



Antidiabetic Potentials of a Novel Polyherbal Preparation Formulated According to Principles of Siddha System of Medicine

Venkataraman Balaji, BSMS, MPhil¹,
Augustine Anne Williams, MSc, MPhil¹,
Sampath Sathish, MSc, MPhil¹, Chinnaiyan Mayilvanan, MSc¹,
Akilavalli Narasimhan, MSc, MPhil¹, and
Karundevi Balasubramanian, MSc, MPhil, PhD¹

Abstract

According to the principles of Siddha system of medicine, the following polyherbal preparation consisting of 5 plant parts in equal ratio namely, *Asparagus racemosus*, *Emblica officinalis*, *Salacia oblonga*, *Syzygium aromaticum*, and *Tinospora cordifolia* was formulated to treat experimental type 2 diabetic rats. So, using plants having aphrodisiac property in the formulation is a rational approach and first of its kind, as there have been no reports so far. Phenolics and other bioactive compounds present in polyherbal preparation may be responsible for lipid-lowering effects and strong antioxidant activity. Polyherbal preparation treatment reverted the activities of glycolytic and gluconeogenic enzymes that are disturbed in diabetic rats. It is concluded that polyherbal preparation treatment improves deranged lipid profile, antioxidant status, glycogen content, and decreases lipid peroxidation, which provides stability to membrane integrity and thus favors insulin receptor to achieve better glucose tolerance through a holistic approach.

Keywords

Siddha, type 2 diabetes, polyherbal preparation, antioxidants

Introduction

The Siddha system of medicine was framed from the results of the devoted pursuit of Siddhars, who are saints, doctors, alchemists, and mysticists all at once. Literature works of Siddhars reveal that Siddha medicine was laid on strong fundamentals. Siddha science strongly believes that what exists in the microcosm is in the macrocosm; in which man is the microcosm and Universe, the macrocosm. Cosmogonesis symbolizes that there are 5 principal elements in nature, namely, earth, water, fire, wind, and ether. They are the origin of all corporeal things, which then die out, resolving themselves again into these elements. It is clearly laid down by Siddhars that there is a very close and intimate relationship between the external world and internal man. Six tastes (sweet, sour, pungent, bitter, salty, and astringent) are formed with the selective unification of these 5 elements. Siddha science defines clearly that 3 cardinal humors—Wind, Bile, and Phlegm (called *Vali*, *Azahal*, *Aiyyam* in Siddha medicine) are responsible for a person's mental and physical qualities and dispositions. Treatment is aimed at the restoration of equilibrium among 3 humors with the help of 6 tastes. The essence of this is simply stated as

“*Food is medicine; medicine is food.*” This phrase infers that what we eat is medicine for our body; the same food is responsible for our illness too.

Our food is composed of 6 tastes, which are also made up of 5 elements namely, sweet (earth and water), sour (water and fire), pungent (air and fire), bitter (air and sky), salt (earth and fire), and astringent (earth and air).¹ Siddhars have framed certain basic guidelines regarding which food to consume and which food to avoid while consuming Siddha medicine. So, food and environment (season) play a major role in maintaining the equilibrium of our body.

Over thousands of years, Siddha system of medicine has developed various practical theories to create polyherbal

¹ University of Madras, Sekkizhar Campus, Chennai, India

Corresponding Author:

Karundevi Balasubramanian, Department of Endocrinology, Dr ALM Postgraduate Institute of Basic Medical Sciences, University of Madras, Sekkizhar Campus, Taramani, Chennai 600113, India
Email: kbala82@rediffmail.com

formulations in which multiple agents contained in one formula act synergistically.² Food stuffs possess 2 major functions. The primary function is nutritional feature (life support) and the secondary function is gustational feature (taste, flavor, and texture). A recent report suggested that antioxidant potency is the tertiary function of food.³

In Siddha system of medicine, diabetes (*Neerizhivu*) is portrayed as *Aiyyam* (symbolizes earth and water) humor derangement disease that can be neutralized by *Vali* (represents air and space) humor predominant drugs. To nullify the detrimental effects of diabetes, the proposed drug should possess either bitter or pungent or astringent taste, which will retract the deranged *Aiyyam* humor, because of its predominant *Vali* humor. So, the proposed polyherbal preparation was selected from the list of known potent plants with antioxidant, immunomodulator, and aphrodisiac activity that have predominant *Vali* humor to treat diabetes as a novel strategic option. The polyherbal preparation containing *Embllica officinalis* ribes, *Salacia oblonga*—stem and root (both having astringent taste), *Syzygium aromaticum* buds (pungent), *Tinospora cordifolia*—stem and root (bitter), *Asparagus racemosus* spears/tubers (aphrodisiac property) was formulated to treat type 2 diabetic rats more tactically. All these ingredients do not have any toxicity because these are all added in regular cuisines being consumed globally. *Asparagus* edible spears are present in local diets of Eastern countries.⁴ *Salacia* roots have been extensively consumed in Japan, the United States, and other countries as a food supplement for the prevention of obesity and diabetes.⁵ *Embllica* and clove are integral part of cuisine for many centuries.⁶ *Tinospora* is an important ingredient in every immune modulator preparations in Siddha and Ayurveda since ancient times. *A racemosus* Willd. (*Asparagaceae*) is an important medicinal plant indigenous to South Asian countries and its medicinal uses have been reported in the Indian and British Pharmacopoeias and also in Indian traditional systems of medicine such as Siddha, Ayurveda, and Unani.⁷ Pharmacological studies with animals have manifested the potency of *A racemosus* extract as an antioxidant,⁸ adaptogen,⁹ and with the strongest focus being on its ability in modulating the immune system.^{10,11} *Embllica* fruit has many pharmacological activities for the treatment of a number of diseases and is a constituent of many hepatoprotective formulations¹²; it possesses antidiabetic activity¹³ and shows presence of tannins, lignans, flavonoids, and alkaloids.¹⁴ *S oblonga*, which has been traditionally used in Siddha medicine is effective for the prevention and treatment of diabetes¹⁵ and possesses α -glucosidase inhibitors such as salacinol^{16,17} and kotakanol.¹⁸ Cloves are the dried flower buds of *S aromaticum* (L.) Merr. & Perry—a tree of the myrtle family (*Myrtaceae*). Phytochemical studies indicate that the clove contains free eugenol, eugenol acetate, caryophyllene, sesquiterpene ester, phenyl propanoid,¹⁹ and β -caryophyllene.²⁰ Oil from clove reportedly modulated physiological responses in streptozotocin-induced diabetic rats.²¹ Food seasoning spice mixtures containing clove have shown to improve glucose metabolism and lipid profile in fructose fed hyper-insulinemic male Wistar rats.²² *T cordifolia* Miers (*Menispermaceae*) has been

known to promote longevity and increase the body's resistance against various diseases.²³ It has also been extensively reported as a general tonic, hepatoprotective, and antidiabetic agent.²⁴ Oral administration of either alcoholic or aqueous extract of *Tinospora* is reported to have hypoglycemic activity in different animal models.²⁵ As low testosterone level is commonly associated with diabetes,²⁶ a herb with aphrodisiac property was chosen in the formulation. This is a rational approach and first of its kind.

It is hypothesized that polyherbal preparation may improve glucose tolerance by modulating hormonal levels, lipid profile, liver function, free radical generation and antioxidant, glycolytic, and gluconeogenic enzymes activities in skeletal muscle and liver of type 2 diabetic male rats.

Materials and Methods

Preparation of Polyherbal Preparation

All the 5 plant parts were purchased from the local market, cleaned, dried in shade for a week. They were then powdered by pulverizer separately, sieved, and mixed in equal quantities. These plant parts were identified by Dr D. Aravindan, an Asst Prof, Department of Medicinal Botany, National Institute of Siddha, Chennai, India and specimen samples were deposited at the department.

Chemicals

All fine chemicals, including streptozotocin, were purchased from Sigma Aldrich (St. Louis, MO, USA) and SRL (Mumbai, India). All other chemicals used were of good quality and analytical grade and obtained from SRL (Mumbai, India). Biochemical kits used in the present study were purchased from Spinreact (Girona, Spain).

