a delicate balance among the various immune responsive factors, which first requires their release from dendritic cells. Furthermore, the characteristics of dendritic cells (eg, maturity) will determine which of the various inflammatory cytokines (predominately interleukins) will be secreted. These cytokines will interact with naïve helper T cells at their surface and will promote either induction of or tolerance to autoimmunity. This figure illustrates pathways that have been implicated in the pathogenesis of MS on their dysfunction, as is highlighted (red arrows = downregulation, purple arrows = upregulation, green lightning bolt = trigger). IL indicates interleukin; MS, multiple sclerosis; T_H, helper T; Tregs, regulatory T cells.

observed dysregulation of the IL-23/IL-17 axis in MS, MoDCs in the blood of patients with MS show increases in IL-23 synthesis in addition to increased ability to induce T_H17 differentiation.²⁸ Based on this evidence, there is a correlation between DCs and MS. Dendritic cells interact with other molecules and when these communications are impaired, the result is the misregulation of various immune control mechanisms and the secretion of cytokines that lead to CNS inflammation and the progression of MS symptoms, as will be discussed below (see Figure 1).

IL-23 and MS

Previous work has established that IL-23 is one of the key molecules responsible for maintaining the activity of T_H17 lymphocytes.^{22,46,47} While transforming growth factor β and IL-6 are also involved in the differentiation of T_H17 cells, stimulation with these cytokines alone is not enough to generate T_H17

cells with effective pathogenicity.²² Rather, it is IL-23 that is mainly released from DCs, subsequently acting on IL-23 receptors (IL-23R) found on naïve CD4⁺ T cells, inducing their specialization into pathogenic T_H17 cells.²⁸ In EAE, the presence of IL-23 has been shown to increase IL-17 levels, which robustly enhances the manifestation of symptoms.⁷ Moreover, IL-23 has also been found to be upregulated in the serum of patients with MS.^{48,49}

It was previously hypothesized that MS, along with several other inflammatory autoimmune diseases, was completely facilitated by T_H1 activation mediated by IL-12. The hypothesis that IL-12-mediated T_H1 cells are the sole cause of autoimmune pathogenesis originates, at least in part, due to early studies focusing on the IL-12p40 subunit. For instance, it was observed in early studies that the knockout of IL-12p40 ameliorated the symptoms of MS in various models.^{50,51} This was accepted until several studies demonstrated that T_H17

lymphocytes served a key role in the pathogenesis of several autoimmune diseases including MS.^{7,52-57} It is now known that IL-23 shares the IL-12p40 subunit with IL-12p70, with IL-12p40 bound to IL-23p19 rather than IL-12p35.⁵⁸ Furthermore, it has recently been shown that IL-23 may play a more important role than IL-12 in MS and other autoimmune diseases.^{59,60} For instance, removal of IL-23p19 was sufficient to cause resistance to EAE onset.^{52,60} Similarly, literature also shows that mice having dysfunction of several parts of the IL-12- T_H1 -IFN- γ pathway were still susceptible to EAE.^{50,51,61}

Despite research supporting that IL-23 serves a greater role in autoimmunity than IL-12, studies continue to focus on the shared IL-12p40 subunit. For example, a study focusing on the potential therapeutic effects of Trabad downregulation of histone demethylase Jmjd2d discussed how this should result in suppression of IL-12p40 levels.⁶² However, this broader T_H1 / T_H17 focus could fall out of favor if more IL-23p19-specific therapeutic approaches to MS are discovered. A potential shift toward IL-23p19-specific therapeutic approaches is possible as research suggests that therapeutically targeting both T_H1 and T_H17 subsets results in unwanted side effects. In humans, it has been shown that autosomal recessive deficiencies in IL-12p40 and IL-12 β 1 subunits results in mendelian susceptibility to mycobacterial disease.⁶³⁻⁶⁸ Likewise, IL-12-deficient mice are susceptible to *Listeria*, *Mycobacterium*, *Leishmania*, and *Toxoplasma*, whereas IL-23-deficient mice are very resistant to these same pathogens.^{69,70} In addition, a recent study comparing the effects of anti-IL-12p40 therapy and anti-IL-23p19 therapy showed that both were effective in reducing EAE symptoms. Anti-IL-12p40 therapy caused several side effects associated with the inhibition of both T_H1 and T_H17 functions, including the paradoxical pathogenic role of decreased IFN- γ levels.⁷¹ The effects of anti-IL-12p40 therapies on patients with MS were observed in phase 2 clinical trials. However, the beneficial effects of targeting this subunit were not determined to be sufficient enough to warrant further development.^{72,73} Unfortunately, within the context of MS, there has also been an issue of clarity with IL-23p19-specific therapies. Interestingly, despite observing therapeutic success using antibodies that target IL-23p19 in other autoimmune diseases and EAE,^{71,74,75} there have been no such clinical trials using this approach in patients with MS to date. Such anti-IL-23p19 therapeutic options include tildrakizumab and guselkumab. Both of these monoclonal antibodies (mAbs) have specifically shown to be promising treatments for plaque psoriasis.^{76,77} Further studies focusing on these antibodies within the context of MS could prove to be a valuable asset to progressing treatment options.

Although there is an interest to use antibodies, it is important to note that even direct antagonism of IL-23p19 with mAbs comes with certain issues that need to be circumvented. Almost all protein-based therapeutic approaches produce neutralizing antibodies, causing a reduction in efficacy.⁷⁸⁻⁸³ For example, research observing the effects of repeated therapeutic

administration of the mAb, natalizumab, shows loss of efficacy over time due to the body developing neutralizing antibodies toward mAbs.^{84,85} One approach used to avoid the issues associated with mAbs is RNA interference (RNAi), specifically RNAi of IL-23 expression within pathogenic APCs. RNA interference is a method of silencing gene expression posttranscriptionally.⁸⁶ In 2006, Vaknin-Dembinsky et al observed that MoDCs from patients with MS transfected with antisense oligos containing a specific complementary RNA sequence to IL-23 decreased the amount of biologically active IL-23. In addition, pathogenic CD4⁺ T cells of patients with MS cultured with the media from antisense-transfected DCs produced less IL-17.²⁸ Antisense oligonucleotides have recently been used in studies to target the messenger RNA (mRNA) transcripts of proteins believed to be important in autoimmune diseases such as Crohn disease.^{87,88} Moreover, a study conducted by Kalantari et al explored the effects of IL-23 knockdown by RNAi in bone marrow-derived DCs (BMDCs), one of the main contributors to IL-23 production and T_H17 differentiation in autoimmune diseases. The BMDCs of EAE mice were extracted and transduced with lentiviral vectors engineered to express small hairpin RNA (shRNA) that complemented sequences of IL-23p19 RNA as well as CD40 RNA. This eliminated the barrier of developing an administration technique that was practical in a clinical setting. When transduced, BMDCs are co-cultured with CD4⁺ T cells of EAE mice, and a decrease in T-cell proliferation to the pathogenic subset can be observed.⁸⁹ In addition, IL-23p19-specific shRNA transduction appears to aid in the maturation of BMDCs toward the tolerogenic subset, increasing IL-10 production, whereas the production of IL-6, IL-12, and IFN- γ is greatly reduced.⁹⁰ Unfortunately, despite the observed success of IL-23-specific antisense treatment, there have not been further studies or any clinical trials that test this specific therapeutic approach. The next step in RNAi therapies targeting IL-23 would be further studies replicating the lentiviral vector approach to better understand the potential effectiveness and pitfalls of implementing this as a clinical treatment.

