

Biomarkers of Angiogenesis in Colorectal Cancer

Supplementary Issue: Biomarkers for Colon Cancer

Luay Mousa¹, Mohamed E. Salem² and Sameh Mikhail¹

¹The Medstar Ohio State University Comprehensive Cancer Center-James Cancer Hospital and Solove Research Institute, Columbus, OH, USA. ²Medstar Georgetown University Hospital, Lombardi Comprehensive Cancer Center, Washington, DC, USA.

ABSTRACT: Colorectal cancer (CRC) is the third most common cancer worldwide and accounts for 10% of all new cancer diagnoses. Angiogenesis is a tightly regulated process that is mediated by a group of angiogenic factors such as vascular endothelial growth factor and its receptors. Given the widespread use of antiangiogenic agents in CRC, there has been considerable interest in the development of methods to identify novel markers that can predict outcome in the treatment of this disease with angiogenesis inhibitors. Multiple biomarkers are in various phases of development and include tissue, serum, and imaging biomarkers. The complexity of the angiogenesis pathway and the overlap between the various angiogenic factors present a significant challenge to biomarker discovery. In our review, we discuss the angiogenesis pathway and the most promising evolving concepts in biomarker discovery, as well as highlight the landmark studies that identify subgroups of patients with CRC who may preferentially benefit from angiogenesis inhibitors.

KEYWORDS: angiogenesis, colorectal cancer, imaging biomarkers, microRNA, circulating tumor cells

SUPPLEMENT: Biomarkers for Colon Cancer

CITATION: Mousa et al. Biomarkers of Angiogenesis in Colorectal Cancer. *Biomarkers in Cancer* 2015:7(S1) 13–19 doi:10.4137/BIC.S25250.

TYPE: Review

RECEIVED: August 4, 2015. **RESUBMITTED:** September 17, 2015. **ACCEPTED FOR PUBLICATION:** September 23, 2015.

ACADEMIC EDITOR: Barbara Guinn, Editor in Chief

PEER REVIEW: Twelve peer reviewers contributed to the peer review report. Reviewers' reports totaled 950 words, excluding any confidential comments to the academic editor.

FUNDING: Authors disclose no funding sources.

COMPETING INTERESTS: SM discloses advisory board fees from Bayer and consulting fees from Personalized Cancer Therapy (Perthera). Other authors disclose no potential conflicts of interest.

COPYRIGHT: © the authors, publisher and licensee Libertas Academica Limited. This is an open-access article distributed under the terms of the Creative Commons CC-BY-NC 3.0 License.

CORRESPONDENCE: sameh.mikhail@osumc.edu

Paper subject to independent expert blind peer review. All editorial decisions made by independent academic editor. Upon submission manuscript was subject to anti-plagiarism scanning. Prior to publication all authors have given signed confirmation of agreement to article publication and compliance with all applicable ethical and legal requirements, including the accuracy of author and contributor information, disclosure of competing interests and funding sources, compliance with ethical requirements relating to human and animal study participants, and compliance with any copyright requirements of third parties. This journal is a member of the Committee on Publication Ethics (COPE).

Published by Libertas Academica. Learn more about this journal.

Introduction

Colorectal cancer (CRC) is the third most common cancer worldwide and accounts for 10% of all new cancer diagnoses.¹ Twenty percent of patients diagnosed with CRC are, unfortunately, found to have metastatic disease at presentation. Furthermore, ~30% of patients who are diagnosed with early-stage CRC sooner or later develop metastatic disease.^{2,3}

Although the median overall survival (OS) of patients diagnosed with metastatic CRC (mCRC) has improved from nine months to >30 months over the past decade, the five-year OS remains at 5%–15%. The poor outcome of patients with mCRC calls for the development of new therapeutic options in addition to further refinement of current treatment strategies.^{4–6}

The growth and proliferation of mCRC depends essentially on two signaling pathways: the vascular endothelial growth factor (VEGF) and the epidermal growth factor receptor (EGFR) pathways. Fortunately, therapeutic agents have been developed to target each of these pathways; their activity is well established and they have been incorporated into routine cancer treatment worldwide. In the United States, 60%–70% of patients with mCRC will receive these agents during the course of their treatment.⁷

Anti-VEGF agents, such as bevacizumab, ziv-aflibercept, regorafenib, and ramucirumab, have all shown efficacy in

the treatment of mCRC and are currently approved by the United States Food and Drug Administration (FDA) for use in mCRC. Since the introduction of antiangiogenic agents, there has been significant interest in the identification of clinical and molecular markers to help predict which subgroup of patients will benefit from inhibition of the angiogenesis pathway.^{8–11}

Herein, we review the process of angiogenesis, paying particular attention to the discovery and use of serum, tissue, and imaging biomarkers that can potentially be used to predict patient tumor response to antiangiogenic agents (Fig. 1).

Angiogenesis Pathways

Angiogenesis is a complex process by which new blood vessels are formed from endothelial precursor. It is a critical step in cancer progression and is considered one of the hallmarks of cancer. This process is mediated through a group of ligands and receptors that work in tight regulation.^{12,13} A group of glycoproteins, including the VEGFs (VEGF-A, VEGF-B, VEGF-C, and VEGF-D) and the placental growth factor (PIGF), act as effectors of angiogenesis.^{14–17} These factors interact with three VEGF receptors (VEGFR-1, VEGFR-2, and VEGFR-3) and two neuropilin co-receptors (NRP1 and NRP2).^{18–20} The *VEGF-A* gene consists of eight exons