Induction of Type 2 Diabetes

Rats were made diabetic (type 2) by a single intraperitoneal injection of streptozotocin (35 mg/kg body weight) after 30 days of high-fat diet containing cholesterol 2 g, cholic acid 1 g, coconut oil 30 mL, standard rat feed 100 g, and 25% fructose feeding through drinking water²⁷ and were continued till the end of the study (30 days). The low dose of streptozotocin was given to generate a slight trauma to beta cells of pancreas to mimic the condition of chronic hypoinsulinemic insulin resistance condition.

Experimental Design

Rats were divided into the following groups:

Group 1: Control.

Group 2: Rats were made diabetic (type 2) by a single intraperitoneal injection of streptozotocin (35 mg/kg body weight) after 30 days of high-fat diet and high fructose feeding through drinking water (25%).

Group 3: Diabetic (type 2) rats were treated with polyherbal preparation (500 mg/kg body weight) twice a day (8.00 AM and 8.00 PM) for 30 days after 5 days of streptozotocin treatment.

Group 4: Diabetic (type 2) rats were treated with metformin (50 mg/kg bodyweight per day) for 30 days after 5 days of streptozotocin treatment

Group 5: Control rats were treated with polyherbal preparation (500 mg/kg body weight) twice a day (8.00 AM and 8.00 PM) for 30 days.

At the end of the 30-day treatment, animals were anesthetized with sodium thiopentone (40 mg/kg body weight), blood was collected through cardiac puncture, sera were separated and stored at -80°C until the assay of hormones, and 20 mL of isotonic sodium chloride solution was perfused through the left ventricle to clear blood from the organs. Liver and gastrocnemius muscle were excised and used for the assay of various parameters. Tissues were minced and homogenized (10% w/v) with 0.1 M Tris-HCl buffer (pH 7.4) in ice-cold condition. The homogenates were centrifuged at $1000 \times g$ for 10 minutes and then the supernatants were separated and used for the assay of various parameters.

Oral Glucose Tolerance Test and Plasma Glucose

Rats of all groups were subjected to oral glucose tolerance test 2 days prior to killing. All animals were fasted overnight and the following morning, they were subjected to oral glucose tolerance test by giving an oral dose of glucose (1 mL/100 g body weight, 50% w/v glucose solution) after collecting blood by puncturing the orbital sinus with the help of heparinized microhematocrit capillary tubes for estimating fasting blood glucose and insulin. Blood samples were collected subsequently at 60, 120, and 180 minutes and centrifuged for 10 minutes at $800 \times g$ at 4°C within 30 minutes to prevent autoglycolysis by leukocytes. Plasma glucose was estimated by glucose oxidase-peroxidase method (CPC Diagnostics, Spain). Results are expressed as mg/dL.

Radioimmunoassay of Insulin and Testosterone

Serum insulin was assayed using ^{125}I -labeled radioimmunoassay kit obtained from DiaSorin (Saluggia, Italy). The limit of detection is 3.0 $\mu\text{IU/mL}$. The percentage cross-reactivity of insulin antibody to rat insulin was 100% and to C-peptide was $<0.01\%$. Intra-assay coefficient of variation was $<10.6\%$ and inter-assay coefficient of variation was $<10.8\%$. Results are expressed as $\mu\text{IU/mL}$.

Serum testosterone was assayed using liquid-phase radioimmunoassay kit obtained from DiaSorin. The limit of detection is 0.02 ng/mL. Cross-reactivity of testosterone antiserum to other steroids such as 5- α -dihydrotestosterone and androstenedione is 6.9% and 1.1%, respectively. Intra-assay coefficient of variation was $<8\%$ and inter-assay coefficient of variation was $<7.6\%$. Results are expressed as ng/mL.

Liver Function Test and Lipid Profile

The activities of serum aspartate aminotransferase, alanine aminotransferase, alkaline phosphatase, bilirubin (total and direct), total cholesterol, triglycerides, low-density lipoprotein cholesterol, high-density lipoprotein cholesterol, urea, and creatinine were estimated using commercially available kits according to manufacturer's instructions.

Determination of Lipid Peroxidation and Reactive Oxygen Species Generation

Lipid peroxidation was measured by the previously published method.²⁸ The malondialdehyde content of the sample is expressed as nanomoles of malondialdehyde formed/min/mg protein. Hydrogen peroxide generation was assessed by the spectrophotometric method.²⁹ The hydrogen peroxide content of the sample is expressed as $\mu\text{mol/min/mg}$ protein. Hydroxyl radical (OH^*) production was quantified³⁰ and expressed as $\mu\text{mol/min/mg}$ protein.

Determination of Antioxidant Enzymes

The activities of superoxide dismutase,³¹ catalase,³² glutathione peroxidase,³³ glutathione-S-transferase,³⁴ glutathione reductase,³⁵ and reduced glutathione³⁶ were assayed by previously published method. Protein was estimated by the method of Lowry et al.³⁷

Estimation of Glycolytic, Gluconeogenic Enzyme, and Glucose-6-Phosphate Dehydrogenase

Hexokinase was assayed³⁸ by determining the rate of disappearance of glucose at 37°C in a reaction mixture. Hexokinase activity is expressed as nanomoles of glucose-6-phosphate liberated/min/mg protein. Aldolase was estimated³⁹ and its activity is expressed as nanomoles of glyceraldehyde liberated/min/mg protein. Glucose-6-phosphatase was assayed⁴⁰ and its activity is expressed as nanomoles of inorganic phosphorus liberated/min/mg protein. Glucose-6-phosphate dehydrogenase was assayed⁴¹ and its activity is expressed as units/mg protein.

Estimation of Markers of Kidney Function, Calcium, Lactate Dehydrogenase, and γ -Glutamyl Transpeptidase

Urea, creatinine, calcium, lactate dehydrogenase, and γ -glutamyl transpeptidase were estimated according to the manufacturer's instructions using biochemical kits from Spinreact, Spain.

Estimation of Glycogen Content

Glycogen was estimated by the method of Hassid and Abraham.⁴² The amount of glycogen is expressed as mg/g wet tissue.

Statistical Analysis

Data were subjected to statistical analysis using one-way analysis of variance and Duncan's multiple range test to assess the significance of individual variations between the control and treatment groups using a computer based software (SPSS 7.5 for Windows student version) and expressed as mean \pm standard error of the mean. In Duncan's test, the significance was considered at the level of $P < .05$.

Results

Fasting Blood Glucose and Oral Glucose Tolerance Test

The fasting blood glucose level was elevated substantially (Figure 1A) in diabetic group and it was significantly decreased when treated with polyherbal preparation. In metformin-treated group also blood glucose level was partially ablated. From the