As another alternative to developing antibodies that target the IL-23p19 subunit, associated adenovirus (AAV) vector containing a soluble recombinant IL-23 receptor (sIL-23R) homology region was developed.⁹¹ Expression of this inactive and soluble form of IL-23R requires one injection when administered via an AAV8 vector, thus avoiding continuous administration. The injection of sIL-23R-containing AAV8 into mice improved EAE clinical symptoms by blocking the IL-23/IL-23R interaction via competitive inhibition. Specifically, AAV8 administration of sIL-23R was shown to delay clinical onset, slow disease progression, reduce inflammatory infiltration, and reduce demyelination in the CNS.⁹² From previous studies revealing that AAV8 vectors only function in the periphery, it was presumed that the inhibitory effects of sIL-23R did not occur in the CNS, but rather

inhibition of IL-23 signaling in the periphery allowed less IL-23 to enter the CNS.^{93,94}

However, several questions have been raised that have yet to be answered from these recent findings. In particular, the effects that IL-23R has on inflammatory cytokine transcription factors must be further explored. In mice and humans, sIL-23R administration was found to decrease T_H17 differentiation and STAT3 phosphorylation.^{92,95} STAT3 of the JAK-STAT pathway is an important transcription factor downstream of IL-23R and IL-6R signaling.^{96,97} In the initial stages of T_H17 development, solely IL-6R signaling results in the phosphorylation of STAT3.^{96,97} Phosphorylation of STAT3 results in its association with a transcription factor complex that directly controls the genes for IL-17A, IL-17F, and IL-23R. The upregulation of IL-23R then enhances IL-23R-mediated STAT3 phosphorylation in a positive feedback manner, stabilizing the production of IL-17 in pathogenic T_H17 cells.^{96,97} Contrary to these observations in the IL-23R-STAT3 pathway, gene expressions induced by STAT3 activation (such as *IL-17A*, *IL-17F*, and *IL-23R*) were not decreased in media of myelin oligodendrocyte glycoprotein (MOG₄₀₋₅₅)-cultured splenocytes from EAE mice following sIL-23R treatment compared with untreated MOG₄₀₋₅₅ culture media. Instead, an increase in IFN- γ expression correlated with amelioration of EAE symptoms was observed.⁹² The lack of change in cytokine levels suggests that a transcription factor complex regulates the fate of pathogenic T_H17 commitment rather than single independent transcription factors.⁹⁸ Regarding the increase in the expression of IFN- γ in sIL-23R-treated mice, the accompanying reduction in EAE symptoms that was observed is consistent with current literature suggesting that IFN- γ is a negative regulator of IL-23/IL-17 immune responses.⁹⁹⁻¹⁰⁴ However, there is also literature reporting positive associations with CNS lesions in EAE as well as MS and higher levels of IFN- γ , suggesting a pathogenic role.^{22,50,105-107} To date, the specific role of IFN- γ in the pathogenicity of MS seems paradoxical as the role of IFN- γ changes depending on the stage of the disease.¹⁰⁸⁻¹¹⁰ Because there are no significant changes seen in cytokine levels except IFN- γ , it is possible that inhibition of the IL-23/IL-23R interaction promotes a compensatory mechanism whereby IFN- γ increases. It is also possible that replication of the experiment that Miralles et al⁹² performed could yield changes in cytokine levels. Seeing as this particular therapeutic approach to the negative regulation of T_H17 cells is rather new, further clarification of the exact mechanism of action of sIL-23R on the amelioration of EAE symptoms is warranted.

Associations between MS and the B7/T-Cell Costimulatory Pathway

T-cell costimulatory pathways, particularly the pathway involving B7 molecules (B7-1/CD80 and B7-2/CD86) and the protein receptors they act on (CTLA-4 and CD28), have been implicated in the pathogenesis of MS,^{111,112} but their role in

autoimmune diseases has been highly contested within the literature. A few recent studies have suggested that when faulty, the signals involved in the B7-CD28/CTLA-4 costimulatory pathway can trigger the activation of autoreactive T cells that contribute to the development of MS through the alteration of Treg activity and the induction of pathogenic T_H17 cells.^{113,114} To become activated, T cells require 2 signals,^{115,116} the first of which is antigen-specific due to the communication between the T-cell receptor (TCR) and an antigenic peptide, which is presented by an APC on an MHC molecule. The primary APCs that function in relation to this pathway are DCs, as mentioned previously. The second signal, which directly determines the functional response of the T cells to the TCR-MHC binding, occurs through the interaction of the CTLA-4 and CD28 receptors and the B7 molecules that are already expressed in low levels on DCs.^{111,117} Alternatively, binding of CTLA-4 to the B7 molecules inhibits an immune response by downregulating the costimulatory activation through the prevention of CD28 binding and reducing the antigen-presenting ability of APCs, which diminishes the triggering of T-cell activation.¹¹⁸⁻¹²¹ The balance of T-cell activation is preserved by the immunomodulation activities of DCs in promoting immunity and/or tolerance.¹²² When DCs are dysfunctional, it can lead to aberrant T-cell activation that prevents the body from properly reacting to both threats and benign targets.

Mouse models strengthen the association between the T-cell costimulatory pathway and MS. Using the EAE model, mice were treated with a combination of CTLA-4-Ig (or abatacept, a B7 antagonist that binds to B7 molecules to prevent T-cell stimulation) and MOG₃₅₋₅₅. Following treatment, EAE was exacerbated with more pronounced effects occurring in vivo than in vitro.⁸ It has been shown that ipilimumab, a drug used to treat metastatic melanoma through the inhibition of CTLA-4, and subsequently the downregulation of B7 molecules, mediates the development of MS based on an observed increase in disease progression through clinical and magnetic resonance imaging activity.¹¹⁸ This information, in conjunction with 2 other case studies that showed similar effects in patients undergoing treatment with ipilimumab, suggests that CTLA-4 serves a protective role against MS through its typical inhibitory activity in T cells, which is lost once CTLA-4 is downregulated.^{123,124} CTLA-4 is constitutively expressed on Tregs and binding to the B7 molecules present on the surface of DCs has previously been shown to upregulate CTLA-4 once T cells are activated, which suppresses immune activation.¹²⁵ When CTLA-4 is downregulated, it has been shown to lose its ability to inhibit Tregs, which causes these cells to become overactive and results in the dysfunction of effector T cells, which in turn can promote autoimmunity.¹²⁶ When CTLA-4 is blocked, it has been shown to induce differentiation of naïve T cells into T_H17 cells, which have been implicated in the pathogenesis of MS.¹²⁷ Other studies have recently challenged the position that CTLA-4 has a critical role in the onset or progression of

MS. In a recent study, patients treated with abatacept, a B7 antagonist, did not see a therapeutic effect.¹²⁸ These results are in direct contrast to previous findings suggesting abatacept is a potential therapeutic regulator of neuroinflammation.^{129,130} These contradictory findings may be explained by low disease activity prior to treatment.