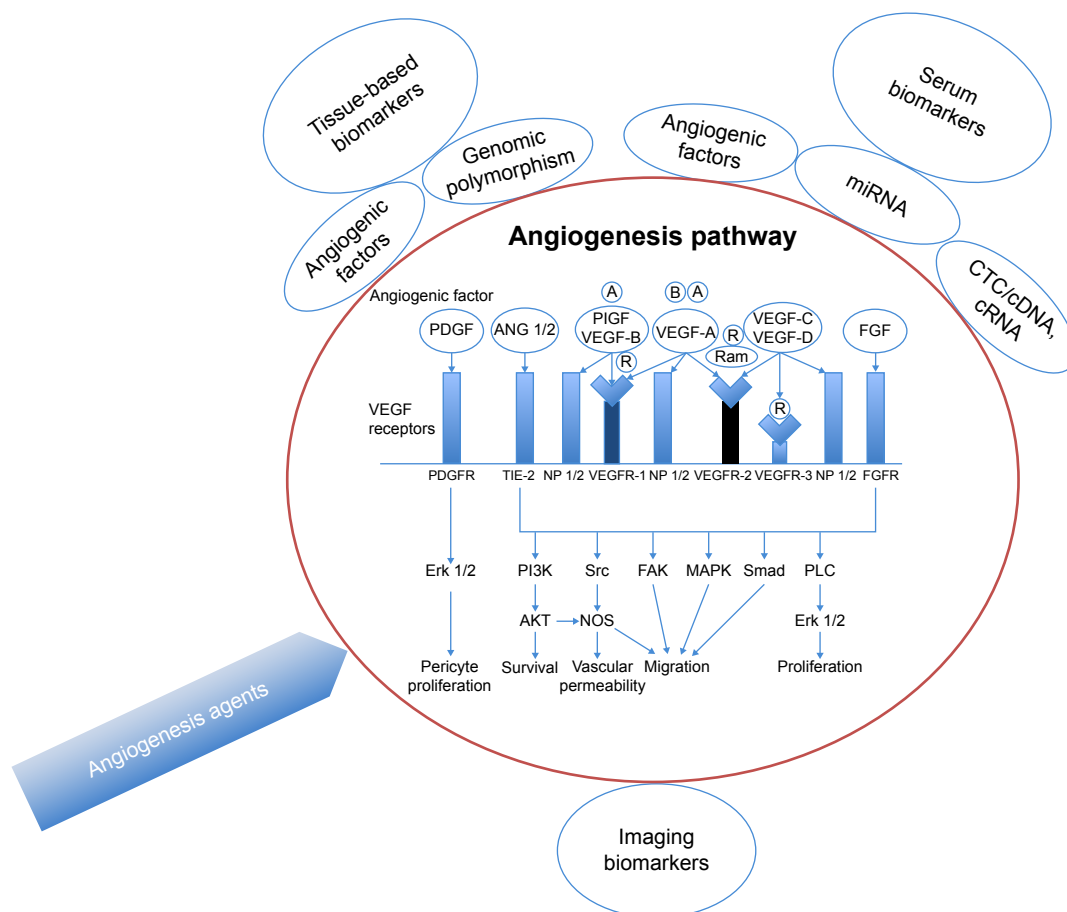


Figure 1. A schematic of the angiogenesis pathway, angiogenesis inhibitors, and the most promising biomarker techniques.

Notes: A: Aflibercept; B: bevacizumab.

Abbreviations: CTCs, circulating tumor cells; ctDNA, circulating tumor DNA; ctRNA, circulating tRNA; mRNA, microRNA; R, regorafenib; Ram, ramucirumab; VEGF, vascular endothelial growth factor.

with splice variants forming different isoforms, namely, VEGFA121, VEGFA165, VEGFA189, and VEGFA209; VEGFA165 is the most biologically active of these isoforms.²¹

The VEGFRs are tyrosine kinase receptors that are primarily located in the vascular endothelial cells.^{14,22,23} The binding of VEGF-A to VEGFR-2 is believed to be the most important activator of angiogenesis.^{12,24,25} This binding initiates a cascade of signals that result in endothelial cell proliferation and migration, increased vascular permeability, alteration of gene expression, and activation of the Ras pathway.^{14,26,27} The role of VEGFR-1 on the other hand is more complex and not fully understood. A soluble form of VEGFR-1 can act as a decoy receptor and prevent VEGF-A from binding to VEGFR-2, which, in turn, prevents signaling pathway activation. However, there is also evidence that VEGFR-1 plays an important role in the development of angiogenesis.²⁸ The third receptor VEGFR-3 is involved in lymphangiogenesis and does not bind to VEGF-A.

The neuropilins (NRP1 and NRP2) are implicated in cell guidance and increased binding of VEGF and its signaling receptors.¹⁵

Several other factors have functions that overlap with VEGF-A, including the PIGF, fibroblast growth factor (FGF), VEGF-C, VEGF-D, angiopoietin, hypoxia-inducible factor (HIF)-1 α and HIF-2 α , integrin, and platelet-derived growth factor.^{14,17}

The overlap between these factors and VEGF, and the multiple isoforms and splice variant forms of VEGF, makes assessment of individual angiogenesis pathway activation or inhibition outcomes, and thus biomarker discovery, particularly challenging.^{29,30}

Biomarkers of Angiogenesis

Tissue-based biomarkers.

Tissue vascular endothelial growth factor. The use of tissue VEGF as a predictive marker has been evaluated in several studies with conflicting results.^{28–34} Some studies indicate potential value for VEGF in the prediction of prognosis for patients with mCRC. For example, Tsai et al compared pre- and posttreatment VEGF expression by immunohistochemistry in 57 patients with mCRC who underwent treatment with 5-fluorouracil (5-FU) and irinotecan (FOLFIRI regimen)

combined with bevacizumab; results indicated that decreased peritherapeutic, low posttreatment, VEGF expressions were significant predictors of response to therapy and six-month progression-free survival (PFS).³⁵

On the other hand, Jubb et al evaluated 312 tissue samples from 813 patients enrolled in a phase III trial of irinotecan and 5-FU with or without bevacizumab. Epithelial and stromal VEGF levels, assayed by in situ hybridization, were not found to be predictive of therapy outcomes.³⁶

These results are confusing and highlight the need for further studies to determine the value of assaying tissue VEGF as a predictive marker.