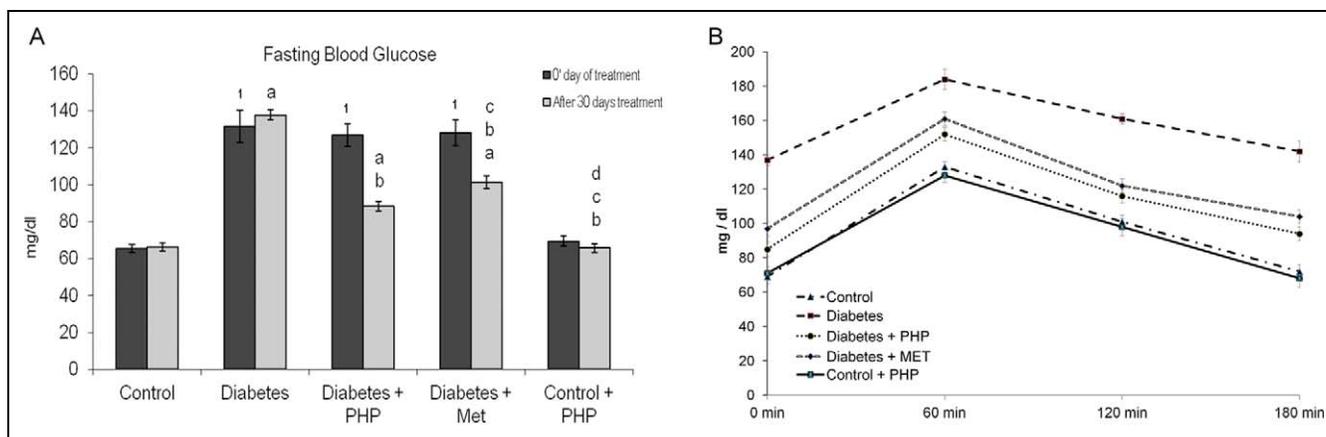


Figure 1. (A) Effect of polyherbal preparation (PHP) on fasting blood glucose of type 2 diabetic adult male rats. Each bar represents mean \pm standard error of the mean (SEM) of 6 animals. Significance at $P < .05$; (1) compared with control; (a) compared with control; (b) compared with diabetes control; (c) compared with diabetes + polyherbal preparation; (d) compared with diabetes + metformin. (B) Effect of polyherbal preparation on oral glucose tolerance of type 2 diabetic adult male rats. After overnight fasting, blood glucose was checked prior to subjecting the animals to an oral dose of glucose (5 g/kg body weight). Blood samples were collected subsequently at 60, 120, and 180 minutes and centrifuged to obtain plasma. Plasma glucose was estimated by glucose oxidase–peroxidase method. Each value represents mean \pm SEM of 6 animals. Significance at $P < .05$

results of oral glucose tolerance test (Figure 1B), it is evident that treatment with polyherbal preparation significantly reduced hyperglycemic excursions than metformin and showed improved tolerance to glucose at all time points.

Effects of Polyherbal Preparation on Serum Insulin and Testosterone

In the diabetic rats, insulin and testosterone levels were decreased. Administration of polyherbal preparation significantly augmented the serum insulin and testosterone levels when compared with type 2 diabetic rats (Figure 2A and B).

Effect of Polyherbal Preparation on Liver Function Markers and Lipid Profile

Diabetic rats showed a significant increase in the levels of serum aspartate aminotransferase, alanine aminotransferase, alkaline phosphatase, bilirubin, cholesterol, triglycerides, and low-density lipoprotein cholesterol and showed a significant decrease in high-density lipoprotein cholesterol level, whereas polyherbal preparation treatment reversed the same effectively (Tables 1 and 2).

Effects of Polyherbal Preparation on Lipid Peroxidation and Reactive Oxygen Species in the Liver and Gastrocnemius Muscle of Type 2 Diabetic Rat

Free radicals and lipid peroxidation in the liver and gastrocnemius muscle were found to be significantly elevated in diabetic rats when compared with control rats. Treatment with polyherbal preparation and metformin proved to be beneficial in reducing the free radical production and lipid peroxidation in the tissues studied (Table 3).

Effects of Polyherbal Preparation on Antioxidant, Glycolytic, and Gluconeogenic Enzyme Activities

Decreased levels of antioxidants were observed in liver and gastrocnemius muscle of type 2 diabetic rats. These derangements were restored significantly towards near normal level in polyherbal preparation- and metformin-treated diabetic rats (Tables 4 and 5). Activities of glycolytic enzymes were reduced in diabetic rats. Treatment with polyherbal preparation and metformin considerably reversed the same in both liver and gastrocnemius muscle (Table 6). Table 7 depicts the activities of glucose-6-phosphatase and glucose-6-phosphate dehydrogenase in normal control and experimental rats. These enzymes were significantly decreased in diabetic rats. Oral administration of polyherbal preparation to diabetic rats restored near normal activities of these enzymes.

Effects of Polyherbal Preparation on Glycogen Content in the Liver and Gastrocnemius Muscle of Type 2 Diabetic Rat

Type 2 diabetes lowered the glycogen concentration in liver and gastrocnemius, but treatment with polyherbal preparation was able to restore the same better than treatment with metformin.

Effects of Polyherbal Preparation on Kidney Markers, Calcium, Lactate Dehydrogenase, and Gamma Glutamyl Transpeptidase

In comparison with control rats, the urea and creatinine levels were increased significantly in diabetic rats. After treatment with polyherbal preparation and metformin, the same

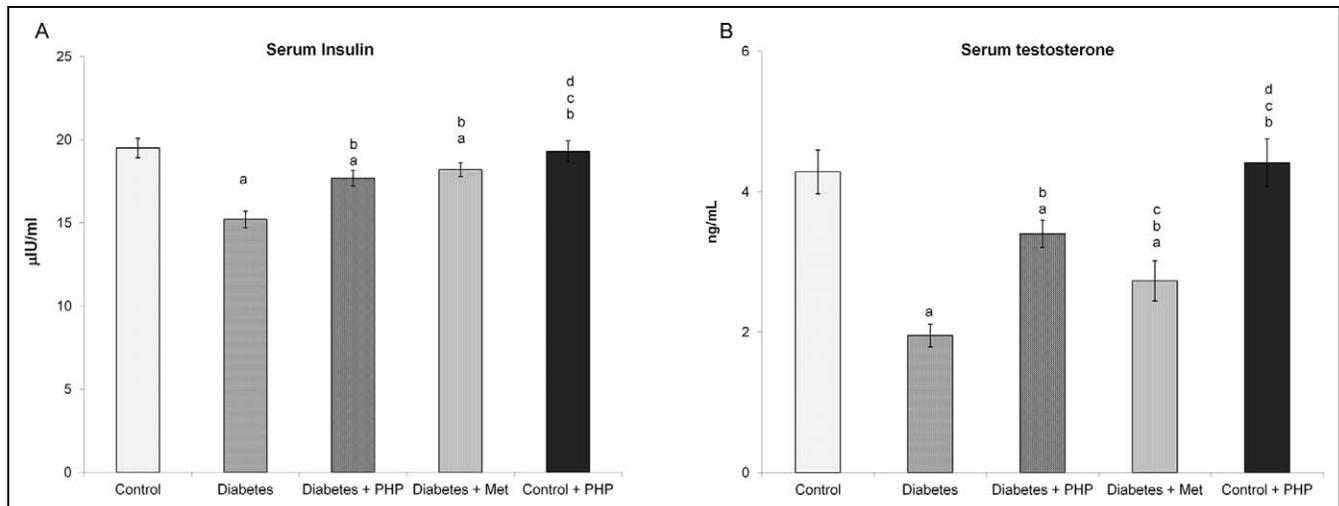


Figure 2. (A) Effect of polyherbal preparation on serum insulin level of type 2 diabetic adult male rats. Each bar represents mean \pm standard error of the mean (SEM) of 6 animals. Significance at $P < .05$: (a) compared with control; (b) compared with diabetes control; (c) compared with diabetes + polyherbal preparation (PHP); (d) compared with diabetes + metformin. (B) Effect of polyherbal preparation on serum testosterone level of type 2 diabetic adult male rats. Each bar represents mean \pm SEM of 6 animals. Significance at $P < .05$: (a) compared with control; (b) compared with diabetes control; (c) compared with diabetes + polyherbal preparation; (d) compared with diabetes + metformin

Table 1. Effect of Polyherbal Preparation on Liver Function Test of Type 2 Diabetic Male Rats*

Group	Alkaline Phosphatase (U/L)	Serum Aspartate Aminotransferase (U/L)	Alanine Aminotransferase (U/L)	Bilirubin (Total)	Bilirubin (Direct)
Normal control	404.85 \pm 7.46	114.83 \pm 1.86	45.87 \pm 2.58	1.39 \pm 0.18	0.47 \pm 0.1
Diabetes	888.66 \pm 13.49 ^a	159.73 \pm 2.77 ^a	146.45 \pm 6.16 ^a	8.38 \pm 1.07 ^a	1.28 \pm 0.15 ^a
Diabetes + polyherbal preparation	583.97 \pm 9.22 ^{a,b}	133.35 \pm 2.46 ^{a,b}	82.88 \pm 2.39 ^{a,b}	3.92 \pm 0.38 ^{a,b}	0.81 \pm 0.03 ^{a,b}
Diabetes + metformin	705.57 \pm 11.02 ^{a,b,c}	134.32 \pm 1.91 ^{a,b,c}	111.56 \pm 6.11 ^{a,b,c}	5.48 \pm 0.31 ^{a,b}	0.88 \pm 0.07 ^{a,b}
Control + polyherbal preparation	397.13 \pm 7.38 ^{b,c,d}	117.78 \pm 2.60 ^{b,c,d}	47.48 \pm 3.08 ^{b,c,d}	1.96 \pm 0.31 ^{b,c,d}	0.4 \pm 0.05 ^{b,c,d}