Furthermore, specific polymorphisms of CTLA-4 have been reported by some groups to have varying effects on risk of MS, particularly the +49 A/G*G-CT60*G haplotype. One study using a Flemish cohort found a significant increase in the +49 G allele in patients with MS compared with their control group.¹²⁰ In contrast, studies performed using Italian, Japanese, Iranian, German, Hungarian, and Polish cohorts were unable to conclude that there was an association between CTLA-4 polymorphisms and MS in their patients.^{131–134} It should be noted that the sample sizes for some of these studies were small. Based on the differing groups in which a connection between CTLA-4 and MS was observed, the association is perhaps indicative of an ethnic component, which requires further investigation to elucidate the true nature of the relationship.

PD-1/PD-L1 Association with MS

Programmed death-ligand 1 (PD-L1 or B7-H1) is another member of the B7-costimulatory pathway that has been linked to MS. PD-L1 is constitutively expressed on DCs and binds to the programmed death receptor (PD-1) on activated T cells, which then upregulates the production of inflammatory cytokines, such as IL-10. It also reduces T-cell proliferation by suppressing T_H1 cell differentiation and DC function.¹³⁵ This pathway is important in subduing T-cell activity, known as T-cell exhaustion, when a virus or other abnormality is present in the CNS to reduce neuroinflammation.¹³⁶ Once the T cell is exhausted, it is only weakly activated, which leaves PD-1 better capable of limiting the T-cell response.¹³⁷ PD-1 is also vital in regulating tolerance and autoimmunity. PD-L1 is upregulated in MS lesions, which results in the downregulation of T-cell response and the prevention of autoimmunity.^{135,138} One hypothesized role of PD-1 is a suppressive effect on APCs, particularly DCs, that infiltrate the CNS, which results in the downregulation of T_H17 cells.¹³⁹ In downregulating this type of pathogenic T cell, PD-1 serves as a modulator of immunity, which in turn reinforces its role in preventing the occurrence of disease.

One important aspect of PD-L1/PD-1 is regarding prominent MS treatments, particularly interferons or signaling proteins released by cells in response to pathogens.¹⁴⁰ IFN- β is most widely used in MS treatment and has been shown in vivo to act through the upregulation of PD-L1, which results in a decrease in neuroinflammation and a therapeutic effect for patients with MS.^{140,141} A previous study illustrated that PD-L1 expression inversely correlates with strong CNS inflammation in MS plaques, further suggesting that PD-L1

acts in a protective manner, downregulating the inflammation associated with T-cell extension into the CNS.¹⁴²

Several studies have shown a correlation between the upregulated expression of PD-1/PD-L1 and the associated MS stage using the EAE mouse model. A study using in vitro techniques first found that PD-L1 overexpression by DCs had the potential to prevent the induction of EAE.¹⁴³ These data were then reinforced by later in vivo studies, where MS was induced following MOG immunization in PD-L1 knockout mice and more severe EAE developed relative to the control.¹³⁵ Similarly, Ortler et al¹⁴² were also able to show using flow cytometry that following immunization with MOG in PD-L1 knockout mice, there was a significant increase in T-cell infiltration in the brain, indicating a correlation between the increased severity of EAE and absence of PD-L1 in mouse models. Other studies have shown that T cells interact with DCs, which then leads to the upregulation of PD-1/PD-L1 expression.^{122,144} As stated previously, elevated PD-1/PD-L1 levels can limit the severity of EAE, if not prevent its induction. This indicates that PD-1 overexpression due to its interactions with DCs may hold therapeutic value. Based on this evidence, one possible avenue of MS treatment may lie in the upregulation of PD-1/PD-L1, which needs to be further explored in later research.

Suppressor of Cytokine Signaling and its Association with MS

A key gene involved in the function of T_H cells and Tregs is SOCS1, a member of the SOCS family. In general, SOCS genes are important in the regulation of cytokine signaling and autoimmune responses through the JAK/STAT pathway.^{145–147} SOCS1 plays a role in the regulation of immune cell homeostasis as well as in inflammation.¹⁴⁸ SOCS1 suppresses DC activity by inhibition of STAT1, which normally activates DCs.¹⁴⁹ SOCS1 has been shown to play a key role in regulating DCs regarding autoimmunity, as SOCS1-deficient DCs are overly susceptible to IL-4 and IFN- γ , 2 inflammatory cytokines. This enables the production of autoreactive antibodies and systemic autoimmunity.¹⁵⁰ Single-nucleotide polymorphisms (SNPs) near SOCS1 have an association with MS by reducing SOCS1 gene expression and initiating the differentiation of T cells to a pathogenic T_H17 phenotype, increasing in the risk of MS onset as described earlier.^{151–153} Research has shown that SNPs that cause the loss of SOCS1 function do not act through nearby genes that associate with SOCS1. Instead, SOCS1-deficient Tregs lose their suppressive activity. This causes the production of increased levels of IL-17, which induces the specialization of T_H17 from naïve T cells.^{151,153} This evidence indicates that a main role of SOCS1 is to suppress systemic autoimmunity, such as is present in MS, through its standard interactions with DCs.

Another gene expressed by APCs, particularly DCs, is C-type lectin domain family 16A (CLEC16A), which associates with SOCS1 in regulating immune response.^{149,150}

Polymorphisms of this MS risk allele (especially a SNP at rs12708716 found within intron 19 of CLEC16A that results in reduced SOCS1 expression) influence the susceptibility for various autoimmune diseases, specifically MS, through the reduction in SOCS1 protein expression in the thymus.⁹ It has been reported that in the absence of the SOCS1 protein, mice experienced systemic inflammation as well as the development of thymocytes.¹⁴⁸ These symptoms are similar to those observed in MS, providing further evidence that the SOCS1 gene plays an important role in the pathogenesis of MS, with reduction in SOCS1 expressions leaving DCs over-responsive to cytokines and resulting in autoimmunity as discussed above. Based on this evidence, SOCS1 and its relationship with CLEC16A and the associated effects on DCs should be further explored. Such studies would aid in determining whether there is therapeutic potential in new treatments for MS geared toward the upregulation of SOCS1 protein expression through targeting of the associated SOCS1 and CLEC16A genes.

Summary

Multiple sclerosis is a neurodegenerative disease characterized by inflammation of the CNS that results in demyelination. Although the cause of MS is still unknown, it is believed to result from a mixture of viral, genetic, and environmental factors. It has largely been attributed to an autoimmune response stemming from EBV due to its prevalence in patients with MS, but immune misregulation plays a clear role in the presentation of disease symptoms. Both T_H1 and T_H17 cells are important to MS pathogenesis and are known to primarily be stimulated by DCs.¹⁶ This pathogenic stimulation is a combination of myelin-specific antigen presentation, costimulation, and cytokine signaling.^{17–19} Therefore, understanding and altering the interaction between DCs and $CD4^+$ T cells is important for developing possible treatments for MS.