Genetic polymorphisms. At least 12 studies were conducted to evaluate the predictive value of VEGF polymorphisms in patients who were receiving treatment with bevacizumab-based regimens.³⁷

Formica et al conducted a prospective study to evaluate the predictive value of *VEGF* gene polymorphisms in 40 patients with mCRC who received bevacizumab-containing first-line chemotherapy.³⁸ The study demonstrated that the *VEGF-1154G>A* polymorphism was associated with improved OS and PFS. Similarly, *VEGF405C>I* was associated with a significant improvement in OS. This observation was further confirmed by Gerger et al who also suggested that germline variants in VEGF-dependent and -independent angiogenesis genes can predict survival and tumor response in patients with mCRC treated with first-line bevacizumab and oxaliplatin-based chemotherapy.³⁹

To further explore the role of genetic polymorphisms in predicting response to angiogenesis inhibitors, investigators evaluated the role of multigene signatures as a predictive tool. Zhang et al analyzed the expression levels of *VEGFA*, *VEGFR1*, and *VEGFR2* genes in two independent colon cancer datasets and suggested that patients whose tumors expressed all these three genes at a low level had a significantly longer mean disease-free survival (DFS; 101 months, 95% confidence interval [CI], 86–116) than patients whose tumors expressed all three genes at high levels (72 months, 95% CI, 54–90). Therefore, the expression of this three-gene signature (*VEGFA*, *VEGFR1*, and *VEGFR2*) was reported to represent robust prognostic indicator in this patient population.⁴⁰

Furthermore, Zhang et al assessed the ability of this three-gene signature to predict response to bevacizumab in a cohort of colon cancer patients ($n = 14$) who had received bevacizumab treatment and for whom clinical response data were available. They found that 71% of patients who did not respond to bevacizumab expressed the three-gene signature at a low level, whereas none of those who responded exhibited a low-level signature (χ^2 test, $P = 0.02$).⁴⁰ These findings are promising but require further validation in larger prospective trials.

Other genetic polymorphisms are being investigated and include polymorphisms in *NRP1*.³⁷

Overall this is an intriguing area of research, but the role of genetic polymorphisms in the prediction of disease outcome should be further validated in larger prospective trials.

Circulating biomarkers.

Serum vascular endothelial growth factor. Analysis of biomarkers in serum represents an attractive strategy for research studies due to ease of specimen acquisition allowing for serial measurements of any biomarker of interest. Circulating VEGF levels are reported to be relevant in the prediction of outcomes of patients with solid tumors. These levels are thought to reflect VEGF-dependent tumor-mediated angiogenesis.⁴¹ However, the predictive value of baseline VEGF and/or changes in VEGF levels during or after treatment with bevacizumab remain a matter of debate.

An exploratory analysis by Duda et al evaluated VEGF, PIGF, and soluble VEGFR-1 in plasma from patients receiving bevacizumab combination treatment and suggested that soluble VEGFR-1 may predict response and toxicity to neoadjuvant bevacizumab-based chemotherapy.⁴²

The importance of baseline levels of VEGF and soluble VEGFR-2 (sVEGFR-2) as prognostic and predictive biomarkers was evaluated in two phase III studies evaluating the role of cediranib, an experimental angiogenesis inhibitor, in mCRC: the HORIZON II study randomized 860 patients to receive FOLFOX or XELOX with ($n = 502$) or without ($n = 358$) cediranib. Similarly, the HORIZON III study randomized 1422 patients to receive modified FOLFOX-6 (mFOLFOX-6) with cediranib ($n = 709$) or bevacizumab ($n = 713$).⁴³ High baseline VEGF was associated with a worse PFS in both the HORIZON II (hazard ratio [HR], 1.41; 95% CI, 1.21–1.65) and the HORIZON III (HR, 1.20; 95% CI, 1.04–1.38) studies, and a worse OS in the HORIZON II study (HR, 1.35; 95% CI, 1.12–1.63). However, sVEGFR2 did not predict PFS in either study (HR, 0.99; 95% CI, 0.85–1.15 [HORIZON II] and HR, 0.98; 95% CI, 0.86–1.13 [HORIZON III]). Similarly, sVEGFR2 did not predict OS in HORIZON II (HR, 0.92; 95% CI, 0.77–1.10) or HORIZON III (HR, 0.92; 95% CI, 0.77–1.10).

Bates et al also evaluated the predictive value of VEGF_{(165)b}, a VEGF splice isoform that binds bevacizumab. High VEGF_{(165)b} appeared to predict resistance to bevacizumab therapy but this observation was not statistically significant,⁴⁴ and further work is necessary to ascertain a true link between the two.

Although of interest, the findings highlighted in this section have not been uniformly confirmed. Therefore, this area of research remains a work in progress and further studies are needed to clarify the role of circulating VEGF as a predictive marker.

Circulating tumor cells and free nucleic acid. Circulating tumor cells (CTCs) and free nucleic acid (CTNA) detection in peripheral blood represents an attractive strategy for diagnosis and response assessment in patients with CRC, as well as to predict patient prognosis and therapeutic



outcomes (Table 1).⁴⁵ The most widely used CTC enumeration platform, Cell Search™ (Veridex LLC), is currently approved by the FDA for clinical use in CRC, breast cancer, and prostate cancer. Investigators have also become increasingly interested in the detection of CTNA (DNA or RNA) to reflect the presence of CTCs. Diehl et al⁴⁶ demonstrated that for every 100 g of tumor, 3.3 g of tumor DNA could enter the blood stream. Circulating tumor DNA may give insight into genetic and epigenetic alterations, as well as being useful for diagnosis and prediction of response to therapy. Early evidence suggests that CTC may have both prognostic and predictive values in patients with mCRC.⁴⁷ High CTC counts (≥ 3 CTC/7.5 mL) were associated with worse PFS and OS and were predictive of a worse outcome following all treatment types administered in the study. It is worth noting that ~50% of patients in this study had received bevacizumab. Interestingly, Rahbari et al⁴⁸ also demonstrated that CTC detection correlated with circulating angiogenic factors and was associated with lower levels of EGF and FGF. This observation suggests that CTCs may prove to be valuable in predicting response to antiangiogenesis agents but is currently hypothesis generating at best.

This technology is promising but continues to have several limitations. The median CTC detection rate is 35%⁴⁹ and CellSearch technology requires subjective CTC verification, not permitting single cell analysis.⁴⁵ Nevertheless, CTCs represent a promising biomarker for the prediction of treatment response in patients with cancer.