*Each group represents mean \pm standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

Table 2. Effect of Polyherbal Preparation on Lipid Profile of Type 2 Diabetic Male Rats*

Group	Cholesterol (mg/dL)	Triglycerides (mg/dL)	Low-Density Lipoprotein Cholesterol (mg/dL)	High-Density Lipoprotein Cholesterol (mg/dL)
Normal control	50.16 \pm 7.20	82.43 \pm 2.85	35.01 \pm 2.36	43.29 \pm 2.17
Diabetes	190.75 \pm 8.97 ^a	293.98 \pm 4.72 ^a	166.07 \pm 13.84 ^a	29.92 \pm 2.30 ^a
Diabetes + polyherbal preparation	132.25 \pm 9.42 ^{a,b}	172.73 \pm 3.03 ^{a,b}	93.70 \pm 3.63 ^{a,b}	36 \pm 1.45 ^{a,b}
Diabetes + metformin	139.28 \pm 8.85 ^{a,b}	205.5 \pm 3.87 ^{a,b,c}	123.32 \pm 4.01 ^{a,b,c}	31.05 \pm 1.39 ^{a,b}
Control + polyherbal preparation	52.32 \pm 7.12 ^{b,c,d}	81.67 \pm 2.77 ^{b,c,d}	38.67 \pm 3.52 ^{b,c,d}	43.28 \pm 1.26 ^{b,c,d}

*Each group represents mean \pm standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

were significantly reduced compared with those in untreated diabetic rats (Table 8). The rise in calcium level was accompanied with significant increase in lactate dehydrogenase and γ -glutamyl transpeptidase levels in diabetic rats than those in control rats. Treatment with polyherbal preparation and metformin decreased the same compared with those in diabetic rats. Administration of polyherbal preparation to normal rats produced no significant change in these parameters (Table 8).

Discussion

Natural products drug industry has contributed nearly half of all small molecules approved in this decade. It has been suggested that the current drug discovery approach of finding “new entity drugs,” if shifted to “combining existing agents” may be helpful.⁴³ It is generally accepted that high-fat diets can be used to generate a valid rodent model for the metabolic syndrome with insulin resistance and compromised beta-cell function.⁴⁴ In the

Table 3. Effect of Polyherbal Preparation on Lipid Peroxidation and Free Radical Generation in Type 2 Diabetic Male Rats*

Group	Lipid Peroxidation (nmoles of Malondialdehyde [MDA] Formed/min/mg Protein)		Hydrogen Peroxide Generation ($\mu\text{mol}/\text{min}/\text{mg}$ Protein)		Hydroxy Radical Generation ($\mu\text{mol}/\text{min}/\text{mg}$ Protein)	
	Liver	Gastrocnemius	Liver	Gastrocnemius	Liver	Gastrocnemius
Normal control	128.94 \pm 2.05	67.28 \pm 4.28	22.81 \pm 0.50	9.03 \pm 0.26	381.11 \pm 6.71	194.21 \pm 5.73
Diabetes	188.83 \pm 3.94 ^a	155.19 \pm 2.29 ^a	57.48 \pm 1.78 ^a	18.75 \pm 0.92 ^a	488.68 \pm 10.40 ^a	285.10 \pm 10.97 ^a
Diabetes + polyherbal preparation	159.55 \pm 2.57 ^{a,b}	98.84 \pm 2.01 ^{a,b}	33.41 \pm 1.16 ^{a,b}	13.10 \pm 0.27 ^{a,b}	407.37 \pm 6.79 ^{a,b}	219.88 \pm 5.04 ^{a,b}
Diabetes + metformin	174.49 \pm 4.58 ^{a,b,c}	124.17 \pm 1.98 ^{a,b}	43.93 \pm 1.48 ^{a,b,c}	15.75 \pm 0.44 ^{a,b,c}	448.69 \pm 5.16 ^{a,c}	252.85 \pm 4.22 ^{a,b,c}
Control + polyherbal preparation	123.23 \pm 1.74 ^{b,c,d}	69.12 \pm 1.30 ^{b,c,d}	20.63 \pm 0.10 ^{b,c,d}	8.35 \pm 0.44 ^{b,c,d}	371.99 \pm 8.27 ^{b,c,d}	186.50 \pm 5.37 ^{b,c,d}

*Each group represents mean \pm standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

Table 4. Effect of Polyherbal Preparation on the Activities of Superoxide Dismutase, Catalase, and Glutathione Peroxidase in Type 2 Diabetic Male Rats*

Group	Superoxide Dismutase (Units ¹ /mg Protein)		Catalase (Units ² /mg Protein)		Glutathione Peroxidase (Units ³ /mg Protein)	
	Liver	Gastrocnemius	Liver	Gastrocnemius	Liver	Gastrocnemius
Normal control	25.45 \pm 1.41	15.66 \pm 0.64	53.54 \pm 2.09	32.34 \pm 2.08	6.85 \pm 0.78	3.04 \pm 0.20
Diabetes	9.01 \pm 1.08 ^a	8.16 \pm 0.19 ^a	15.19 \pm 1.28 ^a	13.56 \pm 1.28 ^a	3.23 \pm 0.39 ^a	1.71 \pm 0.11 ^a
Diabetes + polyherbal preparation	20.08 \pm 0.88 ^{a,b}	13.79 \pm 0.96 ^{a,b}	40.06 \pm 1.05 ^{a,b}	25.45 \pm 1.50 ^{a,b}	5.03 \pm 0.07 ^{a,b}	2.32 \pm 0.11 ^{a,b}
Diabetes + metformin	12.80 \pm 0.82 ^{a,b,c}	10.64 \pm 0.70 ^{a,b,c}	32.15 \pm 2.54 ^{a,b,c}	20.12 \pm 1.35 ^{a,b,c}	3.55 \pm 0.17 ^{a,c}	1.81 \pm 0.02 ^a
Control + polyherbal preparation	28.52 \pm 1.11 ^{b,c,d}	16.19 \pm 1.05 ^{b,c,d}	55.70 \pm 4.21 ^{b,c,d}	33.28 \pm 1.14 ^{b,c,d}	7.17 \pm 0.45 ^{b,c,d}	5.54 \pm 0.23 ^{a,b,c,d}

¹One unit of activity was taken as the enzyme reaction that gave 50% inhibition of nitro blue tetrazolium chloride (NBT) reduction in 1 minute.

²Micromoles of hydrogen peroxide consumed per minute.

³Micrograms of glutathione consumed per minute.

*Each group represents mean \pm standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

Table 5. Effect of Polyherbal Preparation on the Activities of Glutathione-S-Transferase, Glutathione Reductase, and Reduced Glutathione in Type 2 Diabetic Male Rats*

Group	Glutathione-S-Transferase (Units ¹ /mg Protein)		Glutathione Reductase (Units ² /mg Protein)		Reduced Glutathione (nmoles of GSH/mg Protein)	
	Liver	Gastrocnemius	Liver	Gastrocnemius	Liver	Gastrocnemius
Normal control	63.69 \pm 2.20	34.42 \pm 2.42	6.33 \pm 0.26	3.12 \pm 0.43	10.14 \pm 0.12	6.42 \pm 0.12
Diabetes	18.24 \pm 2.80 ^a	7.16 \pm 0.43 ^a	2.80 \pm 0.14 ^a	1.41 \pm 0.14 ^a	3.22 \pm 0.50 ^a	3.01 \pm 0.17 ^a
Diabetes + polyherbal preparation	51.73 \pm 1.84 ^{a,b}	27.07 \pm 0.78 ^{a,b}	4.54 \pm 0.2 ^{a,b}	1.99 \pm 0.08 ^{a,b}	6.56 \pm 0.48 ^{a,b}	4.76 \pm 0.14 ^{a,b}
Diabetes + metformin	37.27 \pm 2.92 ^{a,b,c}	24.41 \pm 1.38 ^{a,b}	3.94 \pm 0.2 ^{a,b,c}	1.73 \pm 0.08 ^{a,b,c}	5.30 \pm 0.17 ^{a,c}	4.34 \pm 0.04 ^{a,b,c}
Control + polyherbal preparation	62.15 \pm 1.74 ^{b,c,d}	36.96 \pm 1.25 ^{b,c,d}	6.23 \pm 0.54 ^{b,c,d}	2.43 \pm 0.17 ^{b,c,d}	10.82 \pm 0.15 ^{b,c,d}	6.94 \pm 0.27 ^{b,c,d}

¹Micromoles of 1-chloro-2,4-dinitrobenzene (CDNB)-glutathione conjugate formed per minute.