Cytokine signaling involving T_H cells and DCs likely plays a significant role in MS pathology. The suppression of cytokine signaling between DCs and $CD4^+$ T_H cells plays a large role in MS progression through its associations with other genes, which in turn helps T cells function within the immune system.^{28,152} When cytokine function is abnormal, this leads to an autoimmune reaction within the body. Thus, cytokine regulation is a good potential therapeutic target for patients with MS.⁹ The upregulation of IL-23 is widely accepted as a key checkpoint in the pathogenesis of MS because of its protective and differentiative role in T_H17 cells.⁷ Finding ways to decrease the elevated serum and CNS levels of IL-23 in MS and EAE models is a major point of interest. Therapeutic approaches such as IL-23p19-specific mAbs, antisense oligos against IL-23 mRNA, and sIL-23R have shown to downregulate the expression of IL-23 in MS models.^{28,89,92} However, further research should be done in the extracted cells of patients with MS to elucidate the role of these potential treatments in disease progression to develop effective clinical versions to use as

treatments. In addition, further study into the IL-23R-STAT3 pathway and its role in IL-17 and IL-23R regulation is a promising avenue that could lead to improved treatment of MS.^{92,98}

Multiple sclerosis onset has also been linked to the B7/T-cell costimulatory pathway, particularly the B7-CD28/CTLA-4 costimulatory pathway. The B7 molecules, CD80 and CD86, interact with receptors found on DCs and when the balance of these interacting proteins is off, exacerbated symptoms of MS often develop. Although it has been suggested that CTLA-4 serves a protective role in the brain that leads to MS once inhibited,^{123,124} there is conflicting research regarding this,^{133,134} calling for additional study to determine the true nature of this relationship. There has also been a lack of consistency in the effects of CTLA-4 polymorphisms on varying cohorts,^{131–134} which indicates a possible ethnic relationship between MS and variations in CTLA-4. The PD-L1/PD-1 costimulatory pathway has also been recently reported to play a role in the progression of MS when downregulated by DCs. Therefore, using therapies that increase PD-1/PD-L1 activity holds promise for slowing MS disease state.

In this review, we have illustrated the need for more research into the impact of DCs on immune regulation in MS. By looking at the interactions between DCs and T cells, there is opportunity to further elucidate the role of ILs, $CD4^+$ T cells, PD-1/PD-L1, CTLA-4, and SOCS1 in MS onset and progression due to the irregular activity found in each during disease. More research into the role of the PD-L1/PD-1 costimulatory pathway may result in further treatment avenues to slow MS progression. We have also demonstrated a need to further analyze the role of IL-23 in the differentiation of T_H17 cells, enabling greater understanding of the role of IL-23R in MS. The effects of the upregulation of SOCS1 in patients having an autoimmune disease also require deeper investigation. Continued focus in these areas may result in potential therapeutic value for patients living with MS, ultimately having a long-term impact and improving their quality of life.

Acknowledgements

The authors would like to thank Tia M Spagnuolo, MS, CF-SLP; Teresa Wasiluk, MS; and John Forsberg, MS, for reviewing the manuscript. They would also like to thank Austin Ford for his initial thoughts on the manuscript.

Author Contributions

GS and AP conceived the concept, wrote the first draft of the manuscript, contributed to the writing of the manuscript, and jointly developed the structure and arguments for the paper. GS, AP, and TW agree with manuscript conclusions and made critical revisions. GS contributed to the formatting and compilation of manuscript. All authors reviewed and approved the final manuscript.