MicroRNA. MicroRNA (miRNA) is a class of small, single-stranded, noncoding RNA that can regulate the expression of multiple genes at the posttranscriptional level. They are thus involved in various cellular functions, including proliferation, apoptosis, regulation of embryonic stem cell development, and cancer cell invasion.⁵⁰ Recent studies have shown that miRNAs in the circulation are remarkably stable. This finding allows them to be robust and reliable biomarkers of cancer therapy.⁵¹ Investigators have demonstrated that miRNAs can modulate tumor angiogenesis through targeting pro-/antiangiogenic factors, including RTK signaling protein, HIF, VEGF, and EGF (Table 2). The involvement of miRNA in tumor angiogenesis has generated interest in

Table 2. Examples of miRNAs that regulate various angiogenesis proteins.

TARGET	miRNA	REFERENCE
RTK	miRNA-145	75–78
HIF	miRNA-22, 107	79,80
VEGF	miRNA-192	81
TSP-1	miRNA-182, 194	82,83
ROS	miRNA 186, 216B, 337-3p, 760	84
EGF	miRNA-121	85

Abbreviations: RTK, receptor tyrosine kinase; HIF, hypoxia-inducible growth factor; VEGF, vascular endothelial growth factor; TSP-1, thrombospondin-1; ROS, reactive oxygen species; EGF, endothelial growth factor.

exploring their utility as predictive biomarkers. miRNA-126 is one of the most studied miRNAs in mCRC.⁵⁰ It is thought to play an important role in the regulation of angiogenesis. High expression of miRNA-126 is associated with increased VEGF-A signaling in endothelial cells and therefore was thought to be a promising biomarker for antiangiogenic therapy. Hansen et al evaluated miRNA-126 as a predictive marker of outcomes in patients enrolled in the NOR-DIC ACT 1 trial.⁵² This phase 3 study evaluated the use of maintenance bevacizumab and erlotinib in patients who had stable disease or a clinical response following six months of oxaliplatin- or irinotecan-based chemotherapy plus bevacizumab. High tumor expression of miRNA-126 was significantly related to longer PFS (HR, 0.49; 95% CI, 0.29–0.84; $P = 0.009$). However, miRNA-126 did not seem to predict response rate in this study. There is still significant interest in evaluating the role of miRNAs as predictive biomarkers in CRC but their use is currently limited to the research setting.

Other angiogenesis markers. Kopetz et al investigated the efficacy of FOLFIRI (fluorouracil, leucovorin, and irinotecan) and bevacizumab in 43 patients with previously untreated mCRC. In an attempt to find potential circulating biomarkers of treatment response and therapeutic resistance, this phase II study assessed the levels of 37 different cytokines and circulating angiogenic factors (CAFs) in patient plasma using multi-bead and enzyme-linked immunosorbent assays. Levels were evaluated at baseline, during treatment, and at the time of disease progression (PD).⁵³ Elevated interleukin 8 (IL-8) levels at baseline were associated with a shorter PFS (11 vs. 15.1 months, $P = 0.03$). Furthermore, the levels of several CAFs already associated with angiogenesis and myeloid recruitment were increased prior to radiographic evidence of PD compared with baseline levels. These factors included basic FGF ($P = 0.046$), hepatocyte growth factor ($P = 0.046$), stromal-derived factor-1 ($P = 0.04$), and macrophage chemoattractant protein-3 ($P < 0.001$). These data suggest that an increase in the levels of certain pro-angiogenic cytokines and myeloid recruitment factors may represent a mechanism of resistance.⁵³

Also in 2010, Goede et al⁵⁴ demonstrated that angiotensin-2, a key regulator of vascular remodeling in

Table 1. Clinical utility of circulating tumor cells and tumor nucleic acid.

ASSAY	CLINICAL UTILITY	REFERENCE
CTC	Prognostic	47,71,72
	Predictive (ALDH1, survivin, MRP5)	73
ctDNA	Prognostic	73
ctRNA	Prognostic	74

Notes: Prognostic: biomarker that provides information on the likely course of the disease in an untreated patient; predictive: biomarker that can be used to identify subgroups of patients who are most likely to respond to a given treatment.

Abbreviations: CTCs, circulating tumor cells; ctDNA, circulating tumor DNA; ctRNA, circulating tRNA.

conjunction with VEGF, is potentially predictive of response rate, PFS, and OS.

Another potential biomarker of bevacizumab treatment outcome is CD133. Pohl et al assayed tumor tissue for germline variations on the 3'UTR-region of the CD133 gene (rs2240688, rs3130, and rs2286455) in 91 patients treated with FOLFOX or FOLFIRI plus bevacizumab. Patients who carried the CC allele in rs2286455 and rs3130 or the combination of the CT with either CT or TT experienced longer PFS (16.5 vs. 8.4 months; $P = 0.010$).⁵⁵ These results are intriguing, but need to be validated in larger studies if these markers are to be proven of true predictive rather than prognostic value.

In order to identify serum biomarkers that may predict the clinical outcomes of regorafenib, an analysis of samples obtained from the CORRECT trial was conducted.⁵⁶ The study, unfortunately, did not reveal any biomarkers that were predictive of clinical outcomes. In univariate analysis, high concentration of soluble TIE-1 was associated with improved OS, but not PFS, compared with a low concentration. In multivariable analysis, this association was, however, not statistically significant. Biomarkers such as IL-8 and PIGF demonstrated a potential prognostic value but neither marker seemed to play a role in predictive treatment response in patients treated with regorafenib.

Baseline levels of IL-8 and PIGF were found to have prognostic value for OS, although only IL-8 was also prognostic for PFS. Neither factor was predictive of OS or PFS.

In summary, the CORRECT trial biomarker analysis did not reveal any biomarkers that were predictive of clinical outcomes.

There is growing evidence to suggest that baseline and posttreatment circulating endothelial cell (CEC) and endothelial progenitor cell number and viability can predict response to antiangiogenic treatment. Willet et al demonstrated a decrease in the blood concentration of viable CECs on day 12 of bevacizumab administration,⁵⁷ compared with baseline, suggesting a possible response to treatment.

Also, in another study, a reduced percentage of viable CECs posttreatment significantly correlated with pathologic complete response ($P < 0.05$) in 32 patients with locally advanced rectal cancer receiving neoadjuvant bevacizumab in combination with standard chemoradiation therapy.⁵⁸

Taken together, these data indicate that select biomarkers appear promising, but their potential utility has to be further evaluated in well-designed prospective studies.