²Nanomoles of glutathione disulfide (GSSG) reduced per minute at 37°C.

*Each group represents mean \pm standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

present study, improved fasting blood glucose level and better tolerance for glucose achieved in polyherbal preparation-treated group may be because of increased insulin sensitivity

in type 2 diabetic rats. Enhanced testosterone level observed in polyherbal preparation-treated rats may partly be responsible for increased sensitivity to insulin.⁴⁵ In accordance with the

Table 6. Effect of Polyherbal Preparation on Activity of Glycolytic Enzymes of Type 2 Diabetic Male Rats*

Group	Hexokinase (nmoles of Glucose-6-Phosphate Formed/min/mg Protein)		Aldolase (nmoles of Glyceraldehyde Formed/min/mg Protein)	
	Liver	Gastrocnemius	Liver	Gastrocnemius
Normal control	12.25 ± 0.59	17.10 ± 0.83	121.06 ± 5.73	61.23 ± 3.04
Diabetes	4.26 ± 0.25 ^a	7.73 ± 0.14 ^a	56.40 ± 2.35 ^a	22.53 ± 1.97 ^a
Diabetes + polyherbal preparation	8.87 ± 0.24 ^{a,b}	12.62 ± 0.71 ^{a,b}	99.65 ± 1.58 ^{a,b}	44.77 ± 2.32 ^{a,b}
Diabetes + metformin	5.95 ± 0.26 ^{a,b,c}	10.27 ± 0.20 ^{a,b}	83.28 ± 2.55 ^{a,b,c}	32.20 ± 2.12 ^{a,b,c}
Control + polyherbal preparation	13.92 ± 0.71 ^{a,b,c,d}	17.21 ± 0.84 ^{b,c,d}	123.6 ± 4.04 ^{b,c,d}	67.19 ± 3.88 ^{b,c,d}

*Each group represents mean ± standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

Table 7. Effect of Polyherbal Preparation on Glucose-6-Phosphatase, Glucose-6-Phosphate Dehydrogenase Activities, and Glycogen Content of Type 2 Diabetic Male Rats*

Group	Glucose-6-Phosphatase (nmoles of Inorganic Phosphorous [Pi] Liberated/min/mg Protein)		Glucose-6-Phosphate Dehydrogenase (units/mg Protein)	Glycogen Content (mg/g Wet Tissue)	
	Liver	Gastrocnemius		Liver	Gastrocnemius
Normal control	1075.13 ± 42.20	239.13 ± 13.04	8.02 ± 1.07	65.73 ± 2.46	43.35 ± 1.89
Diabetes	2001.45 ± 41.71 ^a	551.78 ± 36.59 ^a	3.04 ± 0.51 ^a	28.54 ± 2.04 ^a	14.07 ± 1.71 ^a
Diabetes + polyherbal preparation	1330.09 ± 30.54 ^{a,b}	354.63 ± 35.10 ^{a,b}	6.01 ± 1.06 ^{a,b}	48.70 ± 1.55 ^{a,b}	33.83 ± 1.09 ^{a,b}
Diabetes + metformin	1557.32 ± 31.03 ^{a,b,c}	448.43 ± 42.69 ^{a,b,c}	5.84 ± 1.31 ^{a,b}	43.95 ± 1.39 ^{a,b}	24.38 ± 1.13 ^{a,b,c}
Control + polyherbal preparation	1120.68 ± 22.19 ^{b,c,d}	214.59 ± 14.93 ^{b,c,d}	9.43 ± 1.67 ^{b,c,d}	69.70 ± 2.46 ^{b,c,d}	45.76 ± 2.33 ^{b,c,d}

*Each group represents mean ± standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

Table 8. Effect of Polyherbal Preparation on Urea, Creatinine, Calcium, Lactate Dehydrogenase, and γ -Glutamyl Transpeptidase on Type 2 Diabetic Male Rats*

Group	Urea (mg/dL)	Creatinine (mg/dL)	Calcium (mEq/L)	Lactate Dehydrogenase (IU/L)	γ -Glutamyl Transpeptidase (IU/L)
Normal control	26.78 ± 1.77	0.38 ± 0.013	20.02 ± 1.94	564.18 ± 22.3	2.86 ± 0.13
Diabetes	65.58 ± 2.89 ^a	1.67 ± 0.028 ^a	34.60 ± 1.08 ^a	2736.6 ± 160.96 ^a	14.99 ± 1.58 ^a
Diabetes + polyherbal preparation	43.43 ± 1.66 ^{a,b}	0.51 ± 0.034 ^{a,b}	23.46 ± 1.32 ^b	1123.61 ± 71.86 ^{a,b}	6.45 ± 0.50 ^{a,b}
Diabetes + metformin	49.77 ± 2.65 ^{a,b,c}	0.58 ± 0.028 ^{a,b,c}	28.6 ± 1.19 ^{a,b,c}	1710.5 ± 47.5 ^{a,b,c}	8.18 ± 0.34 ^{a,b,c}
Control + polyherbal preparation	27.62 ± 1.26 ^{b,c,d}	0.327 ± 0.02 ^{b,c,d}	20.74 ± 1.3 ^{b,d}	755.5 ± 43.06 ^{b,c,d}	3.53 ± 0.21 ^{b,c,d}

*Each group represents mean ± standard error of the mean of 6 animals. Significance at $P < .05$: ^acompared with control, ^bcompared with diabetes control, ^ccompared with diabetes + polyherbal preparation, ^dcompared with diabetes + metformin.

present study, polyphenols, phenolic acids, and tannins from strawberry and apple have showed substantial inhibition on both glucose uptake and transport in *Caco-2* intestinal cell monolayers.⁴⁶ Polyphenols from green tea interact with sodium glucose transporter-1 as antagonist-like molecules, controlling the dietary glucose uptake in the intestinal tract,⁴⁷ which hampers the rapid increase of plasma glucose after glucose load, and this may be responsible for the improved glucose tolerance as seen in the current study.

Optimal level of insulin and sex steroids is essential for the regulation of glucose homeostasis. Low testosterone level was reported in obesity with insulin resistance suggesting increased

risk for type 2 diabetes and associated complications.⁴⁸ Increased testosterone observed in the polyherbal preparation-treated group can be attributed to aphrodisiac property of Asparagus, an age-old claim in Siddha system of medicine. Enhanced penile erection index and reduced hesitation time are the observed effects of Asparagus extracts⁴⁹ and dose-dependent proliferation of *LNCaP* cells with *T cordifolia* suggested that androgenic compounds present in the plant appear to act via androgen receptor.⁵⁰ Clove extract was shown to increase 3- β -hydroxysteroid dehydrogenase and 17- β -hydroxysteroid dehydrogenase activities and serum testosterone level.⁵¹ All these reports reinforce the folklore aphrodisiac

claim of Asparagus, clove, and Tinospora and provide a scientific basis for their traditional usage.

Increased levels of aspartate transaminase and alanine transaminase enzymes indicate increased permeability and damage and/or necrosis of hepatocytes⁵² and spillage of these enzymes into blood results in elevation of these enzymes in diabetic condition. Polyherbal preparation treatment was effective in regulating the liver marker enzymes. Tannoid principles of *Emblica* when given to iron overload-induced oxidative stress rats resulted in the reduction of lipid peroxidation and restoration of the deranged activities of liver marker enzymes toward normalcy.⁵³ The increased bilirubin level in diabetic rats may be attributed to fatty liver induced by high fat and fructose diet. Polyherbal preparation treatment restored the above parameters to control levels by decreasing the triglycerides and cholesterol levels and improving liver function.