REFERENCES

- Lill CM. Recent advances and future challenges in the genetics of multiple sclerosis. *Front Neurol*. 2014;5:140.
- Ascherio A, Munger KL, Lennette ET, et al. Epstein-Barr virus antibodies and risk of multiple sclerosis. *JAMA*. 2001;286:3083–3088.
- Angelini DF, Serafini B, Piras E, et al. Increased CD8+ T cell response to Epstein-Barr virus lytic antigens in the active phase of multiple sclerosis. *PLoS Pathog*. 2013;9:e1003220.
- Vereide DT, Seto E, Chiu Y-F, et al. Epstein-Barr virus maintains lymphomas via its miRNAs. *Oncogene*. 2014;33:1258–1264.
- Lünemann JD, Edwards N, Muraro PA, et al. Increased frequency and broadened specificity of latent EBV nuclear antigen-1-specific T cells in multiple sclerosis. *Brain*. 2006;129:1493–1506.
- Graves MC, Benton M, Lea RA, et al. Methylation differences at the HLA-DRB1 locus in CD4+ T-Cells are associated with multiple sclerosis. *Mult Scler*. 2014;20:1033–1041.
- Langrish CL, Chen Y, Blumenschein WM, et al. IL-23 drives a pathogenic T cell population that induces autoimmune inflammation. *J Exp Med*. 2005;201:233–240.
- Vogel I, Kasran A, Cremer J, et al. CD28/CTLA-4/B7 costimulatory pathway blockade affects regulatory T-cell function in autoimmunity. *Eur J Immunol*. 2015;45:1832–1841.
- Leikfoss IS, Mero I-L, Dahle MK, et al. Multiple sclerosis-associated single-nucleotide polymorphisms in CLEC16A correlate with reduced SOCS1 and DEXI expression in the thymus. *Genes and Immun*. 2013;14:62–66.
- Zhou L, Chong MM, Littman DR. Plasticity of CD4+ T cell lineage differentiation. *Immunity*. 2009;30:646–655.
- Kurschus FC. T cell mediated pathogenesis in EAE: molecular mechanisms. *Biomed J*. 2015;38:183–193.
- Luchtman DW, Ellwardt E, Laroche C, Zipp F. IL-17 and related cytokines involved in the pathology and immunotherapy of multiple sclerosis: current and future developments. *Cytokine Growth Factor Rev*. 2014;25:403–413.
- Kivisäkk P, Imitola J, Rasmussen S, et al. Localizing central nervous system immune surveillance: meningeal antigen-presenting cells activate T cells during experimental autoimmune encephalomyelitis. *Ann Neurol*. 2009;65:457–469.
- Goverman J. Autoimmune T cell responses in the central nervous system. *Nat Rev Immunol*. 2009;9:393–407.
- Otterbein LE, Soares MP, Yamashita K, Bach FH. Heme oxygenase-1: unleashing the protective properties of heme. *Trends Immunol*. 2003;24:449–455.
- Satpathy AT, Wu X, Albring JC, Murphy KM. Re(de)fining the dendritic cell lineage. *Nat Immunol*. 2012;13:1145–1154.
- Steinman RM, Banchereau J. Taking dendritic cells into medicine. *Nature*. 2007;449:419–426.
- Chastain EM, Duncan DS, Rodgers JM, Miller SD. The role of antigen presenting cells in multiple sclerosis. *Biochim Biophys Acta*. 2011;1812:265–274.
- Joffe O, Nolte MA, Sporri R, Reis e Sousa C. Inflammatory signals in dendritic cell activation and the induction of adaptive immunity. *Immunol Rev*. 2009;227:234–247.
- Yokoi H, Kato K, Kezuka T, et al. Prevention of experimental autoimmune uveoretinitis by monoclonal antibody to interleukin-12. *Eur J Immunol*. 1997;27:641–646.
- Yoshimura T, Sonoda KH, Ohguro N, et al. Involvement of Th17 cells and the effect of anti-IL-6 therapy in autoimmune uveitis. *Rheumatology (Oxford)*. 2009;48:347–354.
- Cua DJ, Sherlock J, Chen Y, et al. Interleukin-23 rather than interleukin-12 is the critical cytokine for autoimmune inflammation of the brain. *Nature*. 2003;421:744–748.
- Chang J, Kunkel SL, Chang CH. Negative regulation of MyD88-dependent signaling by IL-10 in dendritic cells. *Proc Natl Acad Sci USA*. 2009;106:18327–18332.
- Lande R, Gafa V, Serafini B, et al. Plasmacytoid dendritic cells in multiple sclerosis: intracerebral recruitment and impaired maturation in response to interferon-beta. *J Neuropathol Exp Neurol*. 2008;67:388–401.
- Longhini AL, von Glehn F, Brandao CO, et al. Plasmacytoid dendritic cells are increased in cerebrospinal fluid of untreated patients during multiple sclerosis relapse. *J Neuroinflammation*. 2011;8:2.
- Karni A, Abraham M, Monsonego A, et al. Innate immunity in multiple sclerosis: myeloid dendritic cells in secondary progressive multiple sclerosis are activated and drive a proinflammatory immune response. *J Immunol*. 2006;177:4196–4202.
- Huang YM, Xiao BG, Ozenci V, et al. Multiple sclerosis is associated with high levels of circulating dendritic cells secreting pro-inflammatory cytokines. *J Neuroimmunol*. 1999;99:82–90.
- Vaknin-Dembinsky A, Balashov K, Weiner HL. IL-23 is increased in dendritic cells in multiple sclerosis and down-regulation of IL-23 by antisense oligos increases dendritic cell IL-10 production. *J Immunol*. 2006;176: 7768–7774.
- McMahon EJ, Bailey SL, Castenada CV, Waldner H, Miller SD. Epitope spreading initiates in the CNS in two mouse models of multiple sclerosis. *Nat Med*. 2005;11:335–339.
- Guilliams M, Henri S, Tamoutounour S, et al. From skin dendritic cells to a simplified classification of human and mouse dendritic cell subsets. *Eur J Immunol*. 2010;40:2089–2094.
- Robbins SH, Walzer T, Dembele D, et al. Novel insights into the relationships between dendritic cell subsets in human and mouse revealed by genome-wide expression profiling. *Genome Biol*. 2008;9:R17.
- Crozat K, Guiton R, Contreras V, et al. The XC chemokine receptor 1 is a conserved selective marker of mammalian cells homologous to mouse CD8alpha+ dendritic cells. *J Exp Med*. 2010;207:1283–1292.
- Jongbloed SL, Kassianos AJ, McDonald KJ, et al. Human CD141+ (BDCA-3)+ dendritic cells (DCs) represent a unique myeloid DC subset that cross-presents necrotic cell antigens. *J Exp Med*. 2010;207:1247–1260.
- Poulin LF, Salio M, Griessinger E, et al. Characterization of human DNGR-1+ BDCA3+ leukocytes as putative equivalents of mouse CD8alpha+ dendritic cells. *J Exp Med*. 2010;207:1261–1271.
- Bachem A, Guttler S, Hartung E, et al. Superior antigen cross-presentation and XCR1 expression define human CD11c+CD141+ cells as homologues of mouse CD8+ dendritic cells. *J Exp Med*. 2010;207:1273–1281.
- Haniffa M, Shin A, Bigley V, et al. Human tissues contain CD141hi cross-presenting dendritic cells with functional homology to mouse CD103+ non-lymphoid dendritic cells. *Immunity*. 2012;37:60–73.
- Segura E, Touzot M, Bohineust A, et al. Human inflammatory dendritic cells induce Th17 cell differentiation. *Immunity*. 2013;38:336–348.
- Helft J, Ginhoux F, Bogunovic M, Merad M. Origin and functional heterogeneity of non-lymphoid tissue dendritic cells in mice. *Immunol Rev*. 2010;234:55–75.
- Serafini B, Rosicarelli B, Magliozzi R, et al. Dendritic cells in multiple sclerosis lesions: maturation stage, myelin uptake, and interaction with proliferating T cells. *J Neuropathol Exp Neurol*. 2006;65:124–141.
- Dittel BN, Visintin Merchant RM, Janeway CA. Presentation of the self antigen myelin basic protein by dendritic cells leads to experimental autoimmune encephalomyelitis. *J Immunol*. 1999;163:32–39.
- Greter M, Heppner FL, Lemos MP, et al. Dendritic cells permit immune invasion of the CNS in an animal model of multiple sclerosis. *Nat Med*. 2005;11:328–334.
- Stasiolek M, Bayas A, Kruse N, et al. Impaired maturation and altered regulatory function of plasmacytoid dendritic cells in multiple sclerosis. *Brain*. 2006;129:1293–1305.
- Pashenkov M, Huang YM, Kostulas V, et al. Two subsets of dendritic cells are present in human cerebrospinal fluid. *Brain*. 2001;124:480–492.
- León B, López-Bravo M, Ardavin C. Monocyte-derived dendritic cells formed at the infection site control the induction of protective T helper 1 responses against *Leishmania*. *Immunity*. 2007;26:519–531.
- Shortman K, Naik SH. Steady-state and inflammatory dendritic-cell development. *Nat Rev Immunol*. 2007;7:19–30.
- Mangalam A, Luckey D, Basal E, et al. HLA-DQ8 (DQB1*0302)-restricted Th17 cells exacerbate experimental autoimmune encephalomyelitis in HLA-DR3-transgenic mice. *J Immunol*. 2009;182:5131–5139.
- Tzartos JS, Friese MA, Craner MJ, et al. Interleukin-17 production in central nervous system-infiltrating T cells and glial cells is associated with active disease in multiple sclerosis. *Am J Pathol*. 2008;172:146–155.
- Krakauer M, Sorensen P, Khademi M, Olsson T, Sellebjerg F. Increased IL-10 mRNA and IL-23 mRNA expression in multiple sclerosis: interferon-beta treatment increases IL-10 mRNA expression while reducing IL-23 mRNA expression. *Mult Scler*. 2008;14:622–630.
- Alexander J, Harris M, Wells S, et al. Alterations in serum MMP-8, MMP-9, IL-12p40 and IL-23 in multiple sclerosis patients treated with interferon-beta1b. *Mult Scler*. 2010;16:801–809.
- Becher B, Durell B, Noelle R. Experimental autoimmune encephalitis and inflammation in the absence of interleukin-12. *J Clin Invest*. 2002;110:493–497.
- Gran B, Zhang G, Yu S, et al. IL-12p35-deficient mice are susceptible to experimental autoimmune encephalomyelitis: evidence for redundancy in the IL-12 system in the induction of central nervous system autoimmune demyelination. *J Immunol*. 2002;169:7104–7110.
- Komiyama Y, Nakae S, Matsuki T, et al. IL-17 plays an important role in the development of experimental autoimmune encephalomyelitis. *J Immunol*. 2006;177:566–573.
- Sutton C, Brereton C, Keogh B, Mills KHG, Lavelle EC. A crucial role for interleukin (IL)-1 in the induction of IL-17-producing T cells that mediate autoimmune encephalomyelitis. *J Exp Med*. 2006;203:1685–1691.
- Wilson NJ, Boniface K, Chan JR, et al. Development, cytokine profile and function of human interleukin 17-producing helper T cells. *Nat Immunol*. 2007;8:950–957.