Imaging biomarkers. Recent developments in imaging technologies have led to significant improvements in the management of patients with CRC. Diagnostic techniques, such as diffusion-weighted imaging (DWI), fluorodeoxyglucose positron emission tomography (FDG-PET), and dynamic contrast-enhanced magnetic resonance imaging (DCE-MRI), are transitioning from bench-to-bedside and appear to be useful in providing insights into tumor biology and possibly response to treatment.^{59–61}

Functional imaging allows investigators to map the distribution of tumor mass and surrounding tissues corrected for injected dose and patient weight. FDG-PET provides information on tumor cell viability after treatment, and it has been shown that metabolic changes in response to treatment occur before any structurally detectable change (eg, tumor shrinkage). In the neoadjuvant setting, serial FDG-PET examinations may assist treatment planning. It is thought to be an important marker in rectal cancer due to the fact that sequential FDG-PET after neoadjuvant chemoradiation can predict response to therapy and has been shown to be an independent predictor of DFS and OS.^{62,63}

Whole-body DWI is being explored in mCRC but there is currently no evidence to suggest that this technique can replace PET/CT.⁶⁴ DWI has been shown to be feasible as an early marker of treatment response because cell death and vascular alterations typically occur before tumor size changes.⁶⁵

Another potential prognostic biomarker in mCRC is the dynamic contrast-enhanced CT (DCE-CT) or perfusion CT. This technology uses quantitative vascular parameters such as blood flow (BF), blood volume, and mean transient time. Goh et al reported that tumor BF was significantly lower in primary tumors of patients who ultimately developed metastatic disease,⁶⁶ whereas Hayano et al⁶⁷ found that low BF of tumors correlated with poorer outcomes and tumor progression. In terms of the role of DCE-CT as a predictive biomarker for chemoradiation, the data are controversial; Bellomi et al⁶⁸ suggested that low perfusion values at baseline were associated with a poorer response, whereas Sahani et al⁶⁹ reported the contrary.

Similarly, DCE-MRI is a functional imaging modality that is being investigated as an imaging biomarker. This is an imaging modality that may be useful as a predictive marker in patients receiving angiogenesis inhibitors.⁷⁰ It evaluates the extravasation of paramagnetic contrast agents, follows their uptake, and changes in signal intensity over time. Preliminary evidence suggests that contrast-enhanced imaging may be a useful tool to predict patient treatment response to angiogenesis inhibitors.

Although none of these imaging technologies have entered routine clinical practice yet, they highlight important scientific concepts. It is conceivable that one or more imaging markers will enter clinical practice in the not too distant future.

Conclusion

The poor outcome of mCRC, as well as the widespread use of antiangiogenic agents, has prompted the need for reliable measures to predict response to treatment. Given the complexity of the angiogenesis pathway, the discovery of biomarkers that predict angiogenesis inhibitor-related outcomes has been challenging.

Circulating and tissue biomarkers show promising potential as predictive biomarkers. Additionally, functional



imaging is emerging as a superior tool with to predict response to treatment with antiangiogenic agents. Although several studies have produced encouraging results, definitive data is lacking. Large-scale studies are needed to confirm that promising biomarkers are truly predictive rather than just prognostic in patients with mCRC. In the absence of biomarkers, the decision to treat patients with angiogenesis inhibitors remains a clinical decision based on the perceived balance between the benefit and toxicities of antiangiogenic agents.

Author Contributions

Conceived the concepts: SM and LM. Analyzed the data: SM and LM. Wrote the first draft of the manuscript: SM, MS and LM. Contributed to the writing of the manuscript: SM, MS and LM. Agree with manuscript results and conclusions: SM, LM, MS and MH. Jointly developed the structure and arguments for the paper: SM, LM. Made critical revisions and approved final version: SM, LM and MS. All authors reviewed and approved of the final manuscript. We thank Marion Hartley for her assistance in editing the manuscript.