Free radical generation and oxidative stress can be responsible for accumulation of lipids and deranged antioxidant status. *S oblonga*, which contains compounds such as peroxisome proliferator-activated receptors- α agonists (enhances uptake and beta-oxidation of fatty acids in liver⁵⁴) was shown to normalize fatty liver and triglycerides.^{55,56} Another polyphenol-containing plant, *E officinalis* was shown to regulate lipid profile in rat model of metabolic syndrome.⁵⁷ Fructose feeding leads to hypertriglyceridemia by increasing the formation of glycerol-3-phosphate, a precursor of lipid synthesis.⁵⁸ Increased triglycerides demonstrate that diabetic rats had more severe insulin resistance. Polyherbal preparation may inhibit the pathway of cholesterol synthesis and increase high-density lipoprotein/low-density lipoprotein ratio and this may be because of the activation of low-density lipoprotein receptors in hepatocytes, which are responsible for utilizing low-density lipoprotein and reduce its level in the serum. Flavonoids, anthocyanins, triterpenes, tannins, and saponins present in polyherbal preparation can be helpful in preventing diet-induced body fat accumulation.⁵⁹ High concentrations of lipid peroxides and hydrogen and hydroxyl peroxides in tissues of diabetic rats were shown to increase the generation of free radicals. Aroma chemicals such as eugenol, thymol, and benzyl alcohol in the *S aromaticum* extracts, which are shown to have inhibitory effect on malonaldehyde formation by 48%,⁶⁰ may be responsible for the decreased lipid peroxidation in polyherbal preparation-treated diabetic rats. The protective role of flavonoids, tannoids, saponins, phenolics, and terpenoids⁶¹ present in polyherbal preparation is likely attributed to reduction of oxidative stress and associated complications.

Reduced activities of superoxide dismutase and catalase in diabetes may be because of increased reactive oxygen radicals.⁶² The reduction of these enzymes can result in various deleterious effects. Increased activity of these enzymes due to polyherbal preparation treatment may quench diabetes-induced free radical generation. Chronic hyperglycemia in diabetic condition increases the polyol pathway as well as advanced glycation end products formation and free radical generation rates, leading to increased oxidation of reduced glutathione. Reduced glutathione is known to protect cellular

system against the toxic effects of lipid peroxidation.⁶³ Reduced glutathione, a free radical scavenger acts as a co-substrate for glutathione peroxidase activity and a cofactor for many enzymes and form conjugates in endo- and xenobiotic reactions.⁶⁴ Glutathione peroxidase metabolizes hydrogen peroxide to water by using reduced glutathione as a hydrogen donor.⁶⁵ The significant recovery of reduced glutathione content and reduced glutathione-dependent enzyme glutathione peroxidase that was observed in polyherbal preparation-treated diabetic rats is likely due to the presence of ellagic acid, gallic acid, tannoids,⁶⁶ flavanoids, and proanthocyanidins⁶⁷ in the polyherbal preparation.

Increased glutathione reductase will in turn boost reduced glutathione levels, which would help in reducing oxidative stress.⁶⁸ Elevated glutathione (reduced) level can be induced by Tinospora and protect cellular proteins against oxidation.⁶⁹ High phenolic content in Asparagus,⁷⁰ *Emblica*,¹⁴ and clove⁶⁰ species may explain greater radical scavenging capacity of polyherbal preparation. Glycolysis is a main metabolic pathway that provides energy by using glucose. Hexokinase and aldolase are the most sensitive enzymes of glycolytic pathway in diabetic condition.⁷¹ The enhanced activity of glycolytic enzymes is suggestive of improved insulin sensitivity and glucose tolerance and the associated increased glucose utilization. The decrease in glucose-6-phosphate dehydrogenase activity in diabetic condition indicates the diminished functioning of hexose monophosphate shunt pathway and thereby impaired production of reducing equivalents. Insulin is reported to increase this enzyme activity in a dose-dependent manner.⁷² Administration of polyherbal preparation and metformin increased the activity of glucose-6-phosphate dehydrogenase. This is likely due to improved insulin sensitivity and decreased insulin resistance.

It has been reported that clove represses phosphoenolpyruvate carboxykinase and glucose-6-phosphatase gene expression by affecting the expression of a transcription factor peroxisome proliferator-activated receptor- γ coactivator-1. It is an important coactivator for gluconeogenic genes.^{73,74} A significant reduction in glucose-6-phosphatase enzyme activity observed in the present study may be correlated.

Increased levels of urea, creatinine, γ -glutamyl transpeptidase are suggestive of the renal tissue damage in type 2 diabetic rats. Reversal of the same in polyherbal preparation-treated group may be attributed to the strong curative action of *T cordifolia*.²⁵ γ -Glutamyl transpeptidase, a membrane-bound enzyme, is present in proximal renal tubule, liver, pancreas (ductules and acinar cells), and intestine. Decrease in serum γ -glutamyl transpeptidase activity with concomitant amelioration of reduced glutathione content observed in the present study suggests antioxidant action of quercetin, an active principle of *E officinalis* that neutralized the generated reactive oxygen species and thereby maintained cell membrane integrity and viability.⁷⁵

Serum calcium regulation is altered in diabetes mellitus,⁷⁶ and high serum calcium levels are associated with high levels of glucose, blood pressure and total cholesterol.⁷⁷⁻⁸⁰ Further

studies are needed to identify the mechanism behind decreased calcium level observed in polyherbal preparation-treated rats.

In diabetes, there is a decrease in liver weight because of enhanced catabolic processes such as glycogenolysis, lipolysis, and proteolysis.⁸¹ Glycogen level in muscle and liver were decreased in the absence of insulin and recovered on insulin treatment.⁸² Administration of polyherbal preparation and metformin significantly increased hepatic glycogen content compared with diabetic group, which could be because of improvement in insulin sensitivity and inhibition of glucose-6-phosphatase in the liver, thereby preventing gluconeogenesis.

Conclusion

The novel Siddha polyherbal preparation supplemented in the present study exhibited reversal of deteriorated liver marker enzymes and lipid profile of type 2 diabetic rats and showed strong antioxidant activities. Improved glucose tolerance and hepatic gluconeogenic enzymes were also achieved suggesting its potential therapeutic effect for the management of type 2 diabetes through a holistic approach. Clinical trials employing such novel herbal preparations would be of great interest and beneficial to disease management and human welfare at large.

Acknowledgments

We thank Ms L. Sheerin Banu for correcting the article for proper English usage.

Author Contributions

BV conceptualized the study, collected the data, developed the tables, analyzed the data, and wrote the first draft of the manuscript. AWA, SS, MC, and AN developed the tables and helped in statistical analysis. BK conceptualized the study, critically analyzed and discussed the data, and corrected and reviewed the article.

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This study received financial assistance from DST-PURSE (grant letter no. RPO/PURSE/2009/4063 dated December 2, 2009, Department of Science & Technology, Government of India) and UGC-SAP-DRS (sanction letter no. F-3/58/2009/(SAPIII) dated September 17, 2010, University Grants Commission, New Delhi, India) programmes.

Ethical Approval

Ethical clearance was obtained from the Institutional Animal Ethical Committee (IAEC No. 03/019/2009 dated April 1, 2009).