55. Seiderer J, Elben I, Diegelmann J, et al. Role of the novel Th17 cytokine IL-17F in inflammatory bowel disease (IBD): upregulated colonic IL-17F expression in active Crohn's disease and analysis of the IL17F p.His161Arg polymorphism in IBD. *Inflamm Bowel Dis*. 2008;14:437–445.
56. Figueroa-Vega N, Alfonso-Perez M, Benedicto I, et al. Increased circulating pro-inflammatory cytokines and Th17 lymphocytes in Hashimoto's thyroiditis. *J Clin Endocrinol Metab*. 2010;95:953–962.
57. Sarkar S, Cooney LA, Fox DA. The role of T helper type 17 cells in inflammatory arthritis. *Clin Exper Immunol*. 2010;159:225–237.
58. Oppmann B, Lesley R, Blom B, et al. Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity*. 2000;13:715–725.
59. Lee E, Trepicchio WL, Oestreicher JL, et al. Increased expression of interleukin 23 p19 and p40 in lesional skin of patients with psoriasis vulgaris. *J Exp Med*. 2004;199:125–130.
60. Haak S, Croxford AL, Kreymborg K, et al. IL-17A and IL-17F do not contribute vitally to autoimmune neuro-inflammation in mice. *J Clin Invest*. 2009;119:61–69.
61. Ferber IA, Brocke S, Taylor-Edwards C, et al. Mice with a disrupted IFN-gamma gene are susceptible to the induction of experimental autoimmune encephalomyelitis (EAE). *J Immunol*. 1996;156:5–7.
62. Teng M, Bownman E, McElwee J, et al. IL-12 and IL-23 cytokines: from discovery to targeted therapies for immune-mediated inflammatory diseases. *Nat Med*. 2015;21:719–729.
63. Casanova JL, Abel L. Genetic dissection of immunity to mycobacteria: the human model. *Annu Rev Immunol*. 2002;20:581–620.
64. Filipe-Santos O, Bustamante J, Chappier A, et al. Inborn errors of IL-12/23- and IFN- γ -mediated immunity: molecular, cellular, and clinical features. *Semin Immunol*. 2006;18:347–361.
65. Al-Muhsen S, Casanova JL. The genetic heterogeneity of mendelian susceptibility to mycobacterial diseases. *J Allergy Clin Immunol*. 2008;122:1043–1051.
66. Bogunovic D, Byun M, Durfee LA, et al. Mycobacterial disease and impaired IFN- γ immunity in humans with inherited ISG15 deficiency. *Science*. 2012;337:1684–1688.
67. Alcaïs A, Fieschi C, Abel L, Casanova JL. Tuberculosis in children and adults: two distinct genetic diseases. *J Exp Med*. 2005;202:1617–1621.
68. Abel L, El-Baghdadi J, Bousfiha AA, Casanova JL, Schurr E. Human genetics of tuberculosis: a long and winding road. *Phil Trans R Soc Lond B*. 2014;369:20130428.
69. Khader SA, Pearl JE, Sakamoto K, et al. IL-23 compensates for the absence of IL-12p70 and is essential for the IL-17 response during tuberculosis but is dispensable for protection and antigen-specific IFN- γ responses if IL-12p70 is available. *J Immunol*. 2005;175:788–795.
70. Lieberman LA, Cardillo F, Owyang AM, et al. IL-23 provides a limited mechanism of resistance to acute toxoplasmosis in the absence of IL-12. *J Immunol*. 2004;173:1887–1893.
71. Chen Y, Langrish CL, McKenzie B, et al. Anti-IL-23 therapy inhibits multiple inflammatory pathways and ameliorates autoimmune encephalomyelitis. *J Clin Invest*. 2006;116:1317–1326.
72. Segal BM, Constantinescu CS, Raychaudhuri A, et al. Repeated subcutaneous injections of IL12/23 p40 neutralising antibody, ustekinumab, in patients with relapsing-remitting multiple sclerosis: a phase II, double-blind, placebo-controlled, randomised, dose-ranging study. *Lancet Neurol*. 2008;7:796–804.
73. Vollmer TL, Wynn DR, Alam MS, Valdes J. A phase 2 24-week randomized placebo-controlled double-blind study examining the efficacy and safety of an anti-interleukin-12 and -23 monoclonal antibody in patients with relapsing-remitting or secondary progressive multiple sclerosis. *Mult Scler*. 2011;17:181–191.
74. Sandborn WJ, Gasink C, Gao L-L, et al. Ustekinumab induction and maintenance therapy in refractory Crohn's disease. *N Engl J Med*. 2012;367:1519–1528.
75. Gordon KB, Langley RG, Gottlieb AB, et al. A phase III, randomized, controlled trial of the fully human IL-12/23 mAb briakinumab in moderate-to-severe psoriasis. *J Invest Dermatol*. 2011;132:304–314.
76. Gordon KB, Duffin KC, Bissonnette R, et al. A phase 2 trial of guselkumab versus adalimumab for plaque psoriasis. *N Engl J Med*. 2015;373:136–144.
77. Papp K, Thaçi D, Reich K, et al. Tildrakizumab (MK-3222), an anti-interleukin-23p19 monoclonal antibody, improves psoriasis in a phase IIb randomized placebo-controlled trial. *Br J Dermatol*. 2015;173: 930–939.
78. Polman CH, Bertolotto A, Deisenhammer F, et al. Recommendations for clinical use of data on neutralising antibodies to interferon-beta therapy in multiple sclerosis. *Lancet Neurol*. 2010;9:740–750.
79. Wolbink GJ, Vis M, Lems W, et al. Development of antiinfiximab antibodies and relationship to clinical response in patients with rheumatoid arthritis. *Arthritis Rheum*. 2006;54:711–715.
80. Baert F, Noman M, Vermeire S, et al. Influence of immunogenicity on the long-term efficacy of infliximab in Crohn's disease. *N Engl J Med*. 2003;348:601–608.