REFERENCES

- Global Burden of Disease Cancer Collaboration; Fitzmaurice C, Dicker D, et al. The global burden of cancer 2013. *JAMA Oncol.* 2015;1(4):505–527.
- Schmoll HJ, Van Cutsem E, Stein A, et al. ESMO consensus guidelines for management of patients with colon and rectal cancer. A personalized approach to clinical decision making. *Ann Oncol.* 2012;23(10):2479–2516.
- Böckelman C, Engelmann BE, Kaprio T, Hansen TF, Glimelius B. Risk of recurrence in patients with colon cancer stage II and III: a systematic review and meta-analysis of recent literature. *Acta Oncol.* 2015;54(1):5–16.
- Heinemann V, von Weikersthal LF, Decker T, et al. FOLFIRI plus cetuximab versus FOLFIRI plus bevacizumab as first-line treatment for patients with metastatic colorectal cancer (FIRE-3): a randomised, open-label, phase 3 trial. *Lancet Oncol.* 2014;15(10):1065–1075.
- Simmonds PC, Primrose JN, Colquitt JL, Garden OJ, Poston GJ, Rees M. Surgical resection of hepatic metastases from colorectal cancer: a systematic review of published studies. *Br J Cancer.* 2006;94(7):982–999.
- Brenner H, Bouvier AM, Foschi R, et al; EURO-CARE Working Group. Progress in colorectal cancer survival in Europe from the late 1980s to the early 21st century: the EURO-CARE study. *Int J Cancer.* 2012;131(7):1649–1658.
- Peeters M, Price T, Van Laethem JL. Anti-epidermal growth factor receptor monotherapy in the treatment of metastatic colorectal cancer: where are we today? *Oncologist.* 2009;14(1):29–39.
- Van Cutsem E, Tabernero J, Lakomy R, et al. Addition of aflibercept to fluorouracil, leucovorin, and irinotecan improves survival in a phase III randomized trial in patients with metastatic colorectal cancer previously treated with an oxaliplatin-based regimen. *J Clin Oncol.* 2012;30(28):3499–3506.
- Hurwitz H, Fehrenbacher L, Novotny W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N Engl J Med.* 2004;350(23):2335–2342.
- Grothey A, Van Cutsem E, Sobrero A, et al; CORRECT Study Group. Regorafenib monotherapy for previously treated metastatic colorectal cancer (CORRECT): an international, multicentre, randomised, placebo-controlled, phase 3 trial. *Lancet.* 2013;381(9863):303.
- Garcia-Carbonero R, Rivera F, Maurel J, et al. An open-label phase II study evaluating the safety and efficacy of ramucirumab combined with mFOLFOX-6 as first-line therapy for metastatic colorectal cancer. *Oncologist.* 2014;19(4):350–351.
- Jain RK. Normalization of tumor vasculature: an emerging concept in antiangiogenic therapy. *Science.* 2005;307(5706):58–62.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell.* 2000;100(1):57–70.
- Kerbel RS. Tumor angiogenesis. *N Engl J Med.* 2008;358(19):2039–2049.
- Soker S, Takashima S, Miao HQ, Neufeld G, Klagsbrun M. Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor. *Cell.* 1998;92(6):735.
- Ebos JM, Lee CR, Bogdanovic E, et al. Vascular endothelial growth factor-mediated decrease in plasma soluble vascular endothelial growth factor receptor-2 levels as a surrogate biomarker for tumor growth. *Cancer Res.* 2008;68(2):521–529.
- Cao Y. Positive and negative modulation of angiogenesis by VEGFR1 ligands. *Sci Signal.* 2009;2(59):re1.
- Bernatchez PN, Rollin S, Soker S, Sirois MG. Relative effects of VEGF-A and VEGF-C on endothelial cell proliferation, migration and PAF synthesis: role of neuropilin-1. *J Cell Biochem.* 2002;85(3):629–639.
- Gagnon ML, Bielenberg DR, Gechtman Z, et al. Identification of a natural soluble neuropilin-1 that binds vascular endothelial growth factor: in vivo expression and antitumor activity. *Proc Natl Acad Sci U S A.* 2000;97(6):2573–2578.
- Saban MR, Backer JM, Backer MV, et al. VEGF receptors and neuropilins are expressed in the urothelial and neuronal cells in normal mouse urinary bladder and are upregulated in inflammation. *Am J Physiol Renal Physiol.* 2008;295(1):F60–F72.
- Harper SJ, Bates DO. VEGF-A splicing: the key to anti-angiogenic therapeutics? *Nat Rev Cancer.* 2008;8(11):880–887.
- Tammela T, Zarkada G, Nurmi H, et al. VEGFR-3 controls tip to stalk conversion at vessel fusion sites by reinforcing Notch signalling. *Nat Cell Biol.* 2011;13(10):1202–1213.
- Benedito R, Rocha SF, Woeste M, et al. Notch-dependent VEGFR3 upregulation allows angiogenesis without VEGF-VEGFR2 signalling. *Nature.* 2012;484(7392):110.
- Brown LF, Berse B, Jackman RW, et al. Expression of vascular permeability factor (vascular endothelial growth factor) and its receptors in adenocarcinomas of the gastrointestinal tract. *Cancer Res.* 1993;53(19):4727–4735.
- Jantus-Lewintre E, Sanmartin E, Sirera R, et al. Combined VEGF-A and VEGFR-2 concentrations in plasma: diagnostic and prognostic implications in patients with advanced NSCLC. *Lung Cancer.* 2011;74(2):326–331.
- Takahashi T, Ueno H, Shibuya M. VEGF activates protein kinase C-dependent, but Ras-independent Raf-MEK-MAP kinase pathway for DNA synthesis in primary endothelial cells. *Oncogene.* 1999;18(13):2221.
- Meadows KN, Bryant P, Pumiglia K. Vascular endothelial growth factor induction of the angiogenic phenotype requires Ras activation. *J Biol Chem.* 2001;276(52):49289–49298.
- Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. *Nat Med.* 2003;9(6):669–676.
- Jubb AM, Miller KD, Rugo HS, et al. Impact of exploratory biomarkers on the treatment effect of bevacizumab in metastatic breast cancer. *Clin Cancer Res.* 2011;17(2):372–381.
- Pohl M, Werner N, Munding J, et al. Biomarkers of anti-angiogenic therapy in metastatic colorectal cancer (mCRC): original data and review of the literature. *Z Gastroenterol.* 2011;49(10):1398–1406.
- Chung S, Dwabe S, Elshimali Y, Sukhija H, Aroh C, Vadgama JV. Identification of novel biomarkers for metastatic colorectal cancer using angiogenesis-antibody array and intracellular signaling array. *PLoS One.* 2015;10(8):e0134948.
- Ciardiello F, Troiani T, Bianco R, et al. Interaction between the epidermal growth factor receptor (EGFR) and the vascular endothelial growth factor (VEGF) pathways: a rational approach for multi-target anticancer therapy. *Ann Oncol.* 2006;17(suppl 7):vii109.
- Jürgensmeier JM, Schmoll HJ, Robertson JD, et al. Prognostic and predictive value of VEGF, sVEGFR-2 and CEA in mCRC studies comparing cediranib, bevacizumab and chemotherapy. *Br J Cancer.* 2013;108(6):1316.
- Kara O, Duman BB, Kara B, Erdogan S, Parsak CK, Sakman G. Analysis of PTEN, VEGF, HER2 and P53 status in determining colorectal cancer benefit from bevacizumab therapy. *Asian Pac J Cancer Prev.* 2012;13(12):6397.
- Tsai HL, Lin CH, Huang CW, et al. Decreased peritumoral VEGF expression could be a predictor of responsiveness to first-line FOLFIRI plus bevacizumab in mCRC patients. *Int J Clin Exp Pathol.* 2015;8(2):1900–1910.
- Jubb AM, Hurwitz HI, Bai W, et al. Impact of vascular endothelial growth factor-A expression, thrombospondin-2 expression, and microvessel density on the treatment effect of bevacizumab in metastatic colorectal cancer. *J Clin Oncol.* 2006;24(2):217–227.
- Eng L, Azad AK, Habbous S, et al. Vascular endothelial growth factor pathway polymorphisms as prognostic and pharmacogenetic factors in cancer: a systematic review and meta-analysis. *Clin Cancer Res.* 2012;18(17):4526–4537.
- Formica V, Palmirotta R, Del Monte G, et al. Predictive value of VEGF gene polymorphisms for metastatic colorectal cancer patients receiving first-line treatment including fluorouracil, irinotecan, and bevacizumab. *Int J Colorectal Dis.* 2011;26(2):143–151.
- Garger A, LaBonte M, Lenz HJ. Molecular predictors of response to antiangiogenesis therapies. *Cancer J.* 2011;17(2):134–141.
- Zhang SD, McCrudden CM, Meng C, Lin Y, Kwok HF. The significance of combining VEGFA, FLT1, and KDR expressions in colon cancer patient prognosis and predicting response to bevacizumab. *Oncotargets Ther.* 2015;8:835–843.
- Carrillo-de Santa Pau E, Carrillo Arias F, Caso Pelaez E, et al. Vascular endothelial growth factor (VEGF) serum levels are associated with survival in early stages of lung cancer patients. *Cancer Invest.* 2010;28(4):393–398.