References

1. Thas JJ. Siddha medicine—background and principles and the application for skin diseases. *Clin Dermatol*. 2008;26:62-78.
2. Ji HF, Li XJ, Zhang HY. Natural products and drug discovery. Can thousands of years of ancient medical knowledge lead us to

- new and powerful drug combinations in the fight against cancer and dementia? *EMBO Rep*. 2009;10:194-200.
3. Niwano Y, Saito K, Yoshizaki F, Kohno M, Ozawa T. Extensive screening for herbal extracts with potent antioxidant properties. *J Clin Biochem Nutr*. 2011;48:78-84.
4. Wiboonpun N, Phuwapraisirisan P, Tip-pyang S. Identification of antioxidant compound from *Asparagus racemosus*. *Phytother Res*. 2004;18:771-773.
5. Jihong Y, Shaozhong L, Jingfeng S, et al. Effects of *Salacia chinensis* extract on reproductive outcome in rats. *Food Chemical Toxicol*. 2011;49:57-60.
6. Scartezzini P, Speroni E. Review on some plants of Indian traditional medicine with antioxidant activity. *J Ethnopharmacol*. 2000;71:23-43.
7. Sharma P, Chauhan PS, Dutt P, et al. A unique immuno-stimulant steroidal sapogenin acid from the roots of *Asparagus racemosus*. *Steroids*. 2011;76:358-364.
8. Parihar MS, Hemnani T. Experimental excitotoxicity provokes oxidative damage in mice brain and attenuation by extract of *Asparagus racemosus*. *J Neural Transm*. 2004;111:1-12.
9. Bhattacharya SK, Bhattacharya A, Chakrabarti A. Adaptogenic activity of Siotone, a polyherbal formulation of Ayurvedic rasayanas. *Indian J Exp Biol*. 2000;38:119-128.
10. Diwanay S, Chitre D, Patwardhan B. Immunoprotection by botanical drugs in cancer chemotherapy. *J Ethnopharmacol*. 2004;90:49-55.
11. Dhuley JN. Effect of some Indian herbs on macrophage functions in ochratoxin A treated mice. *J Ethnopharmacol*. 1997;58:15-20.
12. Antarkar DS, Vaidya AB, Doshi JC, et al. A double-blind clinical trial of Arogya-wardhani—an ayurvedic drug—in acute viral hepatitis. *Indian J Med Res*. 1980;72:588-593.
13. Sabu MC, Kuttan R. Anti-diabetic activity of medicinal plants and its relationship with their antioxidant property. *J Ethnopharmacol*. 2002;81:155-160.
14. Zhang YJ, Abe T, Tanaka T, Yang CR, Kouno I. Phyllanemblinins A-F, new ellagitannins from *Phyllanthus emblica*. *J Nat Prod*. 2001;64:1527-1532.
15. Wolf BW, Weisbrode SE. Safety evaluation of an extract from *Salacia oblonga*. *Food Chem Toxicol*. 2003;41:867-874.
16. Ghavami A, Johnston BD, Pinto BM. A new class of glycosidase inhibitor: synthesis of salacinol and its stereoisomers. *J Org Chem*. 2001;66:2312-2317.
17. Yoshikawa M, Morikawa T, Matsuda H, Tanabe G, Muraoka O. Absolute stereostructure of potent alpha-glucosidase inhibitor, Salacinol, with unique thiosugar sulfonium sulfate inner salt structure from *Salacia reticulata*. *Bioorg Med Chem*. 2002;10:1547-1554.
18. Yoshikawa M, Murakami T, Yashiro K, Matsuda H. Kotalanol, a potent alpha-glucosidase inhibitor with thiosugar sulfonium sulfate structure, from antidiabetic ayurvedic medicine *Salacia reticulata*. *Chem Pharm Bull (Tokyo)*. 1998;46:1339-1340.
19. Miyazawa M, Hisama M. Antimutagenic activity of phenylpropenoids from clove (*Syzygium aromaticum*). *J Agric Food Chem*. 2003;51:6413-6422.
20. Ghelardini C, Galeotti N, Di Cesare Mannelli L, Mazzanti G, Bartolini A. Local anaesthetic activity of beta-caryophyllene. *Farmaco*. 2001;56:387-389.

21. Nangle MR, Gibson TM, Cotter MA, Cameron NE. Effects of eugenol on nerve and vascular dysfunction in streptozotocin-diabetic rats. *Planta Med.* 2006;72:494-500.
22. Rajamani S, Suganthi R, Ravichandran MK, Anuradha CV. Food seasoning spices mixture improves glucose metabolism and lipid profile in fructose-fed hyperinsulinemic rats. *J Med Food.* 2005;8:502-507.
23. Bhatt AD, Bhatt NS. Indigenous drugs and liver disease. *Indian J Gastroenterol.* 1996;15:63-67.
24. Mathew S, Kuttan G. Antioxidant activity of *Tinospora cordifolia* and its usefulness in the amelioration of cyclophosphamide induced toxicity. *J Exp Clin Cancer Res.* 1997;16:407-411.
25. Stanely P, Prince M, Menon VP. Hypoglycaemic and other related actions of *Tinospora cordifolia* roots in alloxan-induced diabetic rats. *J Ethnopharmacol.* 2000;70:9-15.
26. Saad F. The role of testosterone in type 2 diabetes and metabolic syndrome in men. *Arq Bras Endocrinol Metabol.* 2009;53:901-907.
27. Nampurath GK, Mathew SP, Khanna V, Zachariah RT, Kanji S, Chamallamudi MR. Assessment of hypolipidaemic activity of three thiazolidin-4-ones in mice given high-fat diet and fructose. *Chem Biol Interact.* 2008;171:363-368.
28. Devasagayam TP, Tarachand U. Decreased lipid peroxidation in the rat kidney during gestation. *Biochem Biophys Res Commun.* 1987;145:134-138.
29. Pick E, Keisari Y. Superoxide anion and hydrogen peroxide production by chemically elicited peritoneal macrophages—induction by multiple nonphagocytic stimuli. *Cell Immunol.* 1981;59:301-318.
30. Puntarulo S, Cederbaum AI. Comparison of the ability of ferric complexes to catalyze microsomal chemiluminescence, lipid peroxidation, and hydroxyl radical generation. *Arch Biochem Biophys.* 1988;264:482-491.
31. Misra HP, Fridovich I. The role of superoxide anion in the auto-oxidation of epinephrine and a simple assay for superoxide dismutase. *J Biol Chem.* 1972;247:3170-3175.
32. Takahara S, Hamilton HB, Neel JV, Kobara TY, Ogura Y, Nishimura ET. Hypocatalasemia: a new genetic carrier state. *J Clin Invest.* 1960;39:610-619.
33. Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. *Science.* 1973;179:588-590.
34. Habig WH, Pabst MJ, Jakoby WB. Glutathione S-transferases. The first enzymatic step in mercapturic acid formation. *J Biol Chem.* 1974;249:7130-7139.
35. Blakytyn R, Harding JJ. Glycation (non-enzymic glycosylation) inactivates glutathione reductase. *Biochem J.* 1992;288 (pt 1): 303-307.
36. Sedlak J, Lindsay RH. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. *Anal Biochem.* 1968;25:192-205.
37. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem.* 1951; 193:265-275.
38. MacKenzie NM, Keeler KD. A flow microfluorimetric analysis of the binding of immunoglobulins to Fc gamma receptors on brush borders of the neonatal mouse jejunal epithelium. *Immunology.* 1984;51:529-533.
39. Thomas DB, Keeler KD. Flip-flop in the Lyt 2 phenotype of T cells from radiation chimaeras between Thy 1 congenic donor and recipient mice. *Immunology.* 1984;51:563-570.
40. Wing AM, Keele S, Margolin DI. Motor disorder and the timing of repetitive movements. *Ann N Y Acad Sci.* 1984;423:183-192.
41. Beutler E, Morrison M. Localization and characteristics of hexose 6-phosphate dehydrogenase (glucose dehydrogenase). *J Biol Chem.* 1967;242:5289-5293.
42. Hassid W, Abraham S. Chemical procedures for analysis of polysaccharides. In: S Colowick, N Kaplan, eds. *Methods in Enzymology.* Vol 3. New York, NY: Academic Press; 1957:34-36.
43. Kong DX, Li XJ, Zhang HY. Where is the hope for drug discovery? Let history tell the future. *Drug Discov Today.* 2009;14: 115-119.
44. Oakes ND, Cooney GJ, Camilleri S, Chisholm DJ, Kraegen EW. Mechanisms of liver and muscle insulin resistance induced by chronic high-fat feeding. *Diabetes.* 1997;46:1768-1774.
45. Grossmann M, Thomas MC, Panagiotopoulos S, et al. Low testosterone levels are common and associated with insulin resistance in men with diabetes. *J Clin Endocrinol Metab.* 2008;93:1834-1840.
46. Manzano S, Williamson G. Polyphenols and phenolic acids from strawberry and apple decrease glucose uptake and transport by human intestinal Caco-2 cells. *Mol Nutr Food Res.* 2010;54: 1773-1780.
47. Kobayashi Y, Suzuki M, Satsu H, et al. Green tea polyphenols inhibit the sodium-dependent glucose transporter of intestinal epithelial cells by a competitive mechanism. *J Agric Food Chem.* 2000;48:5618-5623.
48. Kapoor D, Aldred H, Clark S, Channer KS, Jones TH. Clinical and biochemical assessment of hypogonadism in men with type 2 diabetes: correlations with bioavailable testosterone and visceral adiposity. *Diabetes Care.* 2007;30:911-917.
49. Thakur M, Chauhan NS, Bhargava S, Dixit VK. A comparative study on aphrodisiac activity of some ayurvedic herbs in male albino rats. *Arch Sex Behav.* 2009;38:1009-1015.
50. Kapur P, Pereira BM, Wuttke W, Jarry H. Androgenic action of *Tinospora cordifolia* ethanolic extract in prostate cancer cell line LNCaP. *Phytomedicine.* 2009;16:679-682.
51. Mishra RK, Singh SK. Safety assessment of *Syzygium aromaticum* flower bud (clove) extract with respect to testicular function in mice. *Food Chem Toxicol.* 2008;46:3333-3338.
52. Goldberg DM, Watts C. Serum enzyme changes as evidence of liver reaction to oral alcohol. *Gastroenterology.* 1965;49:256-261.
53. Bhattacharya A, Kumar M, Ghosal S, Bhattacharya SK. Effect of bioactive tannoid principles of *Emblica officinalis* on iron-induced hepatic toxicity in rats. *Phytomedicine.* 2000;7:173-175.
54. Reddy JK, Hashimoto T. Peroxisomal beta-oxidation and peroxisome proliferator-activated receptor alpha: an adaptive metabolic system. *Annu Rev Nutr.* 2001;21:193-230.
55. Fischer M, You M, Matsumoto M, Crabb DW. Peroxisome proliferator-activated receptor alpha (PPARalpha) agonist treatment reverses PPARalpha dysfunction and abnormalities in hepatic lipid metabolism in ethanol-fed mice. *J Biol Chem.* 2003;278: 27997-28004.