81. Bartelds GM, Wijbrandts CA, Nurmohamed MT, et al. Clinical response to adalimumab: relationship to antiadalimumab antibodies and serum adalimumab concentrations in rheumatoid arthritis. *Ann Rheum Dis*. 2007;66:921–926.
82. Bartelds GM, Krieckaert CL, Nurmohamed MT, et al. Development of anti-drug antibodies against adalimumab and association with disease activity and treatment failure during long-term follow-up. *JAMA*. 2011;305:1460–1468.
83. Farrell RA, Giovannoni G. Measuring and management of anti-interferon beta antibodies in subjects with multiple sclerosis. *Mult Scler*. 2007;13:567–577.
84. Calabresi PA, Giovannoni G, Confavreux C, et al. The incidence and significance of anti-natalizumab antibodies: results from AFFIRM and SENTINEL. *Neurology*. 2007;69:1391–1403.
85. Vennegoor A, Rispens T, Strijbis EM, et al. Clinical relevance of serum natalizumab concentration and anti-natalizumab antibodies in multiple sclerosis. *Mult Scler*. 2013;19:593–600.
86. Grimm D, Kay MA. RNAi and gene therapy: a mutual attraction. *Hematology Am Soc Hematol Educ Program*. 2007;2007:437–481.
87. Monteleone G, Fantini MC, Onali S, et al. Phase I clinical trial of Smad7 knockdown using antisense oligonucleotide in patients with active Crohn's disease. *Mol Ther*. 2012;20:870–876.
88. Geremia A, Arancibia-Carcamo CV, Fleming MP, et al. IL-23-responsive innate lymphoid cells are increased in inflammatory bowel disease. *J Exp Med*. 2011;208:1127–1133.
89. Kalantari T, Karimi MH, Ciric B, et al. Tolerogenic dendritic cells produced by lentiviral-mediated CD40- and interleukin-23p19-specific shRNA can ameliorate experimental autoimmune encephalomyelitis by suppressing T helper type 17 cells. *Clin Exper Immunol*. 2014;176:180–189.
90. Krajina T, Leithhauser R, Muller P, Trobonjaca Z, Reimann J. Colonic lamina propria dendritic cells in mice with CD4+ T cell-induced colitis. *Eur J Immunol*. 2003;33:1073–1083.
91. Guo W, Luo C, Wang C, et al. Suppression of human and mouse Th17 differentiation and autoimmunity by an endogenous Interleukin 23 receptor cytokine-binding homology region. *Int J Biochem Cell Biol*. 2014;55:304–310.
92. Miralles M, Eixarch H, Tejero M, et al. Clinical and histopathological amelioration of experimental autoimmune encephalomyelitis by AAV vectors expressing a soluble interleukin-23 receptor [published online ahead of print June 7, 2017]. *Neurotherapeutics*. doi:10.1007/s13311-017-0545-8.
93. Nakai H, Fuess S, Storm TA, et al. Unrestricted hepatocyte transduction with adeno-associated virus serotype 8 vectors in mice. *J Virol*. 2005;79:214–224.
94. Wang Z, Zhu T, Qiao C, et al. Adeno-associated virus serotype 8 efficiently delivers genes to muscle and heart. *Nat Biotechnol*. 2005;23:321–328.
95. Yu RY, Gallagher G. A naturally occurring, soluble antagonist of human IL-23 inhibits the development and in vitro function of human Th17 cells. *J Immunol*. 2010;185:7302–7308.
96. Parham C, Chirica M, Timans J, et al. A receptor for the heterodimeric cytokine IL-23 is composed of IL-12R β 1 and a novel cytokine receptor subunit. *J Immunol*. 2002;168:5699–5708.
97. Ghoreschi K, Laurence A, Yang X, et al. Generation of pathogenic T(H)17 cells in the absence of TGF- β signalling. *Nature*. 2010;467:967–971.
98. Oestreich KJ, Weinmann AS. Master regulators or lineage-specifying? Changing views on CD4+ T cell transcription factors. *Nat Rev Immunol*. 2012;12:799–804.
99. Harrington LE, Hatton RD, Mangan PR, et al. Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol*. 2005;6:1123–1132.
100. Park H, Li Z, Yang XO, et al. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat Immunol*. 2005;6:1133–1141.
101. Billiau A, Heremans H, Vandekerckhove F, et al. Enhancement of experimental allergic encephalomyelitis in mice by antibodies against IFN- γ . *J Immunol*. 1988;140:1506–1510.
102. Heremans H, Dillen C, Groenen M, et al. Chronic relapsing experimental autoimmune encephalomyelitis (CREAE) in mice: enhancement by monoclonal antibodies against interferon- γ . *Eur J Immunol*. 1996;26:2393–2398.
103. Lublin FD, Knobler RL, Kalman B, et al. Monoclonal anti-gamma interferon antibodies enhance experimental allergic encephalomyelitis. *Autoimmunity*. 1993;16:267–274.
104. Willenborg DO, Fordham S, Bernard CC, et al. IFN- γ plays a critical down-regulatory role in the induction and effector phase of myelin oligodendrocyte glycoprotein-induced autoimmune encephalomyelitis. *J Immunol*. 1996;157:3223–3227.
105. Pettinelli CB, McFarlin DE. Adoptive transfer of experimental allergic encephalomyelitis in SJL/J mice after in vitro activation of lymph node cells by myelin basic protein: requirement for Lyt 1+ 2- T lymphocytes. *J Immunol*. 1981;127:1420–1423.
106. Panitch HS. Interferons in multiple sclerosis: a review of the evidence. *Drugs*. 1992;44:946–962.
107. Chitnis T, Najafian N, Benou C, et al. Effect of targeted disruption of STAT4 and STAT6 on the induction of experimental autoimmune encephalomyelitis. *J Clin Invest*. 2001;108:739–747.