42. Duda DG, Willett CG, Ancukiewicz M, et al. Plasma soluble VEGFR-1 is a potential dual biomarker of response and toxicity for bevacizumab with chemotherapy in locally advanced rectal cancer. *Oncologist*. 2010;15(6):577–583.
43. Hoff PM, Hochhaus A, Pestalozzi BC, et al. Cediranib plus FOLFOX/CAPOX versus placebo plus FOLFOX/CAPOX in patients with previously untreated metastatic colorectal cancer: a randomized, double-blind, phase III study (HORIZON II). *J Clin Oncol*. 2012;30(29):3596–3603.
44. Bates DO, Catalano PJ, Symonds KE, et al. Association between VEGF splice isoforms and progression-free survival in metastatic colorectal cancer patients treated with bevacizumab. *Clin Cancer Res*. 2012;18(22):6384–6391.
45. Lim SH, Becker TM, Chua W, et al. Circulating tumour cells and circulating free nucleic acid as prognostic and predictive biomarkers in colorectal cancer. *Cancer Lett*. 2014;346(1):24–33.
46. Diehl F, Li M, Dressman D, et al. Detection and quantification of mutations in the plasma of patients with colorectal tumors. *Proc Natl Acad Sci U S A*. 2005;102(45):16368–16373.
47. Cohen SJ, Punt CJ, Iannotti N, et al. Relationship of circulating tumor cells to tumor response, progression-free survival, and overall survival in patients with metastatic colorectal cancer. *J Clin Oncol*. 2008;26(19):3213–3221.
48. Rahbari NN, Reissfelder C, Mühlbauer M, et al. Correlation of circulating angiogenic factors with circulating tumor cells and disease recurrence in patients undergoing curative resection for colorectal liver metastases. *Ann Surg Oncol*. 2011;18(8):2182–2191.
49. Groot Koerkamp B, Rahbari NN, Büchler MW, Koch M, Weitz J. Circulating tumor cells and prognosis of patients with resectable colorectal liver metastases or widespread metastatic colorectal cancer: a meta-analysis. *Ann Surg Oncol*. 2013;20(7):2156–2165.
50. Stiegelbauer V, Perakis S, Deutsch A, Ling H, Gerger A, Pichler M. MicroRNAs as novel predictive biomarkers and therapeutic targets in colorectal cancer. *World J Gastroenterol*. 2014;20(33):11727–11735.
51. Shivapurkar N, Mikhail S, Navarro R, et al. Decrease in blood miR-296 predicts chemotherapy resistance and poor clinical outcome in patients receiving systemic chemotherapy for metastatic colon cancer. *Int J Colorectal Dis*. 2013;28(6):887.
52. Hansen TF, Christensen RD, Andersen RF, Sørensen FB, Johnsson A, Jakobsen A. MicroRNA-126 and epidermal growth factor-like domain 7-an angiogenic couple of importance in metastatic colorectal cancer. Results from the Nordic ACT trial. *Br J Cancer*. 2013;109(5):1243–1251.
53. Kopetz S, Hoff PM, Morris JS, et al. Phase II trial of infusional fluorouracil, irinotecan, and bevacizumab for metastatic colorectal cancer: efficacy and circulating angiogenic biomarkers associated with therapeutic resistance. *J Clin Oncol*. 2010;28(3):453–459.
54. Goede V, Coutelle O, Neuneier J, et al. Identification of serum angiopoietin-2 as a biomarker for clinical outcome of colorectal cancer patients treated with bevacizumab-containing therapy. *Br J Cancer*. 2010;103(9):1407–1414.
55. Pohl A, El-Khoueiry A, Yang D, et al. Pharmacogenetic profiling of CD133 is associated with response rate (RR) and progression-free survival (PFS) in patients with metastatic colorectal cancer (mCRC), treated with bevacizumab-based chemotherapy. *Pharmacogenomics J*. 2013;13(2):173–180.
56. Tabernero J, Lenz HJ, Siena S, et al. Analysis of circulating DNA and protein biomarkers to predict the clinical activity of regorafenib and assess prognosis in patients with metastatic colorectal cancer: a retrospective, exploratory analysis of the CORRECT trial. *Lancet Oncol*. 2015;16(8):937–948.
57. Willett CG, Boucher Y, Duda DG, et al. Surrogate markers for antiangiogenic therapy and dose-limiting toxicities for bevacizumab with radiation and chemotherapy: continued experience of a phase I trial in rectal cancer patients. *J Clin Oncol*. 2005;23(31):8136–8139.
58. Willett CG, Duda DG, di Tomaso E, et al. Efficacy, safety, and biomarkers of neoadjuvant bevacizumab, radiation therapy, and fluorouracil in rectal cancer: a multidisciplinary phase II study. *J Clin Oncol*. 2009;27(18):3020–3026.
59. Torigan DA, Huang SS, Houseni M, Alavi A. Functional imaging of cancer with emphasis on molecular techniques. *CA Cancer J Clin*. 2007;57(4):206–224.
60. Beets-Tan RG, Beets GL, Vliegen RF, et al. Accuracy of magnetic resonance imaging in prediction of tumour-free resection margin in rectal cancer surgery. *Lancet*. 2001;357(9255):497–504.
61. Nickel MC, Bipat S, Stoker J. Diagnostic imaging of colorectal liver metastases with CT, MR imaging, FDG PET, and/or FDG PET/CT: a meta-analysis of prospective studies including patients who have not previously undergone treatment. *Radiology*. 2010;257(3):674–684.
62. Kalf J, Duong C, Drummond EG, Matthews JP, Hicks RJ. Findings on 18F-FDG PET scans after neoadjuvant chemoradiation provides prognostic stratification in patients with locally advanced rectal carcinoma subsequently treated by radical surgery. *J Nucl Med*. 2006;47(1):14–22.