56. Ye JM, Iglesias MA, Watson DG, et al. PPARalpha /gamma ragaglitazar eliminates fatty liver and enhances insulin action in fat-fed rats in the absence of hepatomegaly. *Am J Physiol Endocrinol Metab.* 2003;284:E531-E540.
57. Kim HY, Okubo T, Juneja LR, Yokozawa T. The protective role of amla (*Emblica officinalis* Gaertn.) against fructose-induced metabolic syndrome in a rat model. *Br J Nutr.* 2010;103:502-512.
58. Zavaroni I, Chen YD, Reaven GM. Studies of the mechanism of fructose-induced hypertriglyceridemia in the rat. *Metabolism.* 1982;31:1077-1083.
59. de Melo CL, Queiroz MG, Fonseca SG, et al. Oleonic acid, a natural triterpenoid improves blood glucose tolerance in normal mice and ameliorates visceral obesity in mice fed a high-fat diet. *Chem Biol Interact.* 2010;185:59-65.
60. Lee KG, Shibamoto T. Inhibition of malonaldehyde formation from blood plasma oxidation by aroma extracts and aroma components isolated from clove and eucalyptus. *Food Chem Toxicol.* 2001;39:1199-1204.
61. Bhatia IS, Bajaj KL. Chemical constituents of the seeds and bark of *Syzygium cumini*. *Planta Med.* 1975;28:346-352.
62. Wohaieb SA, Godin DV. Alterations in free radical tissue-defense mechanisms in streptozocin-induced diabetes in rat. Effects of insulin treatment. *Diabetes.* 1987;36:1014-1018.
63. Nicotera P, Orrenius S. Role of thiols in protection against biological reactive intermediates. *Adv Exp Med Biol.* 1986;197:41-51.
64. Gregus Z, Fekete T, Halász E, Klaassen CD. Lipoic acid impairs glycine conjugation of benzoic acid and renal excretion of benzoylglycine. *Drug Metab Dispos.* 1996;24:682-688.
65. Sies H. Damage to plasmid DNA by singlet oxygen and its protection. *Mutat Res.* 1993;299:183-191.
66. Monagas M, Hernández-Ledesma B, Gómez-Cordovés C, Bartolomé B. Commercial dietary ingredients from *Vitis vinifera* L. leaves and grape skins: antioxidant and chemical characterization. *J Agric Food Chem.* 2006;54:319-327.
67. Liu X, Zhao M, Luo W, Yang B, Jiang Y. Identification of volatile components in *Phyllanthus emblica* L. and their antimicrobial activity. *J Med Food.* 2009;12:423-428.
68. Meister A. Glutathione-ascorbic acid antioxidant system in animals. *J Biol Chem.* 1994;269:9397-9400.
69. Singh RP, Banerjee S, Kumar PV, Raveesha KA, Rao AR. *Tinospora cordifolia* induces enzymes of carcinogen/drug metabolism and antioxidant system, and inhibits lipid peroxidation in mice. *Phytomedicine.* 2006;13:74-84.
70. Goyal RK, Singh J, Lal H. *Asparagus racemosus*—an update. *Indian J Med Sci.* 2003;57:408-414.
71. Shimizu T, Parker JC, Najafi H, Matschinsky FM. Control of glucose metabolism in pancreatic beta-cells by glucokinase, hexokinase, and phosphofructokinase. Model study with cell lines derived from beta-cells. *Diabetes.* 1988;37:1524-1530.
72. Weber G, Convery HJ. Insulin: inducer of glucose 6-phosphate dehydrogenase. *Life Sci.* 1966;5:1139-1146.
73. Yoon JC, Puigserver P, Chen G, et al. Control of hepatic gluconeogenesis through the transcriptional coactivator PGC-1. *Nature.* 2001;413:131-138.
74. Boustead JN, Stadelmaier BT, Eeds AM, et al. Hepatocyte nuclear factor-4 alpha mediates the stimulatory effect of peroxisome proliferator-activated receptor gamma co-activator-1 alpha (PGC-1 alpha) on glucose-6-phosphatase catalytic subunit gene transcription in H4IIE cells. *Biochem J.* 2003;369(pt 1):17-22.
75. Sultana S, Ahmed S, Jahangir T. *Emblica officinalis* and hepatocarcinogenesis: a chemopreventive study in Wistar rats. *J Ethnopharmacol.* 2008;118:1-6.
76. Levy J, Stern Z, Gutman A, Naparstek Y, Gavin JR 3rd, Avioli LV. Plasma calcium and phosphate levels in an adult noninsulin-dependent diabetic population. *Calcif Tissue Int.* 1986;39:316-318.
77. Kesteloot H, Geboers J. Calcium and blood pressure. *Lancet.* 1982;1:813-815.
78. Lind L, Jakobsson S, Lithell H, Wengle B, Ljunghall S. Relation of serum calcium concentration to metabolic risk factors for cardiovascular disease. *BMJ.* 1988;297:960-963.
79. Jorde R, Sundsfjord J, Fitzgerald P, Bønaa KH. Serum calcium and cardiovascular risk factors and diseases: the Tromsø study. *Hypertension.* 1999;34:484-490.
80. Sun G, Vasdev S, Martin GR, Gadag V, Zhang H. Altered calcium homeostasis is correlated with abnormalities of fasting serum glucose, insulin resistance, and beta-cell function in the Newfoundland population. *Diabetes.* 2005;54:3336-3339.
81. Yadav UC, Moorthy K, Baquer NZ. Combined treatment of sodium orthovanadate and *Momordica charantia* fruit extract prevents alterations in lipid profile and lipogenic enzymes in alloxan diabetic rats. *Mol Cell Biochem.* 2005;268:111-120.
82. Vats V, Yadav SP, Grover JK. Ethanolic extract of *Ocimum sanctum* leaves partially attenuates streptozotocin-induced alterations in glycogen content and carbohydrate metabolism in rats. *J Ethnopharmacol.* 2004;90:155-160.