108. Furlan R, Brambilla E, Ruffini F, et al. Intrathecal delivery of IFN-gamma protects C57BL/6 mice from chronic-progressive experimental autoimmune encephalomyelitis by increasing apoptosis of central nervous system-infiltrating lymphocytes. *J Immunol.* 2001;167:1821–1829.
109. Tanuma N, Shin T, Kogure K, Matsumoto Y. Differential role of TNF- α and IFN- γ in the brain of rats with chronic relapsing autoimmune encephalomyelitis. *J Neuroimmunol.* 1999;96:73–79.
110. Naves R, Singh SP, Cashman KS, et al. The interdependent, overlapping, and differential roles of type I and II IFNs in the pathogenesis of experimental autoimmune encephalomyelitis. *J Immunol.* 2013;191:2967–2977.
111. Camperio C, Muscolini M, Volpe E, et al. CD28 ligation in the absence of TCR stimulation up-regulates IL-17A and pro-inflammatory cytokines in relapsing-remitting multiple sclerosis T lymphocytes. *Immunol Lett.* 2014;158:134–142.
112. Broux B, Mizze MR, Vanheusen M, et al. IL-15 amplifies the pathogenic properties of CD4⁺CD28⁻ T cells in multiple sclerosis. *J Immunol.* 2015;194:2099–2109.
113. Markovic-Plese S, Cortese I, Wandinger K, McFarland H, Martin R. CD4⁺CD28⁻ costimulation-independent T cells in multiple sclerosis. *J Clin Invest.* 2001;108:1185–1194.
114. Viglietta V, Baecher-Allan C, Weiner HL, Hafler DA. Loss of functional suppression by CD4⁺CD25⁺ regulatory T cells in patients with multiple sclerosis. *J Exp Med.* 2004;199:971–979.
115. Haanstra KG, Dijkman K, Bashir N, et al. Selective blockade of CD28-mediated T cell costimulation protects rhesus monkeys against acute fatal experimental autoimmune encephalomyelitis. *J Immunol.* 2015;194:1454–1466.
116. Carbone F, De Rosa V, Carrieri P, et al. Regulatory T cell proliferative potential is impaired in human autoimmune disease. *Nat Med.* 2014;20:69–74.
117. Ligiers A, Teleshova N, Masterman T, Huang W-X, Hillert J. CTLA-4 gene expression is influenced by promoter and exon 1 polymorphisms. *Genes Immun.* 2001;2:145–152.
118. Gerdes LA, Held K, Beltrán E, et al. CTLA4 as immunological checkpoint in the development of multiple sclerosis. *Ann Neurol.* 2016;80:294–300.
119. Wagner M, Sobczynski M, Karabon L, et al. Polymorphisms in CD28 CTLA-4 CD80 and CD86 genes may influence the risk of multiple sclerosis and its age of onset. *J Neuroimmunol.* 2015;288:79–86.
120. Suppiah V, Alloza I, Heggarty S, et al. The CTLA4 +49 A/G*G-CT60*G haplotype is associated with susceptibility to multiple sclerosis in Flanders. *J Neuroimmunol.* 2005;164:148–153.
121. Podojil JR, Miller SD. Targeting the B7 family of co-stimulatory molecules: successes and challenges. *BioDrugs.* 2013;27:1–13.
122. Xie Z-X, Zhang H-L, Wu X-J, et al. Role of the immunogenic and tolerogenic subsets of dendritic cells in multiple sclerosis. *Mediators Inflamm.* 2015;2015:513295.
123. Cao Y, Nylander A, Ramanan S, et al. CNS demyelination and enhanced myelin-reactive responses after ipilimumab treatment. *Neurology.* 2016;86:1553–1556.
124. Gettings EJ, Hackett CT, Scott TF. Severe relapse in a multiple sclerosis patient associated with ipilimumab treatment of melanoma. *Mult Scler.* 2015;21:670.
125. Chitnis T, Khoury SJ. Role of costimulatory pathways in the pathogenesis of multiple sclerosis and experimental autoimmune encephalomyelitis. *J Allergy Clin Immunol.* 2003;112:837–849.
126. Walker LSK, Sansom DM. The emerging role of CTLA4 as a cell-extrinsic regulator of T cell responses. *Nat Rev Immunol.* 2011;11:852–863.
127. Schildberg FA, Klein SR, Freeman GJ, Sharpe AH. Coinhibitory Pathways in the B7-CD28 Ligand-Receptor Family. *Immunity.* 2016;44:955.
128. Khoury SJ, Rochon J, Ding L, et al. ACCLAIM: a randomized trial of abatacept (CTLA4-Ig) for relapsing-remitting multiple sclerosis. *Mult Scler.* 2016;23:686–695.
129. Viglietta V, Bourcier K, Buckle G, et al. CTLA4IG treatment in patients with multiple sclerosis. *Neurology.* 2008;71:917–924.
130. Holley JE, Bremer E, Kendall AC, et al. CD20⁺ inflammatory T-cells are present in blood and brain of multiple sclerosis patients and can be selectively targeted for apoptotic elimination. *Mult Scler Relat Disord.* 2014;3:650–658.
131. Malferrari G, Stella A, Monferini E, et al. CTLA4 and multiple sclerosis in the Italian population. *Exp Mol Pathol.* 2005;78:55–57.
132. Fukazawa T, Kikuchi S, Miyagishi R, et al. CTLA-4 gene polymorphism is not associated with conventional multiple sclerosis in Japanese. *J Neuroimmunol.* 2005;159:225–229.
133. Heidari A, Keramatipour M, Amirzargar AA, et al. CTLA-4 gene polymorphisms (–318C/T, +49A/G, +6230A/G) in Iranian patients with multiple sclerosis. *Iran J Allergy Asthma Immunol.* 2010;9:219–223.
134. Greve B, Simonenko R, Illes Z, et al. Multiple sclerosis and the CTLA4 autoimmune polymorphism CT60: no association in patients from Germany, Hungary and Poland. *Mult Scler.* 2008;14:153–158.
135. Trabattini D, Saresella M, Pacci M, et al. Costimulatory pathways in multiple sclerosis: distinctive expression of PD-1 and PD-L1 in patients with different patterns of disease. *J Immunol.* 2009;183:4984–4993.
136. Schachtele S, Hu S, Sheng W, Mutnal M, Lokensgard J. Glial cells suppress post-encephalitic CD8⁺ T lymphocytes through PD-L1. *GLIA.* 2014;62:1582–1594.
137. Sharpe AH, Wherry EJ, Ahmed R, Freeman GJ. The functions of programmed cell death 1 and its ligands in regulating autoimmunity and infection. *Nat Immunol.* 2007;8:239–245.
138. Javan MR, Aslani S, Zamani MR, et al. Downregulation of immunosuppressive molecules PD-1 and PD-L1 but not PD-L2 in the patients with multiple sclerosis. *Iran J Allergy Asthma Immunol.* 2016;15:296–302.
139. Schreiner B, Bailey SL, Shin T, Chen L, Miller SD. PD-1 ligands expressed on myeloid-derived APC in the CNS regulated T-cell responses in EAE. *Eur J Immunol.* 2008;38:2706–2717.
140. Schreiner B, Mitsdoerffer M, Kieseier BC, et al. Interferon- β enhances monocyte and dendritic cell expression of B7-H1. *J Neuroimmunol.* 2004;155:172–182.
141. Harari D, Kuhn N, Abramovich R, et al. Enhanced in vivo efficacy of a type I interferon superagonist with extended plasma half-life in a mouse model of multiple sclerosis. *J Biol Chem.* 2014;289:29014–29029.
142. Ortler S, Leder C, Mittelbronn M, et al. B7-H1 restricts neuroantigen-specific T cell responses and confines inflammatory CNS damage: implications for the lesion pathogenesis of multiple sclerosis. *Eur J Immunol.* 2008;38:1734–1744.
143. Kuipers H, Muskens F, Willart M, et al. Contribution of the PD-1 ligands/PD-1 signaling pathway to dendritic cell-mediated CD4⁺ T cell activation. *Eur J Immunol.* 2006;36:2472–2482.
144. Yogeve N, Frommer F, Lukas D, et al. Dendritic cells ameliorate autoimmunity in the CNS by controlling the homeostasis of PD-1 receptor⁺ regulatory T cells. *Immunity.* 2012;37:264–275.
145. Kubo M, Hanada T, Yoshimura A. Suppressors of cytokine signaling and immunity. *Nat Immunol.* 2003;4:1169–1176.
146. Bullen DVR, Baldwin TM, Curtis JM, Alexander WS, Handman E. Persistence of lesions in suppressor of cytokine signaling-1-deficient mice infected with *Leishmania major*. *J Immunol.* 2003;170:4267–4272.
147. Pahlevan Kakhki M, Rakhshi N, Heidary M, Behmanesh M, Nikraves A. Expression of suppressor of cytokine signaling 1 (SOCS1) gene dramatically increases in relapsing-remitting multiple sclerosis. *J Neurol Sci.* 2015;350:40–45.
148. Hedrick SM, Catlett IM. Suppressor of cytokine signaling 1 is required for the differentiation of CD4⁺ T cells. *Nat Immunol.* 2005;6:715–721.
149. Hoppenbrouwers I, Aulchenko Y, Janssens A, et al. Replication of CD58 and CLEC16A as genome-wide significant risk genes for multiple sclerosis. *J Hum Genet.* 2009;54:676–680.
150. Hafler D, Compston A, Sawcer S, et al. Risk alleles for multiple sclerosis identified by a genomewide study. *N Engl J Med.* 2007;357:851–862.
151. Lopez de Lapuente A, Pinto-Medel MJ, Astobiza I, et al. Cell-specific effects in different immune subsets associated with SOCS1 genotypes in multiple sclerosis. *Mult Scler.* 2015;21:1498–1512.
152. Durelli L, Conti L, Clerico M, et al. T-helper 17 cells expand in multiple sclerosis and are inhibited by interferon- β . *J Neurosci Res.* 2009;65:499–509.
153. Takahashi R, Nishimoto S, Muto G, et al. SOCS1 is essential for regulatory T cell functions by preventing loss of Foxp3 expression as well as IFN- γ and IL-17A production. *J Exp Med.* 2011;208:2055–2067.