63. Capirci C, Rampin L, Erba PA, et al. Sequential FDG-PET/CT reliably predicts response of locally advanced rectal cancer to neo-adjuvant chemo-radiation therapy. *Eur J Nucl Med Mol Imaging*. 2007;34(10):1583–1593.
64. Ono K, Ochiai R, Yoshida T, et al. Comparison of diffusion-weighted MRI and 2-[fluorine-18]-fluoro-2-deoxy-D-glucose positron emission tomography (FDG-PET) for detecting primary colorectal cancer and regional lymph node metastases. *J Magn Reson Imaging*. 2009;29(2):336–340.
65. Hein PA, Kremser C, Judmaier W, et al. Diffusion-weighted magnetic resonance imaging for monitoring diffusion changes in rectal carcinoma during combined, preoperative chemoradiation: preliminary results of a prospective study. *Eur J Radiol*. 2003;45(3):214–222.
66. Goh V, Halligan S, Wellsted DM, Bartram CI. Can perfusion CT assessment of primary colorectal adenocarcinoma blood flow at staging predict for subsequent metastatic disease? A pilot study. *Eur Radiol*. 2009;19(1):79–89.
67. Hayano K, Shuto K, Koda K, Yanagawa N, Okazumi S, Matsubara H. Quantitative measurement of blood flow using perfusion CT for assessing clinicopathologic features and prognosis in patients with rectal cancer. *Dis Colon Rectum*. 2009;52(9):1624–1629.
68. Bellomi M, Petralia G, Sonzogni A, Zampino MG, Rocca A. CT perfusion for the monitoring of neoadjuvant chemotherapy and radiation therapy in rectal carcinoma: initial experience. *Radiology*. 2007;244(2):486–493.
69. Sahani DV, Kalva SP, Hamberg LM, et al. Assessing tumor perfusion and treatment response in rectal cancer with multisession CT: initial observations. *Radiology*. 2005;234(3):785–792.
70. Hong HS, Kim SH, Park HJ, et al. Correlations of dynamic contrast-enhanced magnetic resonance imaging with morphologic, angiogenic, and molecular prognostic factors in rectal cancer. *Yonsei Med J*. 2013;54(1):123–130.
71. Matsusaka S, Suenaga M, Mishima Y, et al. Circulating tumor cells as a surrogate marker for determining response to chemotherapy in Japanese patients with metastatic colorectal cancer. *Cancer Sci*. 2011;102(6):1188–1192.
72. Sastre J, Maestro ML, Gómez-España A, et al. Circulating tumor cell count is a prognostic factor in metastatic colorectal cancer patients receiving first-line chemotherapy plus bevacizumab: a Spanish Cooperative Group for the Treatment of Digestive Tumors study. *Oncologist*. 2012;17(7):947–955.
73. Gazzaniga P, Gradilone A, Petracca A, et al. Molecular markers in circulating tumour cells from metastatic colorectal cancer patients. *J Cell Mol Med*. 2010;14(8):2073–2077.
74. Lu CY, Tsai HL, Uen YH, et al. Circulating tumor cells as a surrogate marker for determining clinical outcome to mFOLFOX chemotherapy in patients with stage III colon cancer. *Br J Cancer*. 2013;108(4):791–797.
75. Akao Y, Khoo F, Kumazaki M, Shinohara H, Miki K, Yamada N. Extracellular disposal of tumor-suppressor miRs-145 and -34a via microvesicles and 5-FU resistance of human colon cancer cells. *Int J Mol Sci*. 2014;15(1):1392–1401.
76. Pekow J, Meckel K, Dougherty U, et al. Tumor suppressors miR-143 and miR-145 and predicted target proteins API5, ERK5, K-RAS, and IRS-1 are differentially expressed in proximal and distal colon. *Am J Physiol Gastrointest Liver Physiol*. 2015;308(3):G179–G187.
77. Xu Q, Liu LZ, Qian X, et al. miR-145 directly targets p70S6K1 in cancer cells to inhibit tumor growth and angiogenesis. *Nucleic Acids Res*. 2012;40(2):761–774.
78. Yu Y, Nangia-Makker P, Farhana L, Rajendra GS, Levi E, Majumdar AP. miR-21 and miR-145 cooperation in regulation of colon cancer stem cells. *Mol Cancer*. 2015;14:98.
79. Yamakuchi M, Yagi S, Ito T, Lowenstein CJ. MicroRNA-22 regulates hypoxia signaling in colon cancer cells. *PLoS One*. 2011;6(5):e20291.
80. Yamakuchi M, Lotterman CD, Bao C, et al. P53-induced microRNA-107 inhibits HIF-1 and tumor angiogenesis. *Proc Natl Acad Sci U S A*. 2010;107(14):6334–6339.
81. Geng L, Chaudhuri A, Talmon G, et al. MicroRNA-192 suppresses liver metastasis of colon cancer. *Oncogene*. 2014;33(46):5332–5340.
82. Sundaram P, Hultine S, Smith LM, et al. p53-responsive miR-194 inhibits thrombospondin-1 and promotes angiogenesis in colon cancers. *Cancer Res*. 2011;71(24):7490–7501.
83. Amodeo V, Bazan V, Fanale D, et al. Effects of anti-miR-182 on TSP-1 expression in human colon cancer cells: there is a sense in antisense? *Expert Opin Ther Targets*. 2013;17(11):1249–1261.
84. Kim SY, Lee YH, Bae YS. MiR-186, miR-216b, miR-337-3p, and miR-760 cooperatively induce cellular senescence by targeting alpha subunit of protein kinase CKII in human colorectal cancer cells. *Biochem Biophys Res Commun*. 2012;429(3–4):173–179.
85. Saxena A, Shoeb M, Ramana KV, Srivastava SK. Aldose reductase inhibition suppresses colon cancer cell viability by modulating microRNA-21 mediated programmed cell death 4 (PDCD4) expression. *Eur J Cancer*. 2013;49(15):3311–3319.